

Revisión

Anti-inflammatory properties of dietary flavonoids

J. González-Gallego, S. Sánchez-Campos y M. J. Tuñón

Ciberehd and Institute of Biomedicine. University of Leon. Spain.

Abstract

Flavonoids are a group of natural substances that are located in sources of vegetal origin. More than 4,000 varieties of flavonoids have been identified. All of them are phenyl-benzopyrones of low molecular weight with a basic structure formed by two benzene rings united through a heterocyclic pyrane or pyrone. Besides their relevance in plants, flavonoids are important for human health. Their antioxidant capacity confers a therapeutic potential in cardiovascular diseases, gastric or duodenal ulcers, cancer or hepatic pathologies. Also important are their antiviral and anti-allergic actions, as well as their anti-thrombotic and anti-inflammatory properties. Prostaglandins and nitric oxide biosynthesis is involved in inflammation, and isoforms of inducible nitric oxide synthase (iNOS) and of cyclooxygenase (COX-2) are responsible for the production of a great amount of these mediators. It has been demonstrated that flavonoids are able to inhibit both enzymes, as well as other mediators of the inflammatory process such as reactive C protein or adhesion molecules. Modulation of the cascade of molecular events leading to the overexpression of those mediators include inhibition of transcription factors such as nuclear factor kappa B and AP-1, through the inhibition of protein kinases involved in signal transduction. Increased antioxidant defenses through activation of the NF-E2 related factor 2 (Nrf2) also contribute to the anti-inflammatory capacity of flavonoids.

(*Nutr Hosp.* 2007;22:287-93)

Key words: *Flavonoids. Inflammation. Oxidative stress. Nuclear factor kappa B. Nitric oxide.*

PROPIEDADES ANTIINFLAMATORIAS DE LOS FLAVONOIDEOS DE LA DIETA

Resumen

Los flavonoides son un grupo de las sustancias naturales que se encuentran en fuentes de origen vegetal, existiendo más de 4.000 variedades. Todos son fenil-benzopironas de peso molecular bajo con una estructura básica formada por dos anillos heterocíclicos de benceno unidos a través de un pirano o de una pirona. Además de su función en las plantas, los flavonoides son importantes para la salud humana. Su capacidad antioxidante confiere un potencial terapéutico en enfermedades cardiovasculares, úlceras gástricas o duodenales, cáncer o patologías hepáticas. También son importantes sus acciones antivirales y antialérgicas, así como sus características antitrombóticas y antiinflamatorias. La síntesis de prostaglandinas y de óxido nítrico está implicada en la inflamación, e isoformas de la óxido nítrico sintetasa (iNOS) y de la ciclooxigenasa (COX-2) son responsables de la producción de una gran cantidad de estos mediadores. Se ha demostrado que los flavonoides pueden inhibir ambas enzimas, así como otros mediadores del proceso inflamatorio tales como la proteína C reactiva o diversas moléculas de adhesión. La modulación de la cascada de los acontecimientos moleculares que conducen al aumento en la expresión de estos mediadores incluye la inhibición de factores de transcripción tales como el factor nuclear kappaB y el factor AP-1, a través de la inhibición de diferentes proteína quinasas. Otros factores, tales como el incremento de las defensas antioxidantes a través del factor Nrf2 pueden también contribuir a las propiedades antiinflamatorias de los flavonoides.

(*Nutr Hosp.* 2007;22:287-93)

Palabras clave: *Flavonoides. Inflamación. Estrés oxidativo. Factor nuclear kappa B. Óxido nítrico.*

Correspondence: Javier González-Gallego.

Departamento de Ciencias Biomédicas.

Universidad de León.

24071 León (España).

E-mail: jgonga@unileon.es

Recibido: 10-XII-2006.

Aceptado: 1-II-2007.

General characteristics of flavonoids

Flavonoids belong to a group of natural substances containing in their chemical structure a variable number of hydroxyl phenolic groups. They are located in sources of vegetal origin (fruits, seeds, roots, flowers, tea or wine). Their name derives from the Latin *flavus* (yellow) and many of these compounds are responsible for the coloration of flowers, yolks or leaves in autumn. Its function in plants seems to be of attracting pollinators or fruit-eating animals towards with the intention that they can disperse better the seeds. It is the case of the bromeliaceous, which develop their flowers on a species of stem formed by bracts that display an intense reddish colour before or during pollination, changing later to more greenish. In other cases flavonoids contribute to attract the prey. For example, carnivorous plants like the *Drosera*, use anthocyanins in their flowers to attract the insects that soon will devour. Sometimes, flavonoids constitute the adaptive answer of plants to the intense ultraviolet radiation. Effects such as defence against the infections, control of the breathing, participation in the photosynthesis or activation of implied bacterial genes in the fixation of nitrogen have been also described.^{1,2}

More than 4,000 varieties of flavonoids have been identified. All of them are phenyl-benzopyrones of low molecular weight with a basic structure formed by two benzene rings united through a heterocyclic pyrane or pyrone. They present differences in their chemical structure that can facilitate the interaction with certain receptor molecules and/or their respective pathways within the cells, such as apoptosis, cell activation to stress and cascades of protein signalling kinases.³ Based on his molecular structure they are divided in four fundamental groups: flavones, flavanones, flavanoles and anthocyanins (fig. 1). Flavones are characterized by the presence of a double bond in the central aromatic ring. Quercetin, one of the better known flavonoids, is a member of this group along with kaempferol, and it is in abundance in apples, onions, Brussels sprouts or fruits of citrus. Flavones, such as naringenin or hesperedin, are located fundamentally in citrus. Between flavanoles, catechins are mainly found in green and black tea or in red wine. Finally, anthocyanins appear in strawberries, grapes, wine or tea.⁴

