PROSTATIC SPECIFIC ANTIGEN (PSA), PRO-INFLAMMATORY CYTOKINES, AND PROSTATIC PATHOLOGY (BENIGN PROSTATIC HYPERPLASIA AND CANCER). RELATIONSHIP WITH MALIGNANCY

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Summary.- OBJECTIVES: Cancer is a complex process in which cytokines play an important role. Cytokines are low-molecular weight soluble proteins involved in cellular transmission signals and several disorders. Pro-inflammatory cytokines (IL-1, TNF-α and IL-6) are involved in prostate cancer development. The aim of this study was to relate the expression (analyzed by Western blot and immuno-histochemistry) of several pro-inflammatory cytokines (IL-1, TNFα and IL-6) with serum levels of prostate-specific antigen (PSA) in normal (no pathological samples) as pathological samples (hyperplasia and cancer), in order to elucidate their possible role in tumor progression. We are also discussing the possible use of these cytokines as a potential therapeutic target.

METHODS: This study was carried out in 5 normal, 25 benign prostatic hyperplastic (BPH) and 17 prostate cancer (PC) human prostates. Immunohistochemical and Western blot analysis were performed. Serum levels of PSA were assayed by a PSA DPC immulite assays (Diagnostics Products Corporation, Los Angeles, CA)

RESULTS: In BPH, IL-1α, IL-6 and TNF-α were only expressed in patients with PSA serum levels of 0-4 ng/ml or 4-20 ng/ml, but not in the group >20 ng/ml. In PC these cytokines were only expressed in patients with PSA serum levels >4 ng/ml, although the expression of theses cytokines was elevated when PSA levels were >20 ng/ml.

CONCLUSIONS: In PC there might be an association between high expression of pro-inflammatory cytokines (IL-1, TNFα and IL-6), elevated serum levels of PSA and cancer progression. A better understanding of the biologic mechanism of this association may improve the finding of new targets for therapy in these patients.

Keywords: TNFα, IL-1, IL-6, PSA, Prostate cancer.
INTRODUCTION

IL-1 family includes two bioactive ligands (IL-1α and IL-1β) (1), two types of transmembrane receptors (IL-1RI and IL-1RII) (2) and a specific receptor antagonist (IL-1Ra). The affinity of IL-1α, IL-1β and IL-1Ra to receptors IL-1RI and IL-1RII differs depending of the ligand. IL-1α and IL-1Ra present higher affinity to IL-1RI than IL-1β, while IL-1β displays higher affinity to IL-1RII (3). The expression of IL-1 family has been reported in several tumors such as prostate cancer where Ricote et al. (4) propos that IL-1 family may be involved in cell proliferation since immunoexpression of IL-1α and IL-1Ra was a characteristic feature of PC compared with normal prostate samples and BPH.

TNF-α has been proposed as tumoral promoter (5). Different studies relate TNF-α endogenous production with tumour invasion and development of metastasis (6, 7). In cancer prostate have been related the serum levels of TNF-α to the development of the disease, associating the serum levels of TNF-α, the disease development and presence of metastasis (8). The functional duality of TNF-α in tumour tissues is determined by the intracellular signs that contributed your arrival to the cellular surface and binding your membrane receptors (TNFRI and TNFRII) and to activate different transduction pathway. The expression and action of TNF-α and its receptors has been reported in several tumours such as esophageal (9), prostate (10), follicular thyroid (11), skin (12), ovarian (13) and breast (14) cancers.

At the present, several researches are focusing in the search for new biochemical markers capable of predicting prostate cancer and prognosis. In addition, pro-inflammatory cytokines was related to the production of PSA and progression of prostate cancer (26-28).

PSA is a marker organ-specific, with high sensibility but low specificity for the detection of cancer prostate, existing situations that provoke a temporary increase of your levels not related to tumour processes such as manipulations of the prostate gland (rectal tact or biopsies) (23) and benign diseases (hyperplasia) (24). In addition, normal levels have been observed in patients with tumour pathology. In this way, PSA is not indicative of the evolution grade of disease (25).

In the present, different groups of research work on the search of new biochemical marker ca-
pable of predicting the presence and the prediction of cancer prostate. In this way, the pro-inflammatory cytokines have related to PSA production and the progression of cancer prostate (26-28).

PRESENTATION

The object of our study was to relate the expression of several pro-inflammatory cytokines (IL-1, IL-6 and TNF-α), analyzed by Western blot and immunohistochemistry, with serum levels of PSA in normal and pathologic (BPH and PC) prostatic tissue in order to elucidate their possible role in tumor progression. We are also discussing the possible use of these cytokines as a potential therapeutic factor.

MATERIAL AND METHODS

In this study we used: transurethral resections from 25 men (aged from 55 to 85 years) diagnosed clinically and histopathologically with benign prostate hyperplasia (BPH); radical prostatectomies from 17 men (aged from 57 to 88 years) diagnosed with prostate cancer (PC) with Gleason scores 3 to 5; and 5 men samples (aged from 20 to 38 years) obtained at autopsy (8-10 hours after death) without histories of, or reproductive, endocrine or related diseases. Each diagnosed sample was divided into three portions; one portion was immediately processed for immunohistochemistry, and the other two portions was frozen in liquid nitrogen and maintained at –80º C for Western Blotting analysis. All pathological, clinical or personal data were anonymized and separated from any personal identifiers. This study was examined and approved by the Military Hospital of Tunis (HMPIT) and Hospital of Fatouma Bourguiba of Mornastir (Tunisia), and was made with the consent of the patients’ relatives or their family (in autopsy cases).

