



## REVIEW

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**New ophthalmic drug delivery systems****Nuevos sistemas de liberación de fármacos a nivel ocular**Ana Castro-Balado<sup>1,2,3</sup>, Cristina Mondelo-García<sup>1,2</sup>, Irene Zarra-Ferro<sup>1,2</sup>, Anxo Fernández-Ferreiro<sup>1,2</sup><sup>1</sup>Servicio de Farmacia, Área Sanitaria de Santiago de Compostela e Barbanza (SERGAS), Santiago de Compostela. Spain. <sup>2</sup>Grupo de Farmacología Clínica, Instituto de Investigación Sanitaria Santiago de Compostela (IDIS), Santiago de Compostela. Spain. <sup>3</sup>Departamento de Farmacología, Farmacia y Tecnología Farmacéutica, Facultad de Farmacia, Universidad de Santiago de Compostela, Santiago de Compostela. Spain.**Author of correspondence**Anxo Fernández Ferreiro  
Servicio de Farmacia  
Hospital Clínico Universitario  
Santiago de Compostela  
Travesía Choupana, s/n.  
15706 Santiago de Compostela, Spain.Email:  
anxordes@gmail.comReceived 13 December 2019;  
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**Abstract**

For decades, topical ophthalmic drug administration through the use of eye drops has been the most widely used technique for the treatment of eye diseases. The development of galenic formulation has led to the use and commercialization of new formulations, such as suspensions, emulsions, and ophthalmic ointments that increase residence time in the site of action. Recently, new administration systems have been developed, such as devices and inserts that provide the sustained release of active substance. Some of these systems are already available on the market, whereas others are still undergoing clinical trials, such as promising systems based on nanostructures (nanocapsules, cyclodextrins, nanoemulsions, etc.). Similarly, various formulations and devices have been developed in the field of intravitreal administration, with different implants available on the European market for the treatment of age-related macular degeneration (AMD), diabetic macular edema, or infections that affect the posterior segment. This review includes current developments in ophthalmic topical and intravitreal drug administration routes as well as those under investigation.

**Resumen**

Desde hace décadas, la administración tópica oftálmica de fármacos mediante el empleo de colirios ha sido la técnica más empleada para el tratamiento de patologías oculares. El desarrollo de la galénica ha permitido el uso y comercialización de nuevas formulaciones que incrementan el tiempo de residencia en el lugar de acción, como es el caso de las suspensiones, emulsiones y pomadas oftálmicas. Recientemente se han desarrollado nuevos sistemas de administración, como es el caso de dispositivos e insertos que proporcionan una cesión sostenida de principio activo. Algunos de estos sistemas ya se encuentran disponibles en el mercado, mientras que otros todavía están en fase de ensayo clínico, como es el caso también de los prometedores sistemas basados en nanoestructuras (nanocápsulas, ciclodextrinas, nanoemulsiones, etc.). De la misma forma, diversas formulaciones y dispositivos han sido desarrollados en el campo de la administración intravítrea, estando disponibles en el mercado europeo diversos implantes para el tratamiento de la degeneración macular asociada a la edad (DMAE), el edema macular diabético o infecciones que afectan al segmento posterior. En esta revisión se recogen los desarrollos actualmente implementados y en fase de investigación asociados a las vías de administración oftálmica de fármacos tópicos e intravítrea.

**KEYWORDS**

Ophthalmic solutions; Intravitreal injections; Ocular inserts; Drug delivery systems.

**PALABRAS CLAVE**

Soluciones oftálmicas; Inyecciones intravítreas; Insertos oculares; Sistemas de liberación oftálmica.



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## Introduction

The eye is responsible for vision and is one of the most complex and sophisticated sense organs. Its anatomical structure and physiology is especially adapted to allow the passage of light and to provide protection against external agents<sup>1</sup>. At the anatomical level, it is divided into the anterior and posterior segments, which constitute one-third and two-thirds of the total dimension of the eye, respectively. The anterior segment includes the lacrimal apparatus, cornea, conjunctiva, anterior and posterior chambers, iris, ciliary bodies, lens, and aqueous humor; and the posterior segment consists of the sclera, choroid, retina, Bruch's membrane, vitreous humor, optic nerve, and retinal blood vessels<sup>2</sup>. Its different components make it operate as a complex system with numerous barriers that limit drug penetration within the organ<sup>3</sup>.

Ocular drug administration plays an important role in the treatment of common ocular diseases such as glaucoma, macular degeneration, diabetic retinopathy, infections (conjunctivitis, keratitis, endophthalmitis, etc.), and autoimmune disorders (Sjögren syndrome, uveitis, etc.). Each of the ocular administration routes (topical, intravitreal, and periorcular) has advantages and disadvantages, however, a key factor common to all these routes is low drug bioavailability at the target site<sup>4,5</sup>. Therefore, new ocular delivery systems need to be developed such that drugs can be made available at the target site in sufficient concentrations over a precise period of time to produce the desired effect.

The objective of this article is to review current drug delivery systems and those in development for each of the drug administration routes used in the treatment of eye disease.

We conducted an initial search of PubMed, Medline, Cochrane database, and Google Scholar including publications in English and Spanish. Various literature search strategies were conducted combining key terms and Medical Subject Headings (MeSH) to ensure that all relevant papers were found: "ocular drug delivery", "ophthalmic drug delivery", "intraocular drug delivery", "intravitreal delivery", "ocular inserts", and "intraocular devices". Selected articles included original papers and review articles. The ClinicalTrials database was also consulted for additional information on clinical trials related to new forms of ocular administration (<https://clinicaltrials.gov>).

## Physiological barriers

### Tear film

The tear layer is a thin liquid film (3 μm) covering the corneal and conjunctival surfaces. It has different functions, among which its lubricant, nutritional, and antibacterial activity are of special relevance. Under normal conditions, its volume ranges between 8 μL and 10 μL although this can be exceeded in certain physiological and pathological situations. It consists of the lipid, aqueous, and mucin layers<sup>2</sup>. The outermost layer is the lipid layer, whose main function is to retard evaporation from the aqueous layer<sup>6</sup>; the middle layer is the aqueous layer, constituting around 90% of the total volume of tears; and the innermost layer is the mucin layer, constituted by glycoproteins which, due to their hydrophilicity, contribute to the adequate wetting of the cornea and conjunctiva<sup>7</sup>. Different protein compounds are also present, one of the most relevant is lysozyme, which has antimicrobial activity<sup>8</sup>.

Tears are the main barrier to the absorption of topically administered drugs. Drainage through the nasolacrimal duct, protein binding with drug molecules, and continuous tear turnover (1 mL/min) drastically reduce effective drug concentrations at the target site<sup>9,10</sup>.

### Cornea

The cornea is a transparent avascular connective tissue that acts as the first structural barrier of the eye. Together with the tear film, it provides an adequate refractive surface that confers optical functionality<sup>11</sup>. The cornea is the main absorption route of topically administered ocular drugs. It is made up of six different layers: epithelium, Bowman's membrane, stroma, Dua's layer, Descemet's membrane, and endothelium. The outer layer prevents absorption, especially that of hydrophilic substances. Next, the stroma comprises keratocytes and connective tissue, forms 90% of the thickness of the cornea, and constitutes the largest reservoir for hydrophilic substan-

ces<sup>9,12</sup>. The inner layer allows the passage of substances with relative ease, including high molecular weight proteins. Due to the nature of the different layers, drug absorption at the corneal level is fundamentally affected by their hydrophilic-lipophilic balance. However, the presence of erosions and ulcers implies greater drug penetration<sup>13</sup>.