Besides their relevance in plants, flavonoids are important for human health. Many of these compounds are found in the human diet, but unanimity of criteria concerning the average dietary intake do not exist, having been reported values of approximately 250 mg/day in Dutch population,⁵ 128 mg/day in Australian women⁶ or up to 1 g/day in the USA.⁷ An exact estimation is difficult, because it depends on very diverse factors, such as the compound of reference used for the analysis, or the design and methodology of the study. In any case, important differences probably exist between diverse countries, and the Mediterranean

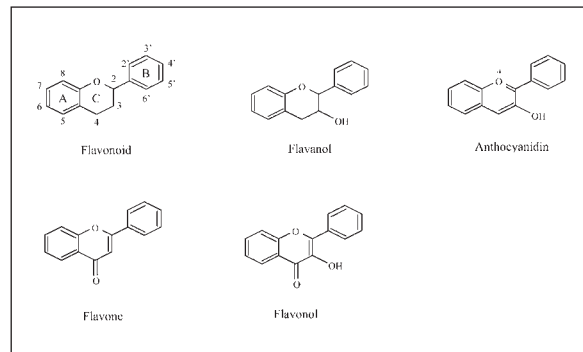


Fig. 1.—Flavonoids. Basic structure and groups.

diet, rich in fruit, olive oil of citrus or vegetables, may suppose an intake considerably greater than that in other surroundings.³ Large differences also exist concerning content in foods, which depends, among others, on genetic factors, such as the vegetal species, light environmental conditions or the processing after the harvest.⁸ Having trustworthy data on this content can have enormous importance to carry out studies on pharmacodynamic effects of flavonoids, and for a better knowledge of optimal consumption levels. Recently, the Department of Agriculture of the United States (USDA) has published the updated version of database from 2003, in which analytical values for 26 selected flavonoids in 393 foods are included.⁹ The Laboratory of Data of Nutrients of USDA has released separated databases for the content in foods of isoflavones and proanthocyanidins that are equally available in the USDA Web page. Given the different biological actions derived from their chemical structure, some studies recommend the diversified consumption of flavonoids from diverse nutritional sources and, at the present time, the possibility of adding specific flavonoids to food which do not naturally contain them is being explored.¹⁰

Flavonoids seem to play an important role in human health and to possess beneficial effects in the prevention of human diseases. A fundamental property of these molecules, responsible for many of their beneficial effects, is the antioxidant capacity, linked to the presence of a series of structural characteristics that allow them, among others, to chelate ions of transition metals such as Fe^{2+} , Cu^{2+} or Zn^{2+} , to catalyze the electron transport, to scavenge reactive oxygen species (ROS) like the superoxide anion, oxygen singlet and lipidic peroxyradicals, or to stabilize free ROS by means of the hydrogenation or formation of complexes with oxidating species.¹¹ The antioxidant capacity of flavonoids confers a therapeutic potential in diseases between which cardiovascular diseases¹², gastric or duodenal ulcers,¹³ cancer¹⁴ or hepatic pathologies are included.¹⁵ Also important are their antiviral and anti-allergic actions, as well as their anti-thrombotic and anti-inflammatory properties.⁴ This last one constitute

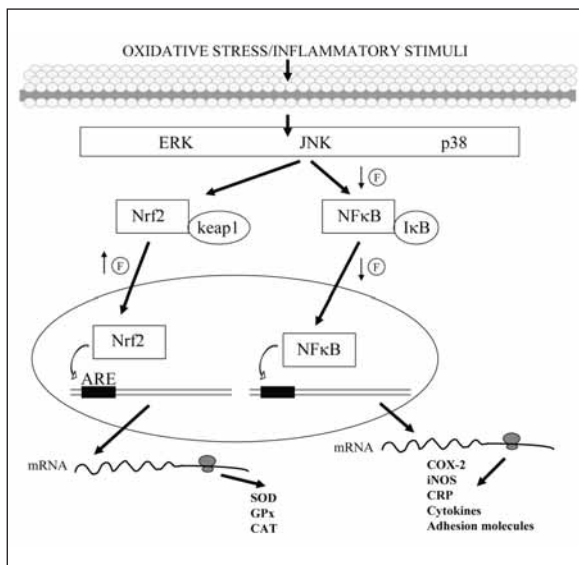


Fig. 2.—Schematic model for some anti-inflammatory effects of flavonoids.

an aspect used for a long time in the Chinese traditional medicine and the cosmetic industry under the form of plant extracts, but recently it has begun to be explored in depth, in order to identify the mechanisms responsible and the possibility for use of flavonoids as anti-inflammatory agents.

Flavonoids and inflammation

Inhibition of inflammation-related enzymes

Effects of flavonoids on a variety of inflammatory processes have been object of diverse reviews¹⁶ and it has been demonstrated that they are able to inhibit a series of enzymes which are activated in the course of the inflammatory process.¹⁷ Prostaglandins and nitric oxide biosynthesis is involved in inflammation, and isoforms of inducible nitric oxide synthase (iNOS) and of cyclooxygenase (COX-2) are responsible for the production of a great amount of these mediators. *In vitro* studies have confirmed that the flavonoid quercetin inhibits nitric oxide production and the expression of iNOS.¹⁸ Differences between various flavonoids exist and, for example, quercetin and kaempferol show little differences in their inhibiting capacity of the expression of iNOS in RAW264.7 cells,¹⁹ but the second inhibits to a greater extent than quercetin nitrite accumulation in culture medium of lipopolysaccharide (LPS)-stimulated J774.2 cells.²⁰ Although the inhibition of iNOS can contribute to the anti-inflammatory effect of flavonoids, the mechanism responsible for such effect is not known in depth. Thus, while in lung adenocarcinoma cell lines quercetin downregulates iNOS at translational level, inhibiting nitric oxide production and protein level,²¹ transcriptional effects have been described in interleukin 1β-

activated hepatocytes.¹⁸ Conflicting results have also been reported concerning regulation of COX-2 expression. Thus, quercetin downregulates COX-2 expression in macrophages.²¹ However, it increases COX-2 expression in human-derived colon cancer cells²³ or does not modify expression in human lung carcinoma cells.¹⁹ Moreover, a unique dose of quercetin is able to reduce the expression of COX-2 in human lymphocytes *in vivo* but not *ex vivo*.²⁴ Our group has recently demonstrated that both quercetin and kaempferol reduce iNOS and COX-2 protein levels in hepatic cells of the Chang Liver line,²⁵ agreeing with similar descriptions of the effect of quercetin in RAW 264.7 macrophages²⁶ or kaempferol in mouse macrophages.²²

