



Nutrición Hospitalaria



Trabajo Original

Paciente crítico

Effect of glutamine on liver injuries induced by intestinal ischemia-reperfusion in rats *Efecto de la glutamina en las lesiones hepáticas inducidas por isquemia-reperfusión intestinal en ratas*

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Abstract

Introduction: Intestinal ischemia-reperfusion (I/R) injury may cause cell and tissue damage, reaching also other organs such as the liver. Because of the involvement of free radicals in I/R injury, treatment options with antioxidants have been studied and tested.

Objective: To evaluate the effect of glutamine (Gln) in the liver of animals with intestinal I/R injury.

Methods: We used 20 male Wistar rats divided into four groups: sham-operated (SO); glutamine + sham-operated (G+SO); intestinal ischemia-reperfusion (I/R); glutamine + intestinal ischemia-reperfusion (G+I/R). The superior mesenteric artery was clamped for 30 minutes and reperused for 15 minutes. Gln (25 mg/kg/day) diluted in 1 ml of saline was administered intraperitoneally on the two days before I/R induction.

Results: Levels of serum aspartate aminotransferase (AST), alanine aminotransferase (ALT), and alkaline phosphatase (ALP), lipid peroxidation (LPO) and expressions of interleukin-6 (IL-6) and nuclear factor kappa B (NF-κB) showed a significant reduction in the G+I/R group as compared with the I/R group. The activity of catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPx), and the levels of glutathione (GSH) showed an increase in the G+I/R group as compared with the I/R group.

Conclusion: Pretreatment with Gln reduced oxidative, tissue damage and showed a decrease expression of inflammatory mediators.

Key words:

DNA damage.
Inflammation.
Lipid peroxidation.
Oxidative stress.

Resumen

Introducción: las lesiones por isquemia-reperfusión (I/R) intestinales pueden causar daño celular y tisular, llegando incluso a otros órganos, como el hígado. Debido a la participación de radicales libres en la lesión I/R, se han estudiado y probado opciones de tratamiento con antioxidantes.

Objetivo: evaluar el efecto de la glutamina (Gln) en el hígado de los animales con lesión intestinal I/R.

Métodos: se utilizaron 20 ratas Wistar macho divididas en cuatro grupos: sham-operadas (SO); glutamina + sham-operadas (G+SO); isquemia-reperfusión intestinal (I/R); glutamina + isquemia-reperfusión intestinal (G+I/R). La arteria mesentérica superior se clampó durante 30 minutos y se llevó a cabo la reperusión durante 15 minutos. Se administró Gln (25 mg/kg/día) diluida en 1 ml de solución salina por vía intraperitoneal en los dos días previos a la inducción de la I/R.

Resultados: los niveles de aspartato aminotransferasa (AST), alanina aminotransferasa (ALT) y fosfatasa alcalina (FA), la peroxidación lipídica (POL) y las expresiones de interleucina-6 (IL-6) y factor nuclear kappa B (NF-κB) mostraron una reducción significativa en el grupo G+I/R en comparación con el grupo I/R. La actividad de la catalasa (CAT), la superóxido dismutasa (SOD), glutatión peroxidasa (GPx), y los niveles de glutatión (GSH) mostraron un aumento en el grupo G+I/R en comparación con el grupo I/R.

Conclusión: el tratamiento previo con Gln redujo el daño tisular oxidativo y mostró una disminución de los mediadores inflamatorios.

Palabras clave:

Daño en el DNA.
Inflamación.
Peroxidación lipídica.
Estrés oxidativo.

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INTRODUCTION

Intestinal ischemia is caused by several clinical conditions such as trauma, shock, strangulated hernia, intestinal obstruction, and small intestine transplant (1). The intestinal mucosa is injured during ischemia due to the reduction and/or obstruction of blood flow. After reintroduction of blood flow, these injuries may get worse, leading to intestinal ischemia-reperfusion (I/R) injury (2).