## Blood-ocular barriers

Blood-ocular barriers protect the eye from circulating substances in the blood and thus prevent many systemically administered drugs from entering the eye. There are two types of blood-ocular barrier: the blood-aqueous barrier, which is formed by the epithelium of the ciliary bodies and protects the anterior segment; and the blood-retinal barrier, which protects the posterior segment and controls the entry of drugs into the vitreous cavity from the systemic circulation<sup>14,15</sup>.

These barriers prevent drug access at the intraocular level, although some factors such as inflammation or edema can alter them and make drug concentrations at this level higher than expected<sup>16,17</sup>.

## Administration routes and drug delivery systems

The drug administration route is a key aspect of all ocular therapies. The topical route is the most frequently used<sup>18</sup>. Drugs administered by this route must meet a set of requirements. Tear fluid has a pH of between 7.4 and 7.7 and may be modified due to eye diseases. Eye drop pH has to be adapted to this range to avoid sensations such as pain, irritation, and tearing, although solutions with a pH other than physiological pH are sometimes used to optimize stability, solubility, and the degree of dissociation of the active principle. The buffering power of carbonic acid, weak organic acids, and proteins present in tear fluid relatively quickly neutralize solutions with a wide pH range (3.5-10.5) providing they are not buffered. The further the pH of the administered solution from the physiological pH of tears, the longer it takes to achieve neutralization<sup>19</sup>.

Sterility is one of the most important requirements because an association has been found between the use of contaminated preparations and different types of eye infections<sup>20</sup>. Clarity is also essential because suspended particles can cause corneal abrasions. The average particle size in most ophthalmic suspensions is less than 10 μm<sup>21</sup>. Finally, the osmolarity should be similar to that of the tear fluid (300.5 ± 7.2 mOsm/kg), although the eye can accept a certain range of osmotic pressure without irritation (osmolarity equivalent to sodium chloride solutions between 0.5% and 1.8%). Generally, slightly hypertonic mixtures are better tolerated than hypotonic ones, but they have the drawback that they cause the osmotic movement of water towards the conjunctival sac, causing dilution of the instilled drug<sup>19,20,22</sup>.

## Drug delivery systems to the anterior segment

After topical drug administration, absorption takes place either through the corneal route or the noncorneal route. Drugs with high corneal permeability, such as water-soluble small molecules (i.e. with molecular weights less than 100 Da<sup>23</sup>), follow the corneal route<sup>15,24</sup>. Such drugs pass through the epithelium, stroma, and endothelium by passive diffusion into the anterior chamber, where they perform their pharmacological function<sup>25,27</sup> or bind to melanin (iris and ciliary body)<sup>13</sup> or plasma proteins<sup>28</sup>. Any remaining drug and metabolites are eliminated through the trabecular meshwork through Schlemm's canal to the systemic blood circulation (conventional route) or through the iris to the uveoscleral outflow pathway (nonconventional route) and thence to the systemic blood circulation<sup>29,32</sup>. A smaller proportion of the drug reaches the posterior chamber through the iris by diffusion through the aqueous humor outflow pathway. As a result, vitreal drug concentrations are 10 to 100 times lower than in the aqueous humor and the cornea, respectively<sup>15</sup>. All these dynamics can be altered by eye movements<sup>33,34</sup> or ophthalmic diseases<sup>25,35,36</sup>.

Drugs with low corneal permeability (high molecular weight molecules) enter the eye through the conjunctiva or the sclera following the noncorneal route<sup>30,37,40</sup>. This route is relevant in the case of small moderately lipophilic molecules such as timolol<sup>41</sup>. In general, the noncorneal route is

of less importance although it can be artificially enhanced with the use of iontophoresis<sup>42</sup>. Drugs reach the vitreous humor through passive diffusion and active transport mechanisms. Once inside the vitreous chamber, the aqueous humor flow either transports the drug to the anterior chamber or it is eliminated by passive diffusion through the retinal pigment epithelium and retinal capillary endothelial cells (blood-retinal barrier) via the choroidal circulation towards the systemic blood circulation<sup>15,30,43</sup>. In comparison to the corneal absorption route, the noncorneal route provides a 20-fold lower drug concentration in the anterior chamber<sup>44</sup>.

#### Topical administration

Eye drops and ointments are the most widely used ophthalmic drug delivery systems (approximately 90%)<sup>18</sup> due to their low production cost and ease of administration (Figure 1). Nevertheless, solutions have very low ocular bioavailability because of significant precorneal drug loss through tearing, blinking, systemic elimination, and tear renewal<sup>18</sup>. Frequent administrations are needed to maintain effective drug concentrations, leading to low therapeutic adherence and consequent therapeutic failure.

One of the most frequently used techniques to increase ocular surface contact time is the use of traditional viscous agents such as hydroxymethyl cellulose, polyvinyl alcohol, hydroxypropyl methylcellulose, polyethylene glycols, and polyvinylpyrrolidone. Natural polymers are also used such as hyaluronic acid, Veegum<sup>®</sup>, alginates, xanthan gum, gelatin, acacia, and tragacanth<sup>21,45</sup>.

Ophthalmic emulsions improve the solubility and bioavailability of insoluble drugs, provide sustained release, improve corneal absorption, and prolong precorneal residence time. In ophthalmology, o/w emulsions are preferred over w/o emulsions because of better tolerability and decreased ocular irritation due to the external aqueous phase<sup>46</sup>. Suspensions can be used as an alternative vehicle with low-solubility drugs, thus increasing the drug particle retention time in suspension as well as contact time. Table 1 shows some examples of ophthalmic emulsions and suspensions currently available on the market.

After solutions and suspensions, ophthalmic ointments are the most popular dosage forms. The vehicle/base for ophthalmic use must not cause eye discomfort and must be compatible with the rest of the formulation components. There is a wide variety of ophthalmic ointments available on the market, most of which have anti-infective, anti-edemic, or anti-inflammatory properties (Table 1). There has been a recent increase in the use of water-soluble bases known as gels because of their advantages over vaseline bases, such as better spreading capacity and their characteristics of pH, lubricity, stability, and low irritability. Polymers used for the preparation of gels generally include PEG 200, PEG 400, carboxymethyl cellulose, carbopol, methyl polymethacrylate, and Lutrol FC-127. These polymers also have mucoadhesive properties that improve drug contact time<sup>21</sup>. Recently, hospital pharmacy services have begun to include these kinds of ingredients in compounded preparations, such as the use of hyaluronic acid in artificial tears and carboxymethyl cellulose in the production of vancomycin and cysteamine eye drops<sup>47</sup>.

Another strategy is the use of prodrugs, such as the prostaglandin F<sub>2α</sub> analogues Bimatoprost (Lumigan<sup>®</sup>), Travaprost (Travatan<sup>®</sup>), and Latanoprost (Xalatan<sup>®</sup>), which are used to reduce intraocular pressure in glaucoma. Furthermore, the Food and Drug Administration (FDA) of the United States has authorized loteprednol etabonate (Lotemax<sup>®</sup>), a topical corticosteroid derived from the chemical structure of prednisolone, for the treatment of eye allergies and inflammations, although it is not available in the European market<sup>48</sup>.