Effects on transcription factors activation

There are several critical steps at which flavonoids can modulate the cascade of molecular events leading to the overexpression of iNOS or COX-2. These include inhibition of protein kinase C, phospholipase C or A2 and phosphodiesterases,³ indirect modulation of iNOS by inhibition of the cyclooxygenase and/or lipoxygenase pathways,²⁷ inhibition of transcription factors AP-1 and C/EBPδ,^{28,29} or effects on the interferon regulatory factor (IRF)-1 and Akt signaling pathway.³⁰ In any case, pathways of induction of iNOS and COX-2 seem to converge in the activation of a transcription essential for the expression of proinflammatory genes, the nuclear factor kappa B (NF-kappaB).³¹ The NF-kappaB is one of the main inducible transcription factors whose modulation triggers a cascade of molecular events, some of which can constitute potential key targets for the treatment of the inflammation. In alterations coursing with inflammation it exists an activation of cells, such as macrophages, which release cytokines (i.e. tumour necrosis factor alpha, TNFα), as well as ROS. The ROS can contribute to the appearance of oxidative stress, mainly in those cases in which an imbalance with enzymatic and nonenzymatic (glutathione, vitamins, and probably flavonoids) antioxidant defenses exist.^{32,33} In situations that course with oxidative stress, this can be an important stimulus for the activation of NF-kappaB, which appears in latent form in the cytoplasm of nonstimulated cells, forming a complex with its inhibitors, the I-kappaBs (IkappaBα and IkappaBβ).³⁴ When the cell is stimulated, NF-kappaB factor activates by means of the phosphorylation and degradation of the I-kappaB proteins and migrates to the nucleus, stimulating the expression of its target genes.³⁵ The phosphorylation of IkappaB involves two IkappaB kinases, IKKα and IKKβ. The degradation of IkappaBα results in rapid changes in the induction of NF-kappaB, whereas the degradation of IkappaBβ is associated with a prolonged activation of NF-kappaB.³⁶ Once activated, NF-kappaB can stimulate the expression of iNOS, with an increase in the nitric oxide formation.³⁷ The reaction of this one with ROS,

such as the superoxide anion, produces the formation of peroxynitrite, which additionally contributes to cellular injury.³⁸ We have demonstrated in an experimental model of ischemia-reperfusion how antioxidant substances such as glycine can block in parallel oxidative stress, the activation of NF-kappaB and nitric oxide production.³⁸ Studies on models of inflammation in isolated hepatocytes also confirm that the protective effects of the immunosuppressant agents FK506 and rapamycin are also related to the inhibition of the commented factors.³⁹

Different studies have shown that flavonoids can modulate the NF-kappaB signalling pathway during inflammation and modify through this the expression of genes involved in the inflammatory process (fig. 2).^{40,41} Reports concerning the relationship between flavonoids and the NF-kappaB pathway are, however, conflicting. Thus, it has been reported that quercetin does not reduce NF-kappaB activation in the renal cortex of rats with established chronic glomerular disease,⁴² and that while diverse flavonoids down-regulate iNOS expression in RAW264.7 cells, they do not suppress DNA binding activities of NF-kappaB.¹⁹ It also appears that only some flavonoids such as kaempferol suppress diverse tyrosine kinase-mediated signalling pathways in prostate cancer cells⁴³ or inhibit TNF α production and TNF α -induced NF-kappaB translocation in osteoblasts.⁴⁴ On the contrary, both quercetin and kaempferol diminish the degradation of IkappaB through the regulation of members of the IKK complex.²⁵ It has also been demonstrated that quercetin prevents LPS-induced IkappaB phosphorylation in bone marrow macrophages,⁴¹ inhibits the activation of NF-kappaB induced by interleukin 1 β in murine fibroblasts⁴⁵ and reduces IkappaB α and IkappaB β phosphorylation in peripheral blood mononuclear cells.¹⁶ Moreover, quercetin inhibits iNOS gene transcription induced by LPS in mouse BV-2 microglia through an effect mediated by attenuation of IkappaB phosphorylation⁴⁶ or abrogates in parallel iNOS overexpression and the activation of NF-kappaB in rat hepatocytes activated by interleukin 1 β .¹⁸ Curcumin reduces hepatic inflammation in mice with experimental steatohepatitis in parallel to an inhibition of NF-kappaB activation and the expression of proinflammatory genes,⁴⁷ and recently a relation between changes in NF-kappaB activation induced by cocoa flavonoids and the expression of COX-2 has been shown in mouse skin cells.⁴⁸ It has been also reported that targeting of NF-kappaB can contribute to the inhibition COX-2 expression by anthocyanidins in RAW 264 cells²⁹ or by amenthoflavone in A549 cells.⁴⁹