Several mechanisms are involved in intestinal I/R injury, such as neutrophil infiltration, intracellular adhesion molecules (ICAM-1), production of proinflammatory cytokines such as interleukin 1 β and 6 (IL-1 β , IL-6) and tumor necrosis factor alpha (TNF- α), exacerbated generation of reactive oxygen species (ROS), and activation of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) (3,4). This condition may damage the integrity of the intestinal barrier, leading to possible bacterial translocation into the bloodstream, thus triggering the onset of the systemic inflammatory response syndrome (SIRS) and the consequent physiological dysfunction of two or more systems, which is known as multiple organ dysfunction syndrome (MODS) (5-7). The liver is one of the organs that undergoes these changes because of the close relationship between intestinal function and balance of liver function. In case of bowel disease, there is imbalance of the intestinal microbiota, thus causing and worsening liver injuries (8,9).

Although some molecular aspects of the mechanism of action causing intestinal I/R injuries have not been well defined, studies have shown that inflammatory cytokines and increased ROS generation play an important role in the pathogenesis of intestinal and systemic injuries after an I/R event (10,11).

Inflammatory mediators, such as IL-6 and TNF- α , are linked to the activation of neutrophils and tissue damage resulting from I/R. Studies have demonstrated that plasma and tissue levels of these cytokines are high in intestinal I/R and MODS. These mediators may be activated by the NF- κ B, which plays an important role in the control of adaptive immune responses by regulating the expression of these pro-inflammatory genes (7).

The oxidative processes resulting from the exacerbated generation of ROS may cause damage to proteins, lipids, and DNA. These processes are involved in the progression of the intestinal I/R injury and the integrity and function of other organs. Experimental studies have shown the association of oxidative damage with intestinal I/R injury in organs such as lung, liver, and kidney (7,11,12). The antioxidant system may compensate for these damages caused by ROS through enzymatic substances, such as catalase (CAT), superoxide dismutase (SOD), and glutathione peroxidase (GPx), and non-enzymatic substances such as glutathione (GSH). Therefore, recent studies have shown that the administration of antioxidants may contribute to reduce oxidative processes and decrease the local and systemic inflammatory process caused by the intestinal I/R injury (1,4,11,13).

Glutamine (Gln) is a hydrophilic, non-essential amino acid found on the surface of proteins, where it interacts with water. Gln is a precursor for the synthesis of nucleic acids and GSH. In addition, Gln plays a key role in the immune defense of the intestinal mucosal barrier by participating in the formation of immunoglobulins,

especially IgA (14,15). Some mechanisms suggest a protective role of glutamine in the intestinal inflammatory response, namely: increased intestinal antioxidant capacity through the synthesis of GSH, reduced apoptosis, and preserved intestinal integrity, thus reducing the local and systemic inflammatory parameters (14,16).

Experimental studies have demonstrated the antioxidant role of Gln in reducing ROS generation and pro-inflammatory cytokines, as well as in preserving tissue GSH in experimental models of liver I/R (17). Fillmann et al. (2007), Marques et al. (2001), and Zhang et al. (2011) showed that pretreatment with Gln reduced oxidative and tissue damages in the gut, stomach, and liver. These authors also found a reduction in the inflammatory processes, suggesting a protective effect of Gln against free radicals (14,15,18).

In our previous study we demonstrated that pretreatment with Gln reduced intestinal lesions resulting from intestinal ischemia and reperfusion model. We observed a reduction in intestinal tissue damage, decreased oxidative damage as well as reducing inflammation mediators studied (7).

Therefore, the objective of the present study was to investigate the antioxidant and anti-inflammatory effect of Gln based on the evaluation of lipid peroxidation (LPO), DNA damage, activity of CAT, SOD, and GPx, GSH levels, and expression of IL-6 and NF- κ B. Our hypothesis suggests that treatment with Gln could reduce oxidative, cell, and inflammatory damage in the liver tissue induced by an intestinal I/R experimental model.