Advances have recently been made in research on nanocarriers. These systems allow the administration of a wide variety of active ingredients (including macromolecules) providing stable conditions for labile compounds and better control of drug delivery, thus improving ocular bioavailability. There is a wide variety of components within this group such as polymeric nanoparticles, lipid nanoparticles, cyclodextrins, liposomes, niosomes, dendrimers, nanosuspensions, and emulsions<sup>21,42,49</sup>, all of which are currently under investigation. These types of systems could be used in the field of compounding, such as cyclodextrins<sup>50,51</sup>, which reduce the toxicity of active ingredients<sup>52</sup> and enhance the solubility of

**Table 1.** Commercial presentations for the topical ophthalmic administration of sustained-release drugs

Type of formulation	Presentation	Active principle	Indication
Emulsions	Restasis <sup>®</sup>	Cyclosporine	Dry eye
	Ikervis 1 mg/mL <sup>®</sup>	Cyclosporine	Dry eye
	AzaSite <sup>®</sup>	Azithromycin	Azithromycin-responsive eye infections
	Refresh Endura <sup>®</sup>	No medication	Dry eye
	Durezol <sup>®</sup>	Difluprednate	Anterior ocular uveitis
Suspensions	Azopt <sup>®</sup>	Brinzolamide	Open-angle glaucoma and ocular hypertension
	Maxidex <sup>®</sup>	Dexamethasone	Non-infectious eye inflammation
	TobraDex <sup>®</sup>	Tobramycin and dexamethasone	Tobramycin-sensitive bacterial eye infections
Ointments	Oftacilox 3 mg/g <sup>®</sup>	Ciprofloxacin	Ciprofloxacin sensitive purulent bacterial conjunctivitis and blepharitis
	Oftalmolosa Cusi Antiedema 50 mg/g <sup>®</sup>	Sodium chloride	Corneal edema reduction
	Oftalmolosa Cusi Dexamethasone 0.5 mg/g <sup>®</sup>	Dexamethasone	Non-infectious eye inflammation
	Tobrex Ophthalmic Ointment 3 mg/g <sup>®</sup>	Tobramycin	Tobramycin-sensitive bacterial infections of the surface of the eye and its annexes
	Oculos Epitelizante Ointment <sup>®</sup> 3 mg/g + 5.5 mg/g + 5 mg/g	Retinol Methionine Gentamicin	Prophylaxis and bacterial infection treatment
Prodrugs	Lumigan 0.3 mg/ml <sup>®</sup>	Bimatoprost	Glaucoma
	Travatan 40 µg/ml <sup>®</sup>	Travaprost	Glaucoma
	Xalatan 50 µg/ml <sup>®</sup>	Latanoprost	Glaucoma

molecules that are barely soluble in vehicles used for ophthalmic administration.

Contact lenses are medical devices that can be loaded with active ingredients, thereby acting as non-invasive drug-delivery systems and providing sustained release in tear fluid located between the cornea and the lens<sup>54</sup>. Generally, drugs and polymers used for these lenses have low affinity. Thus, to achieve controlled sustained release, a series of modifications are needed such as the incorporation of polymeric nanoparticles, microemulsions, micelles, liposomes, diffusion barriers (e.g. vitamin E) and so on, or the use of sophisticated loading techniques such as molecular imprinting, drug-loaded films, ion ligand polymeric systems, or supercritical fluid technology<sup>55,56</sup>. When such modifications are introduced, it is essential that aspects such as water content, mechanical properties, ionic permeability, transparency, and oxygen permeability remain unchanged. Up to the present, hindrances to their commercialization have included issues such as drug stability during processing/manufacturing, achieving zero-order release kinetics, avoiding drug release during the post-manufacturing monomer extraction step, protein adherence, drug release during storage, and cost-benefits<sup>46</sup>. However, they have potential applicability in the field of ophthalmic compounding, because drugs and excipients can be easily loaded onto them under sterile conditions as part of a relatively simple low-cost formulation process. Thus, an ophthalmic drug delivery system could be made available that is easy and comfortable to administer to the patient and has a dosing schedule that leads to better adherence than that obtained under frequent instillation of ophthalmic drops. These aspects would be of interest in the setting of severe eye infections or rare diseases such as ocular cystinosis.

**Inserts and implants in the anterior segment**

The goal of ocular inserts and implants is to improve bioavailability and achieve sustained drug delivery. In order to administer medications to the anterior segment of the eye, these systems can be placed under the eyelid, at the bottom of the conjunctival sac, in the anterior chamber, in the subconjunctival space, or in the episcleral region (Figure 2). However, it is difficult to develop implants for the anterior chamber, because they can be susceptible to movement due to the low viscosity of the aqueous humor, causing irreversible damage to endothelial cells.

These devices can be biodegradable or nonbiodegradable (Table 2). Biodegradable or resorbable devices are noninvasive and do not need to be removed. However, most of these devices have limited action time and therefore may require frequent administration. Furthermore, the complete material and its metabolites must not be toxic. Other challenges include the prevention of accidental device loss and increased tear production after placement, which increases the risk of massive drug release<sup>57</sup>. The Ocuser<sup>®</sup> system was the first commercially available ocular insert for the treatment of glaucoma. This ocular therapeutic system delivers a small amount of pilo-

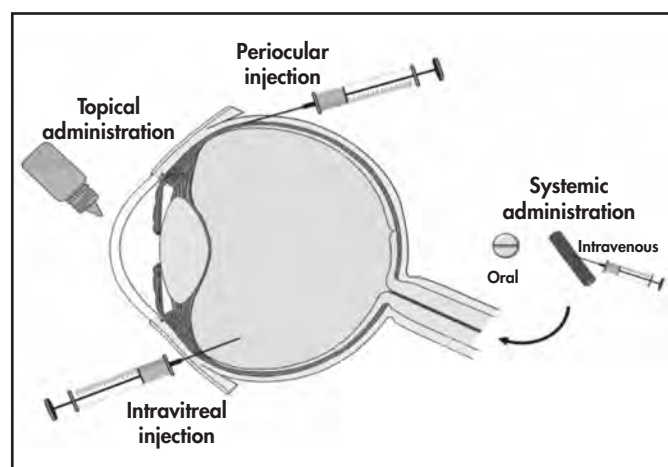
carpine over one week. It consists of two ethylene-vinyl acetate copolymer membranes that provide zero-order kinetics<sup>58</sup>. This insert was withdrawn from the market in 1998 as a consequence of the adverse effects of pilocarpine, such as eyebrow pain and miosis<sup>18</sup>.

The only resorbable conjunctival device on the market is Lacrisert<sup>®</sup>, which is a small hydroxypropyl cellulose tablet placed in the inferior conjunctival fornix. It slowly dissolves and creates an artificial tear film to treat dry eye<sup>59</sup>. However, it can cause blurred vision, leading to device removal in 8.7% of patients<sup>60</sup>. Another cul-de-sac implant is DSP-Visulex, which has currently completed a phase-II clinical trial for the treatment of anterior uveitis with dexamethasone (NCT02309385)<sup>61</sup>.

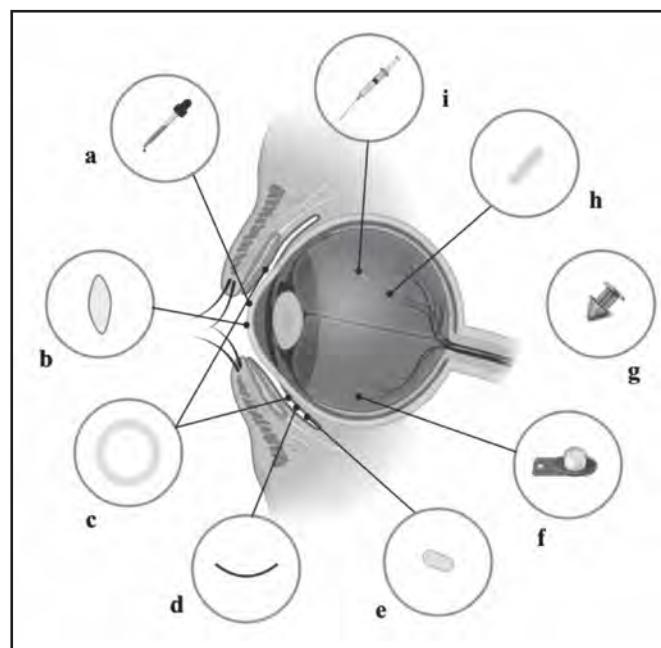
Nonresorbable devices form another group. These include rod-shaped devices, which are placed in the superior or inferior conjunctival fornix to administer drugs through the noncorneal absorption route. Since 2004, Mydrisart<sup>®</sup> has been available on the European market. It is an ethylcellulose tablet loaded with tropicamide hydrochloride and phenylephrine to obtain mydriasis two hours before surgery. However, there are no significant differences in pupil dilation between topical mydriatic eye drops and Mydrisart<sup>®</sup>, although its dilation speed is slower<sup>62</sup>. Another type of device is OphthaCoil<sup>®</sup>, a spiral stainless steel wire, which is placed in the lower conjunctival sac. It can carry medications on its inner lumen (loaded in microspheres or filaments) or outside on the SlipSkin<sup>®</sup> lining. These devices have been tested with pradofloxacin and mydriatic agents (phenylephrine hydrochloride and tropicamide) in Beagle dogs<sup>63</sup> and horses<sup>64</sup>.