Reactive C protein (CRP) is an acute phase reactant whose elevation in serum is considered as indicator of chronic inflammation, and whose interaction with endothelial cells may be the mechanistic link with atherosclerosis.⁵⁰ It has been documented that CRP concentration has a predictive value in cardiovascular

disease, induces adhesion molecules expression⁵¹ and presents another series of proatherogenic effects in endothelial cells.⁵² Although it had been indicated that plasma concentration of CRP is not related to flavonoid intake,⁵³ recent data demonstrate that diverse flavonoides may reduce CRP protein level in hepatic cells and that the effect is dose-dependent (fig. 2).²⁵ It is known that IL-6 induces CRP by means of a mechanism that involves NF-kappaB activation,^{54,55} and that alterations in the intracellular state redox cause an inhibition of the nuclear NF-kappaB translocation and of CRP synthesis in hepatocytes.⁵⁶ Moreover, overexpressed NF-kappaB can participate in the induction of CRP in HepG2 cells by enhancing the effect of C/EBPbeta and the activator of transcription-3.⁵⁷ It is therefore probable that effects of flavonoids on CRP expression could be mediated, at least partly, by the modulation of the NF-kappaB-dependent pathway.

Effects on adhesion molecules

Endothelial dysfunction has an enormous importance in the inflammatory processes, and involves changes in the expression of endothelial adhesion molecules, such as ICAM-1, VCAM-1 and E-selectin, among others,^{58,59} able to trigger the inflammatory process by stimulating the migration and adhesion of leukocytes and originating tissue damage.⁶⁰ NF-kappaB activation is a necessary step in the transcriptional induction of adhesion molecules,⁶¹ as confirms the fact that blockers of NFkappaB also inhibit VCAM-1 expression in endothelial cell stimulated by the TNF α .⁶² In addition, activated endothelial cells release IL-6 that stimulate hepatocyte fibrinogen and CRP production,⁶³ which contributes to the exacerbation of endothelial dysfunction. Flavonoids, in addition to his capacity to inhibit *in vitro* lipoprotein oxidation and to their antithrombotic effects, seem to also exert their beneficial action in cardiovascular diseases by modulating monocyte adhesion during the atherosclerotic inflammatory.⁶⁴ Although all the mechanisms involved in this effect are not known with exactitude, it has been indicated that they could inhibit the expression of inflammatory mediators such as ICAM-1, by acting on NFkappaB activation.^{65,66} Nevertheless, it has also been shown that some flavonoids inhibit the expression of adhesion molecules in activated endothelial cells by process independent from NF-kappaB.⁶⁷

Modulation of proinflammatory gene expression

Effects of flavonoids on the binding capacity of transcription factors such as NF-kappaB or AP-1 may be regulated through the inhibition of protein kinases involved in signal transduction, such as protein kinase C (PKC) and mitogen activated protein kinase (MAPK) (fig. 2). In macrophages and other cell types, LPS activates three kinds of MAPs, extracellular signal related kinase (ERK), Jun N-terminal kinase/stress activated

protein kinase (JNK/SAPK) and p38 kinase.⁶⁸ It has been reported that quercetin inhibits iNOS expression through inhibition of p38 MAPK⁶⁹ and that blocks AP-1 binding in LPS-induced RAW cells by inhibiting JNK/SAPK.⁷⁰ Data from LPS-activated macrophages also show that quercetin is able to suppress proinflammatory cytokines and NF-kappaB activation through ERK and p38 MAPK.⁴⁰ In RAW 264.7 cells, luteolin also inhibits LPS-stimulated pathways through inhibition of some MAP kinases such as ERK and p38 MAPK.⁷¹

Modulation of redox state

An additional mechanism which could contribute to the anti-inflammatory properties of flavonoids is the modulation of the redox state by increases in the endogenous antioxidant defense potential (fig. 2). Organisms have developed a variety of antioxidant defense systems as protection from ROS. The major endogenous antioxidant systems include superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx) and glutathione reductase (GR). SOD catalyzes the dismutation of the superoxide radical anion, and there is a Mn SOD localized in the mitochondria and a Cu/Zn SOD mainly localized in the cytosol. CAT and GPx convert H₂O₂ to H₂O, and GR recycles oxidised glutathione back to reduced glutathione.⁷² The NF-E2 related factor 2 (Nrf2), a member of the cap'n'collar family of basic leucine zipper transcription factors, is a redox-sensitive factor whose nuclear translocation and binding to the antioxidant response elements (ARE) in their promoter regions may result in induction of antioxidant enzymes.⁷³ Thus, binding of Nrf2 to AREs of GPx2 in lungs⁷⁴ or Cu/Zn SOD⁷⁵ in liver have been reported, and Nrf2 is also involved in the regulation of catalase and other antioxidant enzymes in the fibroblasts.⁷⁶ Although changes in Nrf2 signaling in inflammatory diseases is not yet fully addressed,⁷⁷ it is known that polyphenols such as curcumin or epigallocatechin-3-gallate are Nrf2-ARE activators.^{78,79}

Limitations

In spite to the described beneficial effects, it is necessary to be cautious when analyzing potential beneficial effects of flavonoid administration, because these molecules may act as prooxidants at high doses. Thus, quercetin and other flavonoids such as myricetin increase hydroxyl radical production and DNA damage at concentration > 30 µM,⁸⁰ quercetin at concentrations exceeding 75 µM has some intrinsic cytotoxicity in cultured renal tubular cells,⁸¹ and quercetin at high dose may present genotoxic effects.⁸² Moreover, it has been recently reported that quercetin at concentrations > 50 µM is able to participate in the oxidation of NADPH in liver cells, shifting the cellular conditions to a more oxidized state.⁸³ This prooxidant character may explain why exposure of rat aortic smooth muscle cells to quercetin concentrations > 100

µM increases NFkappaB activation⁸⁴ or the fact that similar quercetin doses increase iNOS and COX-2 expression in parallel to the stimulation of the NF-kappa B-dependent pathway in liver cells.²⁵

An additional aspect to stand out is the limited bioavailability of flavonoids, due to its low absorption and rapid elimination. Aglycons and glucosides are absorbed in the small intestine, but they are transformed quickly into methylated, sulfated or glucuronic acid-conjugated derivative. Bacteria of the colon play an important role in the metabolism and absorption of flavonoids and the derivatives do not necessarily present the same biological activity than the original compounds.⁸⁵ Consequently, although the numerous studies published with *in vitro* approaches allow to identify molecular mechanisms of flavonoid effects, data generated must be validated in humans and it is necessary to be very careful when extrapolating results of *in vitro* experiments with purified compounds to *in vivo* situations. Whatever it is the case, the data nowadays available make clear the potential utility that have dietary flavonoids or new flavonoid-based agents for the possible treatment of inflammatory diseases.