In order to determine serum PSA levels before the radical prostatectomy, we uses the PSA DPC immunoassays (gifts of Diagnostics Products Corporation, Los Angeles, CA). These assays were performed according to the manufacturer’s instructions. The primary antibodies (Santa Cruz Biotechnology, Ca, USA) were: goat anti-human TNF-α, rabbit anti-human IL-1α and rabbit anti-human IL-6, at 1:50 (immunohistochemistry) or 1:200 (Western blot) dilutions. In Western blot, mouse anti-chicken α-actin (Amersham, Madrid, Spain) was also used at the same dilution to examine the relative expression of the other proteins.

Procedure specificity was checked using negative and positive controls. For negative controls sections of different group (normal, BPH and cancer) were incubated with preimmune serum at the same immunoglobulin concentration used for each antibody or with blocking peptides (Santa Cruz Biotechnology). As positive controls, histologic sections of thymus (TNF-α and IL-6) and tonsils (IL-1α) were incubated with the same antibodies used in this study.

For each antibody a histologic quantification of immunolabeling was performed. Of each prostate, six histologic sections were selected at random. Using the X40 objective staining intensity (optic density) per unit surface area was measured with an automatic image analyzer (Motic Images Advanced version 3.2, Motic China Group Co., China). For each positively immunostained section, one negative control section was also used, and the optic density of this control section was taken away from that of the stained section. After, means ± SD for each prostatic type were calculated. The same calculation was performed in normal and cancer samples. The statistical significance between means of the different prostate samples was assessed by the Fisher exact and the one-way ANOVA test at p≤0.05 (GraphPad PRISMA 3.0 computer program).

RESULTS

Western blot

In BPH and PC, a single band (at their corresponding molecular weight) was found to IL-1α (17 kDa), TNF-α (17 kDa) and IL-6 (21 kDa) (Figure 1). In normal prostates, only TNF-α and IL-6 were detected (at their corresponding molecular weight) (Figure1).

Average optical densities to IL-1α and IL-6 showed that in PC was more elevated than in BPH, and at the same in BPH was more elevated than in normal prostate. No variations in the average optical densities were found in TNF-α (Table I).

Immunohistochemistry

No immunoreaction was observed in negative controls incubated with pre-immune serum, or using the antibodies preabsorbed with an excess of purified antigens. Staining of thymus (IL-6 and TNF-α) or tonsil (IL-1) sections used as positive controls was always positive.

For each antibody assayed, the percentages of positive cases are shown in Table II.

No immunoreaction was found to IL-1α in normal prostate samples. In BPH patients (Figure 2A) immunostaining was positive in 50% of samples with PSA levels between 0-4 ng/ml and 60% of patients with PSA levels between 4-20 ng/ml (Table II). In PC
samples (Fig. 2B). IL-1α was expressed in the groups with PSA levels between 4-20 ng/ml (75%) and PSA >20 ng/ml (70% of patients).

Immunoreaction to TNFα (Figure 3A-B) was found in NP (40%), BPH (40%) and PC (35.2%) patients (Table II). In BPH (Figure 2C) the percentage of patients’ positives samples decrease when increase PSA levels. In PC (Figure 2D), immunoreaction to TNFα was only positive in two PSA levels groups (PSA levels between 4-20 ng/ml and >20 ng/ml).

In normal prostates, all samples were positives to IL-6. In BPH (Fig. 2E) patients, immunoreaction to IL-6 was positive in the two groups: 0-4 ng/ml (68% of patients) and 4-20 ng/ml of PSA (76.5% of patients). Positive IL-6 immunostaining was seen in 76.5% of PC (Figure 2F, Table II).

**DISCUSSION**

Catalona et al. (30) established a reference range of 0-4 ng/ml to define normal PSA levels, with a range of 4-10 ng/ml increase the probability of cancer, and levels above 10 ng/ml have a high probability of prostate cancer. Although other studies suggested that PSA levels of 10 ng/ml has little correlation with cancer prostate (31). In this manuscript we present PSA levels above 20ng/ml as the group with elevate probability of cancer patients.

In the present study, IL-1α (not detected in normal prostates) was expressed in 56% of BPH and 70% of PC patients. Similar results have been obtained by our group in previous studies (4), suggesting that IL-1α in epithelial cells seems to be involved in increased cell proliferation associated with BPH and Cancer. In patients with elevated expressions of IL-1α had also elevated PSA levels (PSA>20ng/ml). These findings suggest that in both BPH and PC, high expressions of IL-1α in epithelial cells may be related to cell proliferation and high PSA levels.