The Helios<sup>™</sup> device consists of a polypropylene support ring within a silicone matrix loaded with bimatoprost. The results of clinical trials showed that its placement in the eye reduced intraocular pressure in glaucoma patients over six months. However, no significant differences were found between this device and regular 0.5% preservative-free timolol ophthalmic solution. The dropout rate was higher among patients with the Helios device. The main adverse event was mucus discharge<sup>65</sup>.

**Figure 1.** Ocular routes for drug delivery systems<sup>53</sup>.



**Figure 2.** Overview of drug delivery devices. a) Eye drops. b) Drug-loaded contact lenses. c) Conjunctival insert, such as Ocuser<sup>®</sup>. d) The OphthaCoil<sup>®</sup> conjunctival insert: a device placed behind the lower eyelid in the fornix conjunctiva. e) Conjunctival tablet, Lacrisert<sup>®</sup> or Mydrisart<sup>®</sup>: these tablets are placed in the inferior conjunctival fornix. f) Retrisert intravitreal insert: implanted in the vitreous chamber and sewn to the sclera. g) Punctal plug, such as Dextenza or Evolute. h) Intravitreal insert, such as Illuven or Surodex<sup>®</sup>: implanted or injected into the vitreous chamber. i) Intravitreal injection.



**Table 2.** Implants for treatment of anterior segment disease

Implant	Type of system	Drug	Duration	Indication
Ocusert® 5 mg	Oval device (nonbiodegradable)	Pilocarpine	1 week	Glaucoma
Lacrisert® 5 mg	Conjunctival tablet (biodegradable)	Hydroxypropyl cellulose	1 day	Dry eye
Surodex® 60 µg	Anterior chamber implant (biodegradable)	Dexamethasone	10 days	Inflammation after cataract surgery
Dextenza® 0.4 mg	Punctal plug PEG (nonbiodegradable)	Dexamethasone	30 days	Inflammation and postsurgical pain
Evolute®	Punctal plug (nonbiodegradable)	Latanoprost	14 weeks	Glaucoma
Mydrasert® 0.28 mg/5.4 mg	Conjunctival tablet (nonbiodegradable)	Tropicamide/phenylephrine	2 hours	Obtain preoperative mydriasis

Punctal plugs are another type of ocular device that have shown potential as drug delivery systems. These plugs are small biocompatible implants that were initially used to treat dry eye by inserting them into the tear ducts or to block tear drainage. They have the advantages of being non-invasive and maintaining the sustained release of the drug through diffusion. However, they have an increased risk of associated complications, such as conjunctivitis, corneal abrasion, tear duct obstruction, excessive tearing, and plug extrusion<sup>42</sup>. The materials used in their preparation include silicone, hydroxyethyl methacrylate, and polycaprolactone, but they require removal after the complete release of the active ingredient. Clinical trials are currently underway on the administration of latanoprost, bimatoprost, olopatadine, moxifloxacin, and cyclosporine A<sup>18</sup>. In 2018, the FDA authorized the marketing of Dextenza®, which contains dexamethasone 0.4 mg for the treatment of postoperative inflammation and pain. However, this format is not available in the European market.

The transfer of these types of devices and implants to the setting of ophthalmic compounding formulation is unlikely because of the complexity of their preparation and sterilization, which would require equipment not currently available in hospital pharmacy services.

## Drug delivery systems to the posterior segment

In the field of ophthalmology, intravitreal administration has become one of the most widely performed procedures for the administration of anti-VEGF antibodies (bevacizumab, ranibizumab, aflibercept) in patients with age-related vascular degeneration (AMD)<sup>66</sup>.

### Intravitreal injections

In intravitreal administration, the needle is inserted perpendicular to the sclera, between the horizontal and vertical rectus muscles, in order to release the drug in the vitreous chamber where it is distributed and reaches different targets in the posterior eye segment<sup>67</sup> (Figure 1). Depending on the nature of the drug and the formulation used, the vitreous humor can act as an important barrier to the release of the drug or its influence may be insignificant<sup>15</sup>.

After intravitreal administration, drug concentrations at the blood-ocular barriers are clearly higher than those obtained after topical or systemic administration. At this level, drug clearance can occur through the anterior or posterior pathways. Any drug can pass through the anterior route by diffusion through the vitreous humor to the posterior chamber, where the aqueous humor is located, and then eliminated through the trabecular meshwork. However, the posterior route is only available to those drugs that are capable of crossing the endothelium and epithelium that form the blood-ocular barriers. Although these blood-ocular barriers do not allow the passage of proteins and large molecules, they do allow the passage of small molecules<sup>15</sup>. As a consequence, the clearance of small molecules from the vitreous is much faster than that of molecules with high molecular weights<sup>15,53,68</sup>.

A priority in AMD research is to decrease the number of visits to the ophthalmologist and the number of intravitreal injections administered<sup>69</sup>. In this sense, the need to develop new intravitreal drug delivery systems is based on the fact that reductions in the frequency of administration decreases the risk of associated adverse effects and optimizes healthcare costs<sup>5</sup>. Thus, the vast majority of intravitreal drug delivery systems have been studied using anti-VEGF drugs.

### Ocular inserts in the posterior segment

There are various challenges related to the design of new intravitreal drug delivery systems. On the one hand, the size of these systems must be of the order of micrometers or nanometers to fit inside the vitreous chamber, however, they must be also able to contain the appropriate dose to obtain effective concentrations over long periods of time. On the other hand, sustained-release antibody formulations have to maintain antibody stability and bioactivity during storage and release in vivo, which represents another relevant challenge to their development<sup>70</sup>. Therefore, although numerous intravitreal delivery systems are under research, most of them are in very early stages and only a few have been tested in clinical trials<sup>5</sup>. Intravitreal anti-VEGF drug delivery systems in development include hydrogels, liposomes, microparticles, nanoparticles, implants, and composite systems (i.e. combinations of two of the foregoing systems)<sup>71</sup>.

At the clinical level, implants are the only delivery systems that have been marketed for intravitreal administration. They are small solid devices that are surgically implanted or injected into the vitreous humor<sup>46</sup>. In Europe, commercial implants are currently available that provide the prolonged release of dexamethasone (Ozurdex®), fluocinolone (Iluvien®), and ganciclovir (Vitrasert® 4.5 mg, not marketed in Spain) (Table 3). However, there are no implants with anti-VEGF antibodies currently on the market<sup>71</sup>.