MATERIALS AND METHODS

ANIMALS AND GLUTAMINE ADMINISTRATION

Our procedures involving animals followed the guidelines of the Research and Ethics Committee of the Research and Graduate Studies Group (GPPG), Hospital de Clínicas de Porto Alegre (HCPA), approved with the protocol number 12-0241, and the European Council Directive regarding animal experimentation (19,20).

We used 20 male Wistar rats. Their mean weight was 300 grams. The animals were divided into four groups: sham operated (SO) (n = 5), glutamine + sham operated (G+SO) (n = 5), intestinal ischemia-reperfusion (I/R) (n = 5), and glutamine + intestinal ischemia-reperfusion (G+I/R) (n = 5). The animals stayed at the Animal Experimentation Unit (UEA) of the Hospital de Clínicas de Porto Alegre. They were kept in plastic boxes lined with wood shavings and measuring 47 x 34 x 18 cm, on a 12:12-hour light/dark cycle (light from 7 am to 7 pm) and temperature between 20 and 25 °C. Water and feed were given *ad libitum*. The animals were intraperitoneally anesthetized with xylazine hydrochloride (8 mg/kg of body weight) and ketamine hydrochloride (92 mg/kg of body weight). Laparotomy and evisceration were performed to identify the superior mesenteric artery in both groups. In groups SO and G+SO (surgery simulation) the artery was not obstructed and in groups I/R and G+I/R mesenteric artery was occluded for 30 minutes using a microsurgical vascular clamp. After intestinal ischemia was achieved, the microsurgical vascular clamp was

removed and the animals were reperfused for 15 more minutes (adapted from Cho et al., 2013) (3). After this period, the liver was removed for histological analysis and other tests. Finally, the animals were killed by exsanguination under deep anesthesia (20).

Gln was administered intraperitoneally at a dose of 25 mg/kg diluted in 1 ml of saline. This dose was given once daily for 48 hours before ischemia induction (14,15).

LIVER FUNCTION TESTS BASED ON ASPARTATE AMINOTRANSFERASE (AST), ALANINE AMINOTRANSFERASE (ALT) AND ALKALINE PHOSPHATASE (ALP)

Levels of serum AST, ALT, and ALP were determined by the Biochemistry Center of the Hospital de Clínicas de Porto Alegre. To determine AST and ALT levels in the plasma, we used the commercially available enzymatic method (Boehringer Mannheim, Germany) and the results were obtained by kinetic measurement at 567 nm. Levels of ALP were measured with an automated analyzer by the enzymatic method, using p-nitrophenyl phosphate (pNPP) substrate plus water, forming p-nitrophenol, a compound measured with maximum absorption of 400 nm.

LIVER HOMOGENATE

Liver tissues were homogenized for 30 seconds in an ULTRA-TURRAX® disperser (IKA-Werke GmbH & Co. KG) at 4 °C using 1.15% KCl (5 ml/g of tissue) and phenylmethane-sulfonylfluoride (PMSF) at a concentration of 100 mM. Next, the homogenates were centrifuged for ten minutes at 3,000 rpm in a refrigerated centrifuge (SORVALL Super T21, Kendro Laboratory Products, USA). The supernatant was removed and stored in microtubes. The samples were stored at -80 °C for later analyses (21).

LPO

The amount of aldehydes generated by LPO is determined by the method that measures the amount of thiobarbituric acid reacting substances (TBARS). Thiobarbituric acid was added to the samples at 0.37%, whereas trichloroacetic acid was added at 15%. The samples were incubated at 100 °C for 15 minutes and centrifuged at 3,000 rpm (1,612 x g) for ten minutes at 4 °C. Absorbency was determined by spectrophotometry at 535 nm (22).