In a melanoma cell line (A375), Lázár-Molnár et al. (32) found that endogenous IL-1α production was associated with resistance to exogenous anti-proliferative stimuli and also to the increase in the constitutive production of IL-6 in autocrine manner. Previous results from our laboratory revealed increased IL-6 expression in cancer prostate epithelial cells (22). Since the increase in IL-6 has been associated with bad prognosis in prostate cancer (33-34), the evaluation of IL-1α as IL-6 inductor might be important to assess malignancy. Similar results have been described in breast carcinoma, where expression of IL-6 was enhanced and associated with proliferation cellular markers (20).

The effect of IL-6 on prostate cancer cells is contradictory. Some reports suggested that IL-6 could promote prostate cancer cell growth (35-36), whereas other studies could repress prostate cancer cell growth (26). In our study, we found expression of IL-6 in BPH and PC samples but also that high expression of IL-6 was associated with IL-1α (64.7% samples) and high PSA levels (>20 ng/ml).

TNFα has shown contradictory effects. Lejeune y cols. (37) observed that local administration

![Figure 1](image)

**FIGURE 1.** Western blot analysis of IL-1α, TNFα and IL-6 after 1.5% polyacrylamide gel electrophoresis. PN: normal prostate. HBP: benign prostate hyperplasia. CP: prostate carcinoma. The lanes showing a band correspond to a positively stained prostate of each group.

**TABLE I.**

<table>
<thead>
<tr>
<th></th>
<th>IL-1α</th>
<th>IL-6</th>
<th>TNFα</th>
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<tbody>
<tr>
<td>PN</td>
<td>—</td>
<td>7.09±1.2a</td>
<td>17.5±3.1a</td>
</tr>
<tr>
<td>HBP</td>
<td>8.3±0.89a</td>
<td>18.45±2.1b</td>
<td>16.8±1.6a</td>
</tr>
<tr>
<td>CP</td>
<td>20.5±2.3b</td>
<td>19.73±1.6b</td>
<td>18.8±3.6a</td>
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</table>
FIGURE 2. IL-1α showed cytoplasmic immunoreaction of epithelial cells in BPH (A) and PC (B). We showed a BPH (C) image with negative reactions to TNF. In PC (D) TNF immunoreaction was observed in most epithelial cells. IL-6 was also observed in epithelial cells of BPH (E) and PC (F). Bars: 20 μm (A-D) and 30 μm (E-F).
of TNF-α in high doses destroyed selectively the tumour blood vessel of the tumor and therefore his great anticarcinogenic power in melanoma. Nevertheless, different studies suggest that endogenous TNF-α is involved in the growth of tumoral cells and in the interactions with the stroma, increasing the tumour invasion and the development of metastasis (7, 8). In our cancer patients, we found that high expressions of IL-6 were associated with TNF-α (35.2% samples) but also with high PSA levels (>20 ng/ml).

In agreement with our results, we suggest that in BPH, the high expression of pro-inflammatory cytokines might be associated with high production of PSA. In CP, the high expression of these cytokines and the high levels of PSA might be involved in the change of polarity of the epithelial cells as well as the quantity of basal cells. At the same time, these cytokines might be involved indirectly in the increase of circulatory levels of PSA.

### CONCLUSIONS

In CP was an association between the high expression of pro-inflammatory cytokines (TNFα, IL-6, IL-1), inflammation, elevated PSA serum levels and tumor progression. Inhibition of IL-1, IL-6 and TNFα might be a possible target for PC treatment. A better understanding of the biologic mechanism and role played by the elevation of circulating PSA related to the tissue expression of these cytokines may improve clinical management and provide new targets for therapy in these patients.

### ACKNOWLEDGEMENTS

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### TABLE II.

<table>
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<tr>
<th>PSA levels</th>
<th>IL-1α</th>
<th>D. O.</th>
<th>TNF-α</th>
<th>D. O.</th>
<th>IL-6</th>
<th>D. O.</th>
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<tbody>
<tr>
<td>Ng/ml</td>
<td>%</td>
<td></td>
<td>%</td>
<td></td>
<td>%</td>
<td></td>
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<tr>
<td>PN (5)</td>
<td>0 (0%)</td>
<td>1.73±1.51</td>
<td>2 (40%)</td>
<td>12.03±2.9</td>
<td>5 (100%)</td>
<td>4.83±2.65</td>
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<td>&lt;4 (5)</td>
<td>14 (56%)</td>
<td>9.43±2.51</td>
<td>10 (40%)</td>
<td>9.87±3.85</td>
<td>17 (68%)</td>
<td>23.47±4.65</td>
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<tr>
<td>HBP (25)</td>
<td>5 (50%)</td>
<td>5 (33%)</td>
<td>5 (33%)</td>
<td>11 (73%)</td>
<td>11 (73%)</td>
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<td>0-4 (10)</td>
<td>9 (60%)</td>
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<td>&gt;20 (0)</td>
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<tr>
<td>CP (17)</td>
<td>12 (70.5%)</td>
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<td>0-4 (0)</td>
<td>3 (75%)</td>
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<td>4-20 (4)</td>
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<td>&gt;20 (13)</td>
<td>9 (70%)</td>
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(*of special interest, **of outstanding interest)


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