Some implants for the treatment of AMD are under research. The NT/503 cell line is loaded into an encapsulated-cell technology delivery system, implanted into the vitreous cavity every 12 weeks, and produces VEGF antagonists that are released into the retina. This device underwent phase I/II clinical trials: however, although the procedure seemed well tolerated, the phase II trial was halted due to lack of reproducible long-term efficacy<sup>72</sup>. We also draw attention to the Port Delivery System, which is a permanent and reusable drug reservoir that is surgically implanted through an incision in the sclera. It is loaded with ranibizumab solution 20 µL (100 mg/mL) that is released into the vitreous through a semipermeable membrane by passive diffusion. The device carries a customized needle that cleans the device and refills it with ranibizumab. This device is currently being tested in the ARCHWAY phase-III clinical trial (NCT03677934)<sup>73</sup>.

**Table 3.** Intravitreal implants marketed in Europe

Implant	Type of system	Drug	Duration, months	Indication
<b>Ozurdex® 700 µg</b>	Matrix (biodegradable)	Dexamethasone	4	Diabetic macular edema
<b>Iluvien® 190 µg</b>	Reservoir (nonbiodegradable)	Fluocinolone	36	Diabetic macular edema
<b>Vitraser® 4.5 mg</b>	Polymeric (nonbiodegradable)	Ganciclovir	3	Cytomegalovirus retinitis

### Periocular administration: subconjunctival, suprachoroidal, and transscleral routes

Periocular administration routes deliver drugs into the periocular tissues, producing local effects or providing access to intraocular tissues. Periocular administration includes the peribulbar, posterior juxtасcleral, and retrobulbar routes, which are generally used to administer anesthetics; and the subtenon, subconjunctival, suprachoroidal, and transscleral routes, which are commonly used to administer drugs to the posterior segment<sup>2</sup> (Figure 1).

The subconjunctival route refers to the administration of an injection or an implant below the conjunctiva. In the case of injection, an initial accumulation of the drug is formed that acts as a depot that undergoes slow depletion. In this route, the drug avoids elimination by the conjunctival blood and lymphatic circulation and must cross the sclera and choroid to reach the retina<sup>74</sup>. A small incision in the conjunctiva must be made in order to deliver subconjunctival/episcleral implants. Surodex™ is a biodegradable anterior chamber insert approved in Singapore and China, among other countries<sup>75</sup>. It is made of poly(lactic-co-glycolic acid) (PLGA) that provides the sustained release of dexamethasone for approximately 10 days for the treatment of inflammation after cataract surgery<sup>42</sup>.

Drugs administered through the suprachoroidal route slowly diffuse from the suprachoroidal space to the choroid and retina, minimizing systemic adverse effects and those related to the intravitreal route (endophthalmitis, retinal detachment, and cataract formation). Compared to subconjunctival injection, this administration route provides increased greater bioavailability due to its proximity to the retina and choroid<sup>15</sup>, does not interfere with the optical route, and is less invasive than the transscleral route<sup>2,76</sup>. Prolonged action is needed for this administration route to be attractive in a clinical setting. Controlled drug release formulations have been tested for suprachoroidal administration mainly in the form of implants, micro-needles<sup>77</sup>, and micro/nano formulations to provide sustained release. All these approaches are still in preclinical and clinical phases<sup>78</sup>. Of these, we draw attention to the MicroPump®, a rechargeable nonbiodegradable implant comprising a microelectromechanical system that provides a constant release of a set volume of drug in the order of nanolitres<sup>76</sup>. Several phase-III clinical trials are currently underway of the suprachoroidal injection of triamcinolone acetonide in combination with various anti-VEGF agents for the treatment of AMD and posterior uveitis (NCT03203447, NCT02980874, NCT01789320)<sup>46</sup>.

The transscleral route bypasses the anterior segment barrier. Drugs administered through this route encounter the barriers of the sclera, choroidal blood flow, and retinal pigment epithelium. The transscleral route is less invasive and safer than the intravitreal route, but provides less intraocular bioavailability. This is because the removal path of the vitreous chamber moves outward while the drug moves inward (against the natural flow). These drawbacks can be overcome by modifications to the formulation<sup>2,79</sup>. Implants and iontophoretic devices are being developed to address this issue. Transscleral iontophoresis is a less invasive and well-tolerated method that allows drugs to be actively administered through the sclera. A mild electric current is used to allow the diffusion of the active ingredient from the reservoir to the site of action (retina/choroid). Within this approach we highlight the Visulex® iontophoretic device, which can release different corticosteroids via mild electrical currents. It has undergone a phase I/II clinical trial (NCT02309385) for the treatment of anterior uveitis. The success of iontophoresis-mediated drug delivery depends on several factors, such as charge density of the intended mole-

cule, electric current applied, the duration of treatment application, and position of electrode placement<sup>46</sup>.

### Systemic administration

Systemically administered drugs are distributed at the ocular level mainly in the choroid, iris, ciliary body, and retina. Specifically, the choroid receives around 85% of the ocular blood flow and has fenestrated blood vessels, allowing good extravascular distribution of drugs at this level. However, the passage of drugs from the choroid to the retina is obstructed by the pigment epithelial barrier<sup>80</sup>. In addition, the walls of the blood vessels of the iris and retina have very tight junctions between the endothelial cells, which slow down drug penetration. Consequently, small lipophilic compounds administered systemically can cross blood-ocular barriers, reaching and distributing to eye tissues, whereas the penetration of hydrophilic compounds and large molecules is highly restricted<sup>81</sup>.

In certain situations, systemic drug administration can be used to treat certain ocular pathologies. For example, antimicrobials can be administered for the treatment of endophthalmitis. Specifically, levofloxacin, moxifloxacin, linezolid, and meropenem are the best-documented agents that achieve therapeutic levels in the vitreous and penetrate to the site of action<sup>53,82</sup>. In addition, oral acetazolamide is administered for the treatment of glaucoma, systemic corticosteroids and anti-TNF antibodies for uveitis, and intravenous mannitol infusions for ocular hypertension. It is often the case that high doses of these drugs need to be administered such that they reach the ocular tissues in sufficient concentrations to exert the desired pharmacological effect. As a consequence, systemic adverse effects may appear<sup>81</sup>.

Verteporfin (Visudyne®) is currently available on the market for the treatment of exudative or wet AMD or systemic pathological myopia. This drug is administered by intravenous infusion and must be activated by light<sup>83</sup>.

The passage of substances from the bloodstream to the ocular tissues implies that they are sometimes exposed to systemically administered drugs that are not used for the treatment of ocular disease. This circumstance may lead to adverse effects at the ocular level, especially in highly polymedicated elderly patients. Drugs that cause toxicity at the retinal level after systemic administration include chloroquine, sildenafil, chlorpromazine, carmustine, and vigabatrin. Most of these adverse effects are relatively rare and transient under acute drug administration, but they can become more severe and frequent under chronic or long-term administration<sup>81,84</sup>.

### Conclusions

Ocular drug delivery remains a major challenge for ophthalmology. Traditional formulations offer short drug residence times in the site of action, leading to the need for frequent administration, which directly affects therapeutic adherence and ultimately treatment success. Over the last few decades, some improvements have become commercially available, such as the use of emulsions, suspensions, and ointments, although none of these delivery systems achieve sufficiently long residence times. Recent studies have addressed new formulation techniques, such as the use of nanocarriers, medicated contact lenses, and implant/insert-type devices. Despite this activity, only a small number of these administration systems have been marketed for the treatment of diseases that affect the anterior chamber or the

posterior chamber. For this reason, it remains relevant to continue investing in research on the development of new therapeutic strategies in order to improve the treatment and quality of life of patients and their use in routine clinical practice.

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## Conflict of interest

No conflicts of interest.