References

1. McClure, JV: Physiology of flavonoids in plants. En: Cody, V, Middleton, E, Harborne, JB, Eds. *Plant Flavonoids in Biology and Medicine: Biochemical, Pharmacological and Structure-Activity Relationships*. New York: Alan R. Liss, Inc., 1987; 77-85.
2. Firmin JL, Wilson KE, Rossen L, Johnston AWB: Flavonoid activation of modulation genes in *Rhizobium* reversed by other compounds present in plants. *Nature* 1987; 324:90-92.
3. Middleton E, Kandaswami C, Theoharides TC: The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease and cancer. *Pharmacol Rev* 2000; 52:673-751.
4. Nijveldt RJ, Van Nood E, Van Hoorn EC, Boelens PG, Van Norren K, Van Leeuwen, PAM: Flavonoids: a review of probable mechanisms of action and potential applications. *Am J Clin Nutr* 2001; 74:418-425.
5. Hertog MGL, Hollman PCH, Katan MB, Kromhout D: Intake of potentially anticarcinogenic flavonoids and their determinants in adults in the Netherlands. *Nutr Cancer* 1993; 20:21-29.
6. Lyon-Wall P, Autenzio P, Lee E, Moss R, Gie S, Samman S: Catechins are the major sources of flavonoids in a group of Australian women. *Asia Pac J Clin Nutr* 2004; 13:S72.
7. Kuhnau J. The flavonoids: a class of semiessential food components. Their role in human nutrition. *World Rev Nutr Diet* 1976; 24:117-191.
8. Chu YH, Chang CL, Hsu HF: Flavonoid content of several vegetables and their antioxidant activity. *J Sci Food Agric* 2000; 80:561-566.
9. Nutrient Data Laboratory's Web site: *Database for Flavonoid Content of Selected Foods, Release 2*. <http://www.ars.usda.gov/nutrientdata>. Consulta: 30/11/2006.
10. Youdim KA, McDonald J, Kalt W, Joseph JA: Potential role of dietary flavonoids in reducing microvascular endothelium vulnerability to oxidative and inflammatory insults. *J Nutr Biochem* 2002; 13:282-288.
11. Martínez-Flores S, González-Gallego J, Culebras JM, Tuñón, MJ: Los flavonoides: propiedades y acciones antioxidantes. *Nutr Hosp* 2002; XVII:135-142.
12. Yao LH, Jiang YM, Shi J, Tomás-Barberán FA, Datta N, Singanusong R y cols.: Flavonoids in food and their health benefits. *Plant Food Human Nutr* 2004; 59:113-122.