COMET ASSAY

An alkaline comet assay was performed as described by Hartmann and Speit (1999), including some changes suggested by Picada et al. in 2003 (23,24). Blood samples (50 µl) were added to 5 ml of anticoagulant (heparin sodium 25,000 IU/ml solution -

Liquemine®, Roche). Blood cell suspensions (5 µl) were added to 95 ml of 0.75% low melting point agarose (Gilco BRL) and placed on agarose pre-coated microscope slides. After solidification, the slides were placed in lysis buffer (2.5 M NaCl, 100 mM EDTA, 10 mM Tris, pH 10.0). We added 1% Triton X-100 (Sigma) and 10% dimethyl sulfoxide (DMSO) for 48 hours at 4 °C. Next, the slides were subsequently incubated in alkaline buffer (300 mM NaOH and 1 mM EDTA, pH > 13) for 20 minutes at 4 °C. An electric current of 300 mA and 25 V (0.90 V/cm) was applied for 15 minutes to perform DNA electrophoresis. Then the slides were neutralized (0.4 M Tris, pH 7.5), stained with silver, and analyzed by microscope. Images of 100 randomly selected cells (50 cells of each slide) were analyzed from each animal. The cells were also visually classified according to the size of the tail into five categories ranging from undamaged (0) to maximum damage (4), resulting in a single DNA damage score for each animal and for each group. Therefore, the damage index (DI) may range from 0 (completely intact, 100 cells x 0) to 400 (maximum damage, 100 x 4). The damage frequency (%) was calculated based on the number of tailed cells vs tail-less cells.

ACTIVITY OF ANTIOXIDANT ENZYMES: CAT, SOD, GPX AND GSH LEVELS

The analysis of CAT activity was based on the measurement of the reduction of hydrogen peroxide. The activity was detected spectrophotometrically at 240 nm with values expressed as pmol/mg prot (25). The analysis of SOD activity was based on the inhibition of the reaction of epinephrine with superoxide radical, which could be detected spectrophotometrically at 480 nm with values expressed as USOD/mg prot (26). GPx activity was determined based on the consumption of nicotinamide adenine dinucleotide phosphate (NADPH) in the reduction of oxidized glutathione, which could be detected spectrophotometrically at 340 nm within two minutes with the values expressed as nmol/min/mg prot (27). Measurement of GSH levels was detected spectrophotometrically at 412 nm with values expressed as µmol/mg prot, according to the method of Beutler, Duran and Kelly (28).

IMMUNOHISTOCHEMISTRY AND QUANTIFICATION OF THE EXPRESSION OF IL-6 AND NF-κB

The expression of IL-6 and NF-κB in liver tissue was determined by immunohistochemical analysis. Antigen retrieval was performed using buffer at 60 °C, and endogenous peroxidase activity was blocked by incubation in absolute methanol. The slides were incubated with rabbit polyclonal antibody (NF-κB [p65] [SC8008] - Santa Cruz Biotechnology, USA) at 1:100 and (IL-6 [SC1265] - Santa Cruz Biotechnology, USA) at 1:100 overnight at 4 °C. The slides were washed with buffer and incubated with the secondary antibody (anti-mouse IgG-HRP, anti-goat IgG-HRP, Santa Cruz Biotechnology, Santa Cruz, CA, USA) at 1:300 for 30 minutes at room temperature.

The slides were examined by a pathologist, who was unaware of the groups, using a microscope equipped with a digital analysis system including a Zeiss Axioskop 40 microscope (Oberkochen, Germany) connected by a Roper Scientific camera (Media Cybernetics, Rockville, USA) to a computer with an image capture software. The Image-Pro Plus version 4.5 software (Media Cybernetics, Rockville, USA) was used to analyze digital images. The expression was determined by multiplying the mean density of the image by the percentage of positively stained areas (brown-stained areas).

WESTERN BLOT

Western blot analysis was performed on nuclear extracts prepared from liver homogenates as previously described. The supernatant fraction was collected and stored at -80°C in aliquots until use. Lysate proteins were fractionated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to polyvinylidene fluoride (PVDF) membranes. The membranes were then blocked with 5% nonfat dry milk in Tris buffered saline containing 0.05% Tween 20 (TTBS) for one hour at room temperature and probed overnight at 4°C with polyclonal anti-NF- κB (p65) (SC8008/65kDa) (Santa Cruz Biotechnology, Santa Cruz, CA, USA) at 1:200-1,000 dilution with TTBS in 5% nonfat dry milk. Anti- β -actin antibody (A5060/42kDa) (Sigma Aldrich, St Louis, MO, USA) at 1:2,000 dilution with TTBS in 5% nonfat dry milk. After washing with TTBS, the membranes were incubated for one hour at room temperature with secondary anti-mouse IgG-HRP antibody (Santa Cruz Biotechnology, Santa Cruz, CA, USA, 1:4,000). Protein detection was performed by chemiluminescence using a commercial ECL kit (Amersham Pharmacia Biotech, Little Chalfont, UK). The density of the specific bands was quantified using Scion Image software (Scion Corp., Frederick, MD, USA) (29,30).