## Bibliography

- Cabrera FJ, Wang DC, Reddy K, Acharya G, Shin CS. Challenges and opportunities for drug delivery to the posterior of the eye. *Drug Discov Today*. 2019;24(8):1679-84.
- Nayak K, Misra M. A review on recent drug delivery systems for posterior segment of eye. *Biomed Pharmacother*. 2018;107:1564-82. DOI: 10.1016/j.biopha.2018.08.138
- Agrahari V, Agrahari V, Mandal A, Pal D, Mitra AK. How are we improving the delivery to back of the eye? Advances and challenges of novel therapeutic approaches. *Expert Opin Drug Deliv*. 2017;14(10):1145-62. DOI: 10.1080/17425247.2017.1272569
- Ranta VP, Mannerman E, Lumppuro K, Subrizi A, Laukkanen A, Antopolsky M, *et al*. Barrier analysis of periocular drug delivery to the posterior segment. *J Control Release*. 2010;148(1):42-8. DOI: 10.1016/j.jconrel.2010.08.028
- Subrizi A, del Amo EM, Korzhikov-Vlakh V, Tennikova T, Ruponen M, Urtti A. Design principles of ocular drug delivery systems: importance of drug payload, release rate, and material properties. *Drug Discov Today*. 2019;24(8):1446-57. DOI: 10.1016/j.drudis.2019.02.001
- Millar TJ, Schuett BS. The real reason for having a meibomian lipid layer covering the outer surface of the tear film – A review. *Exp Eye Res*. 2015;137:125-38. DOI: 10.1016/j.exer.2015.05.002
- Wang J, Fonn D, Simpson TL, Jones L. Precorneal and pre- and postlens tear film thickness measured indirectly with optical coherence tomography. *Invest Ophthalmol Vis Sci*. 2003;44(6):2524. DOI: 10.1167/iov.02-0731
- Ragland SA, Criss AK. From bacterial killing to immune modulation: Recent insights into the functions of lysozyme. *PLoS Pathog*. 2017;13(9):e1006512. DOI: 10.1371/journal.ppat.1006512
- Prausnitz MR, Noonan JS. Permeability of cornea, sclera, and conjunctiva: A literature analysis for drug delivery to the eye. *J Pharm Sci*. 1998;87(12):1479-88. DOI: 10.1021/js9802594
- Fernández-Ferreiro A, Barcia M, Gil-Martínez M, Méndez J, Luaces-Rodríguez A, Tomé V, *et al*. Critical factors involved in the determination of the optimal concentration of ophthalmic anti-infective compounded drugs. *Int J Clin Pharmacol Ther*. 2016;1. DOI: 10.15344/2456-3501/2016/122
- Del Monte DW, Kim T. Anatomy and physiology of the cornea. *J Cataract Refract Surg*. 2011;37(3):588-98. DOI: 10.1016/j.jcrs.2010.12.037
- Moiseev RV, Morrison PWJ, Steele F, Khutoryanskiy VV. Penetration enhancers in ocular drug delivery. *Pharmaceutics*. 2019;11(7):321. DOI: 10.3390/pharmaceutics11070321
- Agrahari V, Mandal A, Agrahari V, Trinh HM, Joseph M, Ray A, *et al*. A comprehensive insight on ocular pharmacokinetics. *Drug Deliv Transl Res*. 2016;6(6):735-54. DOI: 10.1007/s13346-016-0339-2
- González Barcia M, Esteban Cartelle H. Formulación magistral en oftalmología. En: Aspectos prácticos de la farmacotecnia en un Servicio de Farmacia. Madrid: Master Line & Prodigio; 2011. p. 245-74.
- Del Amo EM, Rimpelä AK, Heikkinen E, Kari OK, Ramsay E, Lajunen T, *et al*. Pharmacokinetic aspects of retinal drug delivery. *Prog Retin Eye Res*. 2017;57:134-85. DOI: 10.1016/j.preteyeres.2016.12.001
- Miyamoto N, de Kozak Y, Normand N, Courtois Y, Jeanny JC, BenEzra D, *et al*. PIGF-1 and VEGFR-1 Pathway Regulation of the External Epithelial Hemato-Ocular Barrier. *Ophthalmic Res*. 2008;40(3-4):203-7. DOI: 10.1159/000119877
- Reimondez-Troitiño S, Csaba N, Alonso MJ, de la Fuente M. Nanotherapies for the treatment of ocular diseases. *Eur J Pharm Biopharm*. 2015;95(Pt B):279-93. DOI: 10.1016/j.ejpb.2015.02.019
- Bertens CJF, Gijs M, van den Biggelaar FJHM, Nuijts RMMA. Topical drug delivery devices: A review. *Exp Eye Res*. 2018;168:149-60. DOI: 10.1016/j.exer.2018.01.010
- Castro-Balado A, González-López J, Blanco-Méndez J. Requerimientos básicos para la elaboración de colirios. En: Fernández-Ferreiro A. Formulación Magistral Oftálmica Antiinfecciosa. Madrid: Sociedad Española de Farmacia Hospitalaria; 2019. p. 63-70.
- Mehta S, Armstrong BK, Kim SJ, Toma H, West JN, Yin H, *et al*. Long-term potency, sterility, and stability of vancomycin, ceftazidime, and moxifloxacin for treatment of bacterial endophthalmitis. *Retina*. 2011;31(7):1316-22. DOI: 10.1097/IAE.0b013e31820039af
- Yellepeddi VK, Palakurthi S. Recent advances in topical ocular drug delivery. *J Ocul Pharmacol Ther*. 2016;32(2):67-82. DOI: 10.1089/jop.2015.0047
- Awwad S, Mohamed Ahmed AHA, Sharma G, Heng JS, Khaw PT, Brocchini S, *et al*. Principles of pharmacology in the eye. *Br J Pharmacol*. 2017;174(23):4205-23. DOI: 10.1111/bph.14024
- Fernández-Ferreiro A, Luaces-Rodríguez A, Lamas-Díaz MJ. La formulación magistral de antiinfecciosos tópicos en oftalmología. En: Formulación Magistral Oftálmica Antiinfecciosa. Madrid: Sociedad Española de Farmacia Hospitalaria; 2019. p. 21-32.
- Edwards A, Prausnitz MR. Predicted permeability of the cornea to topical drugs. *Pharm Res*. 2001;18(11):1497-508. DOI: 10.1023/a:1013061926851
- Ho LC, Conner IP, Do CW, Kim SG, Wu EX, Wollstein G, *et al*. In vivo assessment of aqueous humor dynamics upon chronic ocular hypertension and hypotensive drug treatment using gadolinium-enhanced MRI. *Invest Ophthalmol Vis Sci*. 2014;55(6):3747-57. DOI: 10.1167/iov.14-14263
- Shikamura Y, Yamazaki Y, Matsunaga T, Sato T, Ohtori A, Tojo K. Hydrogel ring for topical drug delivery to the ocular posterior segment. *Curr Eye Res*. 2016;41(5):653-61. DOI: 10.3109/02713683.2015.1050738
- Zhang W, Prausnitz MR, Edwards A. Model of transient drug diffusion across cornea. *J Control Release*. 2004;99(2):241-58. DOI: 10.1016/j.jconrel.2004.07.001
- Pelkonen L, Tengvall-Unadake U, Ruponen M, Kidron H, del Amo EM, Reinisalo M, *et al*. Melanin binding study of clinical drugs with cassette dosing and rapid equilibrium dialysis inserts. *Eur J Pharm Sci*. 2017;109:162-8. DOI: 10.1016/j.ejps.2017.07.027
- Carreon TA, Edwards G, Wang H, Bhattacharya SK. Segmental outflow of aqueous humor in mouse and human. *Exp Eye Res*. 2017;158:59-66. DOI: 10.1016/j.exer.2016.08.001
- Del Amo EM, Urtti A. Rabbit as an animal model for intravitreal pharmacokinetics: Clinical predictability and quality of the published data. *Exp Eye Res*. 2015;137:111-24. DOI: 10.1016/j.exer.2015.05.003
- Goel M, Picciani RG, Lee RK, Bhattacharya SK. Aqueous humor dynamics: A review. *Open Ophthalmol J*. 2010;4:52-9. DOI: 10.2174/1874364101004010052
- Loewen RT, Brown EN, Roy P, Schuman JS, Sigal IA, Loewen NA. Regionally discrete aqueous humor outflow quantification using fluorescein analogs. *PLOS ONE*. 2016;11(3):e0151754. DOI: 10.1371/journal.pone.0151754
- Bonfiglio A, Lagazzo A, Repetto R, Stocchino A. An experimental model of vitreous motion induced by eye rotations. *Eye Vis*. 2015;2(1):10. DOI: 10.1186/s40662-015-0020-8
- Stocchino A, Repetto R, Cafferata C. Eye rotation induced dynamics of a Newtonian fluid within the vitreous cavity: the effect of the chamber shape. *Phys Med Biol*. 2007;52(7):2021-34. DOI: 10.1088/0031-9155/52/7/016