13. Moreira A, Fraga C, Alonso M, Collado PS, Zettler C, Marro ni N, González-Gallego J: Quercetin prevents oxidative stress and NF-kappaB activation in gastric mucosa of portal hyper- tensive rats. *Biochem Pharmacol* 2004; 68:1939-1946.
14. Yang K, Lamprecht SA, Liu Y, Shinozaki H, Fan K, Leung D y cols.: Chemoprevention studies of the flavonoids quercetin and rutin in normal and azoxymethane-treated mouse colon. *Carcinogenesis* 2000; 1:1655-1660.
15. Peres W, Tuñón MJ, Mato S, Collado PS, González-Gallego J. Hepatoprotective effects of the flavonoid quercetin in rats with biliary obstruction. *J Hepatol* 2000; 33:742-750.
16. Nair MP, Mahajan S, Reynolds JL, Aalinkeel R, Nair H, Sch- wartz SA y cols.: The flavonoid quercetin inhibits proinflam- matory cytokine (Tumor necrosis factor alpha) gene expres- sion in normal peripheral mononuclear cells via modulation of the NF-kappaB system. *Clin Vac Immunol* 2002; 13:319-328.
17. Kwon KH, Murakami A, Tanaka T, Ohigashi H: Dietary rutin, but not its aglycon quercetin, ameliorates dextran sulfate so- dium-induced experimental colitis in mice: attenuation of pro- inflammatory gene expression. *Biochem Pharmacol* 2005; 69:395-406.
18. Martínez-Flores S, Gutiérrez-Fernández B, Sánchez-Campos S, González-Gallego J, Tuñón MJ: Quercetin prevents nitric oxide production and nuclear factor kappa B activation in interleukin- 1 β -activated rat hepatocytes. *J Nutr* 2005; 135: 1359-1365.
19. Kim BH, Cho SM, Reddy AM, Kim YS, Min KR, Kim Y: Down-regulatory effect of quercetrin gallate on nuclear factor- kappaB-dependent inducible nitric oxide synthase expression in lipopolysaccharide-stimulated macrophages RAW 264.7. *Biochem Pharmacol* 2005; 69:1577-1583.
20. Olszanecki R, Gebaska A, Kozlovski VI, Gryglewski RJ: Fla- vonoids and nitric oxide synthase. *J Physiol Pharmacol* 2002; 53:571-584.
21. Banerjee T, Van der Vliet A, Ziboh VA: Down regulation of COX-2 and iNOS by amentoflavone and quercetin in A549 human lung adenocarcinoma cell line. *Prostag Leukotr Essent Fatty Acids* 2002; 66:485-492.
22. Raso GM, Meli R, Di Carlo G, Pacilio M, Di Carlo R: Inhibi- tion of inducible nitric oxide synthase and cyclooxygenase-2 expression by flavonoids in macrophage J774A.1. *Life Sci* 2001; 68:921-931.
23. Al-Fayez M, Cai H, Tunstall R, Steward W, Gescher AJ: Dif- ferential modulation of cyclooxygenase-mediated prostaglan- din production by the putative cancer chemopreventive fla- vonoids tricrin, apigenin and quercetin. *Cancer Chemother Pharmacol* 2006; 58:816-825.
24. De Pascual Teresa S, Johnston KL, DuPont MS, O'Leary K, Ne- eds PW, Morgan LM y cols.: Williamson G. Quercetin metaboli- tes downregulate cyclooxygenase transcription in human lymphocytes *ex vivo* but not *in vivo*. *J Nutr* 2004; 134:552-557.
25. García-Mediavilla MV, Crespo I, Collado PS, Esteller A, Sán- chez-Campos S, Tuñón MJ y cols.: Anti-inflammatory effect of the flavones quercetin and kaempferol in Chang Liver cells involves inhibition of inducible nitric oxide synthase, cyclo- oxygenase-2 and reactive C-protein, and down-regulation of the nuclear factor kappaB pathway. *Eur J Pharmacol* 2007; 557:221-229.
26. Jung WJ, Sung MK: Effects of major dietary antioxidants on inflammatory markers of RAW 264.7 macrophages. *Biofac- tors* 2004; 21:131-137.
27. Robak J, Shridi F, Wolbis M, Krolikowska M: *Screening* of the influence of flavonoids on lipoxygenase and cyclooxyge- nase activity, as well as nonenzymic lipid oxidation. *Pol J Pharmacol Pharm* 1988; 40:451-458.
28. Chen CC, Chow MP, Huang WC, Lin YC, Chang YJ: Flavonoids inhibit tumour necrosis factor-alpha-induced up-regulation of in- tercellular adhesion molecule-1 (ICAM-1) in respiratory epithelial cells through activator protein-1 and nuclear factor-kB: struc- ture-activity relationships. *Mol Pharmacol* 2005; 66:683-693.
29. Hou DX, Yanagita T, Uto T, Masuzaki S, Fujii M: Anthocya- nidins inhibit cyclooxygenase-2 expression in LPS-evoked macrophages: structure-activity relationship and molecular mechanism involved. *Biochem Pharmacol* 2005; 70:417-425.