35. Guo T, Sampathkumar S, Fan S, Morris N, Wang F, Toris CB. Aqueous humor dynamics and biomechanics in the ageing Chinese eye. *Br J Ophthalmol*. 2017;101(9):1290-6. DOI: 10.1136/bjophthalmol-2016-309883
36. Li J, Lan B, Li X, Sun S, Lu P, Cheng L. Effect of intraocular pressure (IOP) and choroidal circulation on controlled episcleral drug delivery to retina/vitreous. *J Control Release*. 2016;243:78-85. DOI: 10.1016/j.jconrel.2016.10.001
37. Ambati J, Canakis CS, Miller JW, Gragoudas ES, Edwards A, Weissgold DJ, et al. Diffusion of high molecular weight compounds through sclera. *Invest Ophthalmol Vis Sci*. 2000;41(5):1181-5.
38. Bauer NJ, Motamedi M, Wicksted JP, March WF, Webers CA, Hendrikse F. Non-invasive assessment of ocular pharmacokinetics using confocal raman spectroscopy. *J Ocul Pharmacol Ther*. 1999;15(2):123-34. DOI: 10.1089/jop.1999.15.123
39. Hughes PM, Olejnik O, Chang-Lin JE, Wilson CG. Topical and systemic drug delivery to the posterior segments. *Adv Drug Delivery Reviews*. 2005;57(14):2010-32. DOI: 10.1016/j.addr.2005.09.004
40. Lee SJ, Kim SJ, Kim ES, Geroski DH, McCarey BE, Edelhauser HF. Trans-scleral permeability of Oregon Green 488®. *J Ocul Pharmacol Ther*. 2008;24(6):579-86. DOI: 10.1089/jop.2008.0050
41. Järvinen K, Järvinen T, Urtti A. Ocular absorption following topical delivery. *Adv Drug Delivery Reviews*. 1995;16(1):3-19. DOI: 10.1016/0169-409X(95)00010-5
42. Yavuz B, Kompella UB. Ocular Drug Delivery. En: Whitcup SM, Azar DT, editores. *Pharmacologic therapy of ocular disease. Handbook of Experimental Pharmacology*, Vol. 242. Springer International Cham; 2017; p. 57-93. DOI: 10.1007/164\_2016\_84
43. Kidron H, Del Amo EM, Vellonen KS, Urtti A. Prediction of the vitreal half-life of small molecular drug-like compounds. *Pharm Res*. 2012;29(12):3302-11. DOI: 10.1007/s11095-012-0822-5
44. Ahmed I, Patton TF. Importance of the noncorneal absorption route in topical ocular drug delivery. *Invest Ophthalmol Vis Sci*. 1985;26(4):584-7.
45. Fernández-Ferreiro A, González Barcia M, Gil-Martínez M, Vieites-Prado A, Lema I, Argibay B, et al. In vitro and in vivo ocular safety and eye surface permanence determination by direct and magnetic resonance imaging of ion-sensitive hydrogels based on gellan gum and kappa-carrageenan. *Eur J Pharm Biopharm*. 2015;94:342-51. DOI: 10.1016/j.ejpb.2015.06.003
46. Gote V, Sikder S, Sicotte J, Pal D. Ocular drug delivery: Present innovations and future challenges. *J Pharmacol Exp Ther*. 2019;ijpet.119.256933. DOI: 10.1124/jpet.119.256933
47. Luaces-Rodríguez A, Díaz-Tomé V, González-Barcia M, Silva-Rodríguez J, Herranz M, Gil-Martínez M, et al. Cysteamine polysaccharide hydrogels: Study of extended ocular delivery and biopermanence time by PET imaging. *Int J Pharm*. 2017;528(1):714-22. DOI: 10.1016/j.ijpharm.2017.06.060
48. Yellepeddi VK, Palakurthi S. Recent advances in topical ocular drug delivery. *J Ocul Pharmacol Ther*. 2016;32(2):67-82. DOI: 10.1089/jop.2015.0047
49. Souto EB, Dias-Ferreira J, López-Machado A, Etcheto M, Cano A, Camins Espuny A, et al. Advanced formulation approaches for ocular drug delivery: State-of-the-art and recent patents. *Pharmaceutics*. 2019;11(9):460. DOI: 10.3390/pharmaceutics11090460
50. Fernández-Ferreiro A, Bargiela NF, Varela MS, Martínez MG, Pardo M, Ces AP, et al. Cyclodextrin-polysaccharide-based, in situ-gelled system for ocular antifungal delivery. *Beilstein J Org Chem*. 2014;10(1):2903-11. DOI: 10.3762/bjoc.10.308
51. Díaz-Tomé V, Luaces-Rodríguez A, Silva-Rodríguez J, Blanco-Dorado S, García-Quintanilla L, Ilovo-Taboada J, et al. Ophthalmic econazole hydrogels for the treatment of fungal keratitis. *J Pharm Sci*. 2018;107(5):1342-51. DOI: 10.1016/j.xphs.2017.12.028
52. Fernández-Ferreiro A, González-Barcia M, Gil-Martínez M, Santiago-Varela M, Pardo M, Blanco-Méndez J, et al. Evaluation of the in vitro ocular toxicity of the fortified antibiotic eye drops prepared at the Hospital Pharmacy Departments. *Farm Hosp*. 2016;40(5):352-70. DOI: 10.7399/fh.2016.40.5.10416
53. Luaces-Rodríguez A, González-Barcia M, Blanco-Teijeiro MJ, Gil-Martínez M, González F, Gómez-Ulla F, et al. Review of intraocular pharmacokinetics of anti-infectives commonly used in the treatment of infectious endophthalmitis. *Pharmaceutics*. 2018;10(2):66. DOI: 10.3390/pharmaceutics10020066
54. García-Millán E, Castro-Balado A, Fernández-Ferreiro A, Otero-Espinar FJ. Contact lenses as drug delivery systems. En: Arno F, Rein E. *Recent progress in eye research*. New York: Nova Science Publishers, Inc.; 2017. p. 91-154.
55. Maulvi FA, Soni TG, Shah DO. A review on therapeutic contact lenses for ocular drug delivery. *Drug Delivery*. 2016;23(8):3017-26. DOI: 10.3109/10717544.2016.1138342
56. Álvarez-Lorenzo C, Anguiano-Igea S, Varela-García A, Vivero-López M, Concheiro A. Bioinspired hydrogels for drug-eluting contact lenses. *Acta Biomater*. 2018;5;84:49-62. DOI: 10.1016/j.actbio.2018.11.020
57. Calles JA, Bermúdez J, Vallés E, Allemanni D, Palma S. *Polymers in ophthalmology*. En: Puoci F, editor. *Advanced polymers in medicine* [Internet]. Cham: Springer International Publishing; 2015 [accessed 10/25/2019]; p. 147-76. Available at: [https://doi.org/10.1007/978-3-319-12478-0\\_6](https://doi.org/10.1007/978-3-319-12478-0_6)
58. Ghate D, Edelhauser HF. Ocular drug delivery. *Expert Opin Drug Deliv*. 2006;3(2):275-87. DOI: 10.1517/174252473.2.275
59. Rajasekaran A, Kumaran K, Preetha JP, Karthika K. A comparative review on conventional and advanced ocular drug delivery formulations. *International Journal of PharmTech Research*. 2010;2(1):668-74.
60. Luchs JJ, Nelinson DS, Macy JJ; Group L-0701 S. Efficacy of hydroxypropyl cellulose ophthalmic inserts (LACRISERT) in subsets of patients with dry eye syndrome: findings from a patient registry. *Cornea*. 2010;29(12):1417-27. DOI: 10.1097/ICO.0b013e3181e3f05b
61. Papangkorn K, Truett KR, Vitale AT, Jhaveri C, Scales DK, Foster CS, et al. Novel dexamethasone sodium phosphate treatment (DSP-Visulex) for noninfectious anterior uveitis: A randomized phase I/II clinical trial. *Curr Eye Res*. 2019;44(2):185-93. DOI: 10.1080/02713683.2018.1540707
62. Cagini C, Caricato A, Tosi G, Pascale A, Cesari C, Fiore T. Evaluation of the efficacy and safety of the ophthalmic insert mydrisert in patients undergoing retinal angiography. *Eur J Ophthalmol*. 2014;24(5):728-34. DOI: 10.5301/ejo.5000444
63. Pijs RT, Sonderkamp T, Daube GVV, Krebber R, Hanssen HHL, Nuijts RMM, et al. Studies on a new device for drug delivery to the eye. *Eur J Pharm Biopharm*. 2005;59(2):283-8. DOI: 10.1016/j.ejpb.2004.08.011
64. Pijs RT. *The OphthaCoil: a new vehicle for the delivery of drugs to the eye*. Maastricht: Maastricht University; 2007.
65. Brandt JD, Sall K, DuBiner H, Benza R, Alster Y, Walker G, et al. Six-month intraocular pressure reduction with a topical bimatoprost ocular insert: Results of a phase II randomized controlled study. *Ophthalmology*. 2016;123(8):1685-94. DOI: 10.1016/j.ophtha.2016.04.026
66. García-Quintanilla L, Luaces-Rodríguez A, Gil-Martínez M, Mondelo-García C, Maroñas O, Mangas-Sanjuán V, et al. Pharmacokinetics of intravitreal anti-VEGF drugs in age-related macular degeneration. *Pharmaceutics*. 2019;11(8):365. DOI: 10.3390/pharmaceutics11080365
67. Rezaei KA, Wen JC. Intravitreal injection technique. *MedEdPORTAL*. 2016;12:10502. DOI: 10.15766/mep\_2374-8265.10502
68. Castro-Balado A, Mondelo-García C, González-Barcia M, Zarra-Ferro I, Otero-Espinar FJ, Ruibal-Morell Á, et al. Ocular biodistribution studies using molecular imaging. *Pharmaceutics*. 2019;11(5):237. DOI: 10.3390/pharmaceutics11050237
69. Okada M, Kandasamy R, Chong EW, McGuinness M, Guymer RH. The treat-and-extend injection regimen versus alternate dosing strategies in age-related macular degeneration: A systematic review and meta-analysis. *Am J Ophthalmol*. 2018;192:184-97. DOI: 10.1016/j.ajo.2018.05.026
70. Delplace V, Payne S, Shoichet M. Delivery strategies for treatment of age-related ocular diseases: From a biological understanding to biomaterial solutions. *J Control Release*. 2015;219:652-68. DOI: doi.org/10.1016/j.jconrel.2015.09.065
71. Luaces-Rodríguez A, Mondelo-García C, Zarra-Ferro I, González-Barcia M, Aguiar P, Fernández-Ferreiro A, et al. Intravitreal anti-VEGF drug delivery systems for age-related macular degeneration. *Int J Pharm*. 2020;573:118767. DOI: 10.1016/j.ijpharm.2019.118767
72. Adamson P, Wilde T, Dobrzynski E, Sychterz C, Polsky R, Kurali E, et al. Single ocular injection of a sustained-release anti-VEGF delivers 6 months pharmacokinetics and efficacy in a primate laser CNV model. *J Control Release*. 2016;244:1-13. DOI: 10.1016/j.jconrel.2016.10.026
73. Campochiaro PA, Marcus DM, Awh CC, Regillo C, Adamis AP, Bantseev V, et al. The port delivery system with ranibizumab for neovascular age-related macular degeneration: Results from the randomized phase 2 Ladder clinical trial. *Ophthalmology*. 2019;126(8):1141-54. DOI: 10.1016/j.ophtha.2019.03.036
74. Gaudana R, Ananthula HK, Parenky A, Mitra AK. Ocular drug delivery. *AAPS J*. 2010;12(3):348-60. DOI: 10.1208/s12248-010-9183-3
75. Shah TJ, Conway MD, Peyman GA. Intracameral dexamethasone injection in the treatment of cataract surgery induced inflammation: design, development, and place in therapy. *Clin Ophthalmol*. 2018;12:2223-35. DOI: 10.2147/OPHT.S165722
76. Gutiérrez-Hernández JC, Caffey S, Abdallah W, Calvillo P, González R, Shih J, et al. One-year feasibility study of replenish micropump for intravitreal drug delivery: A pilot study. *Transl Vis Sci Technol*. 2014;3(4):8. DOI: 10.1167/tvst.3.4.1
77. Patel SR, Berezovsky DE, McCarey BE, Zarnitsyn V, Edelhauser HF, Prausnitz MR. Targeted administration into the suprachoroidal space using a microneedle for drug delivery to the posterior segment of the eye. *Invest Ophthalmol Vis Sci*. 2012;53(8):4433-41. DOI: 10.1167/iov5.12-9872
78. Rai UDJP, Young SA, Thrimawithana TR, Abdelkader H, Alani AWG, Pierscionek B, et al. The suprachoroidal pathway: a new drug delivery route to



- the back of the eye. *Drug Discov Today*. 2015;20(4):491-5. DOI: 10.1016/j.drudis.2014.10.010
79. Eljarrat-Binstock E, Pe'er J, Domb AJ. New techniques for drug delivery to the posterior eye segment. *Pharm Res*. 2010;27(4):530-43. DOI: 10.1007/s11095-009-0042-9
80. Cunha-Vaz J. The blood-ocular barriers. *Surv Ophthalmol*. 1979;23(5):279-96. DOI: 10.1016/0039-6257(79)90158-9
81. Vellonen KS, Soini EM, del Amo EM, Urtti A. Prediction of ocular drug distribution from systemic blood circulation. *Mol Pharm*. 2016;13(9):2906-11. DOI: 10.1021/acs.molpharmaceut.5b00729
82. Brockhaus L, Goldblum D, Eggenschwiler L, Zimmerli S, Marzolini C. Revisiting systemic treatment of bacterial endophthalmitis: a review of intravitreal penetration of systemic antibiotics. *Clin Microbiol Infect*. 2019;25(11):1364-9. DOI: 10.1016/j.cmi.2019.01.017
83. Battaglia Parodi M, La Spina C, Berchicci L, Petrucci G, Bandello F. Photosensitizers and photodynamic therapy: Verteporfin. *Dev Ophthalmol*. 2016;55:330-6. DOI: 10.1159/000434704
84. Miguel A, Henriques F, Azevedo LF, Pereira AC. Ophthalmic adverse drug reactions to systemic drugs: a systematic review. *Pharmacoepidemiol Drug Saf*. 2014;23(3):221-33. DOI: 10.1002/pds.3566