30. Ruiz PA, Haller D: Functional diversity of flavonoids in the inhibition of the proinflammatory NF-kappaB, IRF, and Akt signaling pathways in murine intestinal epithelial cell. *J Nutr* 2006; 136:664-671.
31. Jiang B, Xu S, Hou X, Pimentel DR, Brecher P, Cohen RA: Temporal control of NF-kappaB activation by ERK differen- tially regulates interleukin-1beta-induced gene expression. *J Biol Chem* 2004; 279:1323-1329.
32. Sánchez-Campos S, López-Acebo R, González P, Culebras JM, Tuñón MJ, González-Gallego J: Cholestasis and altera- tions of glutathione metabolism induced by FK506 in the rat. *Transplantation* 1998; 68:84-88.
33. Palomero J, Galán AI, Muñoz ME, Tuñón MJ, González-Galle- go J, Jiménez R: Effects of aging on the susceptibility to the toxic effects of cyclosporin in rats: changes in liver glutathione and antioxidant enzymes. *Free Radic Biol Med* 2001; 30:836-845.
34. Romics L, Kodys K, Dolganiuc A, Graham L, Velayudham A, Mandrekar P y cols.: Diverse regulation of NF-kappaB and peroxisome proliferators-activated receptors in murine non-al- coholic fatty liver. *Hepatology* 2004; 40:376-385.
35. Baeuerle PA, Baltimore D: NFkappaB: Ten years after. *Cell* 1996; 87:13-20.
36. Simões A, Porawski M, Alonso M, Collado PS, Marroni N, González-Gallego J: Quercetin prevents oxidative stress and NF-kappaB activation in liver of type 1 diabetic rats. *J Nutr* 2005; 135:299-304.
37. Szabo C, Billiar TR. Novel roles of nitric oxide in hemorrha- gic shock. *Shock* 199; 12:1-9.
38. Mauriz JL, Matilla B, Culebras JM, González P, González- Gallego J: Dietary glycine inhibits activation of nuclear factor kappa B and prevents liver injury in hemorrhagic shock in the rat. *Free Radic Biol Med* 2001; 15:1236-1244.
39. Tuñón MJ, Sánchez-Campos S, Gutiérrez B, Culebras JM, González-Gallego J: Effects of FK506 and rapamycin on ge- neration of reactive oxygen species, nitric oxide production and nuclear factor kappaB activation in rat hepatocytes. *Bio- chem Pharmacol* 2003; 66:439-445.
40. Cho SY, Park SJ, Kwon MJ, Jeong TS, Bok SH, Choi WY y cols.: Quercetin suppresses proinflammatory cytokines pro- duction through MAP kinases and NF-kappaB pathway in lip- polysaccharide-stimulated macrophage. *Mol Cell Biochem* 2003; 243:153-160.
41. Comalada M, Camuesco D, Sierra S, Ballester I, Xaus J, Gál- vez J y cols.: *In vivo* quercetin anti-inflammatory effect invol- ves releases of quercetin, which inhibits inflammation through down-regulation of NF-kappaB pathway. *Eur J Immunol* 2005; 35:584-592.
42. Rangan GK, Wang, Harris DC: Dietary quercetin augments activator protein-1 and does not reduce nuclear factor-kappaB in the renal cortex of rats with established chronic glomerular disease. *Nephron* 2002; 90:313-319.
43. Wang S, DeGroff VL, Clinton SK: Tomato and soy polyphen- ols reduce insulin-like growth factor-I-stimulated prostate cancer cell proliferation and apoptotic resistance *in vitro* via inhibition of intracellular signaling pathways involving tyrosi- ne kinase. *J Nutr* 2003; 133:2367-2376.
44. Pang JL, Ricupero DA, Huang S, Fatma N, Sing DP, Romero JS y cols.: Differential activity of kaempferol and quercetin in attenuating tumor necrosis factor receptor family signaling in bone cells. *Biochem Pharmacol* 2006; 71:818-886.
45. Muraoka K, Shimizu K, Sun X, Tani Y, Izumumi R, Miwa K y cols.: Flavonoids exert diverse inhibitory effects on the acti- vation of NF-kappaB. *Transplant Proc* 2002; 34:1335-1340.
46. Chem JC, Ho FM, Pei-Dawn LC, Chen CP, Jeng KC, Hsu HB, Lee ST, Went Tung W, Lin WW: Inhibition of iNOS by querce- tin is mediated by the inhibition of IkappaB kinase, nuclear fac- tor kappa B, and STAT-1, and depends on heme oxygenase-1 in- duction in mouse BV-2 microglia. *Eur J Pharmacol* 2005; 521:9-20.
47. Leclerq IA, Farell GC, Sempoux C, De la Peña A, Horsmans Y: Curcumin inhibits NF-kappaB activation and reduce the se- verity of experimental steatohepatitis in mice. *J Hepatol* 2004; 41:926-934.

48. Lee KW, Kundu JK, Kim SO, Chun KS, Surh YJ: Cocoa polyphenols inhibit phorbol ester-induced anion formation in cultured HL-60 cells and expression of cyclooxygenase-2 and activation of NF-kappaB and MPAKs in mouse skin *in vivo*. *J Nutr* 2006; 136:1150-1155.
49. Banerjee T, Valacchi G, Ziboh VA, Van der Vliet A: Inhibition of TNF α -induced cyclooxygenase-2 expression by amantoflavone through suppression of NF-kB activation in A549 cells. *Mol Cell Biochem* 2002; 283:105-110.
50. Liang YJ, Shyu KG, Wang BW, Lai LP: C-reactive protein activates the nuclear factor-kB pathway and induces vascular cell adhesion molecule-1 expression through CD32 in human umbilical vein endothelial cells and aortic endothelial cells. *J Molec Cell Cardiol* 2006; 40:412-420.
51. Pasceri V, Cheng JS, Willerson JT, Yeh ET: Modulation of C-reactive protein-mediated monocyte chemoattractant protein-1 induction in human endothelial cells by anti-atherosclerotic drugs. *Circulation* 2001; 103:2531-2534.
52. Venugopal SK, Devaraj S, Jialal I: Macrophage conditioned medium induces the expression of C-reactive protein in human aortic endothelial cells: potential for paracrine/autocrine effects. *Am J Pathol* 2005; 166:1265-1271.
53. Song Y, Manson JE, Buringm JE, Sesso HD, Liu S: Associations of dietary flavonoids with risk of type 2 diabetes, and markers of insulin resistance and systemic inflammation in women: a prospective study and cross-sectional analysis. *J Am Coll Nutr* 2005; 24:376-384.
54. Ahmad N, Chen LC, Gordon MA, Laskin JD, Laskin DL: 2002. Regulation of cyclooxygenase-2 by nitric oxide in activated hepatic macrophages during acute endotoxemia. *J Leuk Biol* 71, 1005-1011.
55. Odontuya G, Hoult JRS, Houghton PJ: Structure-activity relationship for anti-inflammatory effect of luteolin and its derived glycosides. *Phytother Res* 2005; 19:782-786.
56. Maehira F, Miyagi I, Eguchi Y: Selenium regulates transcription factor NF-kappaB activation during the acute phase reaction. *Clin Chim Acta* 2003; 334:163-171.
57. Agrawal A, Samols D, Kushner I: Transcription factor c-Rel enhances C-reactive protein expression by facilitating the binding of C/EBPbeta to the promoter. *Mol Immunol* 2003; 40:373-380.
58. Tomita N, Morishita R, Tomita S, Gibbons GH, Zhang L, Horiuchi M y cols.: Transcription factor decoy for NFkappaB inhibits TNF-alpha-induced cytokine and adhesion molecule expression *in vivo*. *Gene Ther* 2000; 7:1326-1332.
59. Cota-Gómez A, Flore NC, Cruz C, Casullo A, Aw TY, Ichikawa H, Schaack J y cols.: The human immunodeficiency virus-1 Tat protein activates human umbilical vein endothelial cell E-selectin expression via an NF-kappa B-dependent mechanism. *J Biol Che* 2002; 26:14390-14399.
60. Zhu GD, Arendsen DL, Gunawardana IW, Boyd SA, Stewart AO y cols.: Selective inhibition of ICAM-1 and E-selectin expression in human endothelial cells. 2. Aryl modifications of 4-(aryloxythieno(2,3-c)pyridines with fine-tuning at C-2 carbamides. *J Med Chem* 2001; 11:3469-87.
61. Collins T: Endothelial nuclear factor-kappaB and the initiation of the atherosclerotic lesion. *Lab Invest* 1993; 68:499-508.
62. Weber C, Erl W, Pietsch A, Srobel M, Ziegler-Heitbrock HWL, Weber PC: Antioxidants inhibit monocyte adhesion by suppressing nuclear factor-kappaB mobilization and induction of vascular cell adhesion molecule-1 in endothelial cells stimulated to generate radicals. *Arterioscler Thromb* 194; 14:1665-1673.
63. Mortensen RF: C-reactive protein, inflammation, and innate immunity. *Immunol Res* 2001; 24:163-76.
64. Meydany M: Nutrition interventions in aging and age-associated disease. *Ann N Y Acad Sci* 2001; 928:226-235.
65. Rimbach G, Saliou C, Panali R, Virgili F: Interaction between cultured endothelial cells and macrophages *in vitro* model for studying flavonoids in redox-dependent gene expression. *Methods Enzymol* 2001; 335:387-397.
66. Youdim KA, Martin A, Joseph JA: Incorporation of the elderberry anthocyanins by endothelial cells increases protection against oxidative stress. *Free Radic Biol Med* 2000; 29:51-60.
67. Wolle J, Hill RR, Ferguson E, Devall LJ, Trivedi BK, Newton RS y cols.: Selective inhibition of tumor necrosis factor-induced vascular cell adhesion molecule-1 gene expression by a novel flavonoid. *Arterios Throm Vasc Biol* 1996; 16:1501-1508.
68. Weinstein SL, Sanghera JC, Lemke K, DeFRanco AL, Pelech SL: Bacterial lipopolysaccharide induces tyrosine phosphorylation and activation of mitogen-activated protein kinases in macrophages. *J Biol Chem* 1992; 267:14955-14962.
69. Wadsworth TL, Koop DR: Effects of Ginkgo biloba extracts and quercetin on lipopolysaccharide-induced release of nitric oxide. *Chem Biol Interact* 2001; 137:43-58.
70. Wadsworth TL, McDonald TL, Koop D: Effects of Ginkgo biloba extract and quercetin on lipopolysaccharide-induced signaling pathways involved in the release of tumour necrosis factor alpha. *Biochem Pharmacol* 2001; 62:963-974.
71. Xagorari A, Roussos C, Papapetropoulos A: Inhibition of LPS-stimulated pathways in macrophages by the flavonoid luteolin. *Br J Pharmacol* 2002; 136:1058-1064.
72. Alia M, Ramos S, Mateos R, Granado-Serrano AB, Bravo L, Goya L: Quercetin protects human hepatoma HepG2 against oxidative stress induced by tert-butyl hydroperoxide. *Toxicol Appl Pharmacol* 2005; 21:110-118.
73. Brigelius-Flohe R, Banning A: Part of the series: from dietary antioxidants to regulators in cellular signalling and gene regulation. *Free Radic Biol Med* 2006; 40:775-787.
74. Singh A, Rangasamy T, Thimmulappa RK, Lee H, Osburn WG, Brigelius-Flohe R, Kensler TW, Yamamoto M, Biswl S: Glutathione peroxidase 2, the major cigarette smoke-inducible isoform of GPx in lungs, is regulated by Nrf2. *Am J Resp Cell Mol Biol* 2006; 35:639-650.
75. Park EY, Rho HM: The transcriptional activation of the human copper/zinc superoxide dismutase gene by 2,3,7,8-tetrachlorodibenzo-p-dioxin through two different regulator sites, the antioxidant responsive element and xenobiotic responsive element. *Mol Cell Biochem* 2002; 240:47-55.
76. Zhu H, Itoh K, Yamamoto M, Zweier JL, Li Y: Role of Nrf2 signaling in regulation of antioxidants and phase 2 enzymes in cardiac fibroblasts: protection against reactive oxygen and nitrogen species-induced cell injury. *FEBS Lett* 2005; 579:3029-3036.
77. Rahman I, Biswas SK, Kirkham PA: Regulation of inflammation and redox signalling by dietary polyphenols. *Biochem Pharmacol* 72: 1439-1452.
78. Banning A, Deubel S, Kluth D, Zhou Z, Brigelius-Flohe R: The GI-GPx gene is a target for Nrf2. *Mol Cell Biol* 2005; 25:4919-4923.
79. Andreadl CK, Howells LM, Atherfold PA, Manson MM: Involvement of Nrf2, p38, B-Raf, and nuclear factor-kappaB, but not phosphatidylinositol-3-kinase, in induction of hemoxygenase-1 by dietary polyphenols. *Mol Pharmacol* 2006; 69:1033-1040.
80. Laughton MJ, Halliwell B, Evans PJ, Hoult JRS: Antioxidant and pro-oxidant action of the plant phenolics quercetin, gossypol and myricetin. *Biochem Pharmacol* 1989; 38:2859-65.
81. Kuhlmann MK, Horsch E, Burkhardt G, Wagner M, Kohler H: Reduction of cisplatin toxicity in cultured renal tubular cells by the bioflavonoid quercetin. *Arch Toxicol* 1998; 72:536-40.
82. Da Silva J, Hermann SM, Heuser V, Peres W, Marroni N, González-Gallego J y cols.: Evaluation of the genotoxic effect of rutin and quercetin by comet assay and micronucleus test. *Food Chem Toxicol* 2002; 40:941-947.
83. Buss GD, Constantin J, De Lima LC, Teodoro GR, Comar JF, Ishii-Iwamoto EL y cols.: The action of quercetin on the mitochondrial NADH to NAD(+) ratio in the isolated perfused rat liver. *Planta Med* 2005; 71:1118-22.
84. Shih CM, Lin H, Liang YC, Lee WS, Bi WF, Juan SH: Concentration-dependent differential effects of quercetin on rat aortic smooth muscle cells. *Eur J Pharmacol* 2004; 496:41-48.
85. Williams RJ, Spencer JP, Rice-Evans C: Flavonoids: antioxidants or signaling molecules? *Free Radic Biol Med* 2004; 36:838-849.