STATISTICAL ANALYSIS

Data are expressed as means \pm standard error. Statistical significance was calculated using Graphpad InStat, version 3.0 for Windows. We used one-way analysis of variance (ANOVA) and Student-Newman-Keuls for multiple analysis. Analysis of variance with robust standard errors (Welch) was used to investigate the results of the expression of IL-6 and NF- κB between the groups. Results were considered as statistically significant when the significance level was at least 5% ($p < 0.05$).

RESULTS

LIVER FUNCTION TESTS BASED ON AST, ALT AND ALP

The serum levels of AST, ALT and ALP were significantly increased in the I/R group as compared with the SO and G+SO groups, whereas there was a reduction in these levels in the I/R+G

group in comparison with the I/R group (AST*, # $p < 0.001$; ALT*, $p < 0.001$, # $p < 0.05$; ALP*, # $p < 0.01$) (Table I).

LPO AND COMET ASSAY

LPO levels (Fig. 1A) showed a significant increase in the I/R group as compared with the SO and G+SO groups, whereas there was a significant decrease in LPO levels in the G+I/R group as compared with the I/R group (*, # $p < 0.001$).

The results of the comet assay showed an increase in the damage index (Fig. 1B) and damage frequency (Fig. 1C) in the I/R group as compared with the SO and G+SO groups, whereas there was a reduction in these parameters in the G+I/R group in comparison with the I/R (* $p < 0.01$, # $p < 0.05$).

ACTIVITY OF CAT, SOD, GPX AND GSH LEVELS

The activities of CAT (Fig. 2A), SOD (Fig. 2B) and GPx (Fig. 2C) decreased significantly in the I/R group when compared with the SO and G+SO groups, whereas these activities increased in the G+I/R group as compared with the I/R group (* $p < 0.01$, # $p < 0.05$).

The GSH levels (Fig. 3) showed a significant decrease in the I/R group as compared with the SO and G+SO groups, whereas there was a significant increase in the G+I/R group as compared with the I/R group (* $p < 0.01$, # $p < 0.05$).

IMMUNOHISTOCHEMISTRY AND QUANTIFICATION OF THE EXPRESSION OF IL-6 AND NF- κB

The animals of the I/R group showed strong positive staining for IL-6 (Fig. 4) and NF- κB (Fig. 6), which was visible because of the brown staining.

Table I. Effect of glutamine administration on the levels of enzymes of liver cell integrity in the blood of animals subjected to the intestinal ischemia-reperfusion model

Groups	AST	ALT	ALp
SO	101.60 \pm 2.66	51.40 \pm 3.24	160.20 \pm 11.57
G+SO	105.33 \pm 9.58	56.00 \pm 4.97	159.60 \pm 8.76
I/R	291.00 \pm 23.57*	91.80 \pm 8.62*	187.80 \pm 4.82*
G+I/R	145.00 \pm 12.44#	76.00 \pm 3.34#	155.33 \pm 4.75#

Values are expressed as mean \pm SEM. *Significant difference between the I/R group and the SO and G+SO groups. #Significant difference between the G+I/R group and the I/R group. Sham operated (SO), glutamine + sham operated (G+SO), intestinal ischemia-reperfusion (I/R), glutamine + intestinal ischemia-reperfusion (G+I/R). AST - *, # $p < 0.001$; ALT - * $p < 0.001$, # $p < 0.05$; ALP - *, # $p < 0.01$.

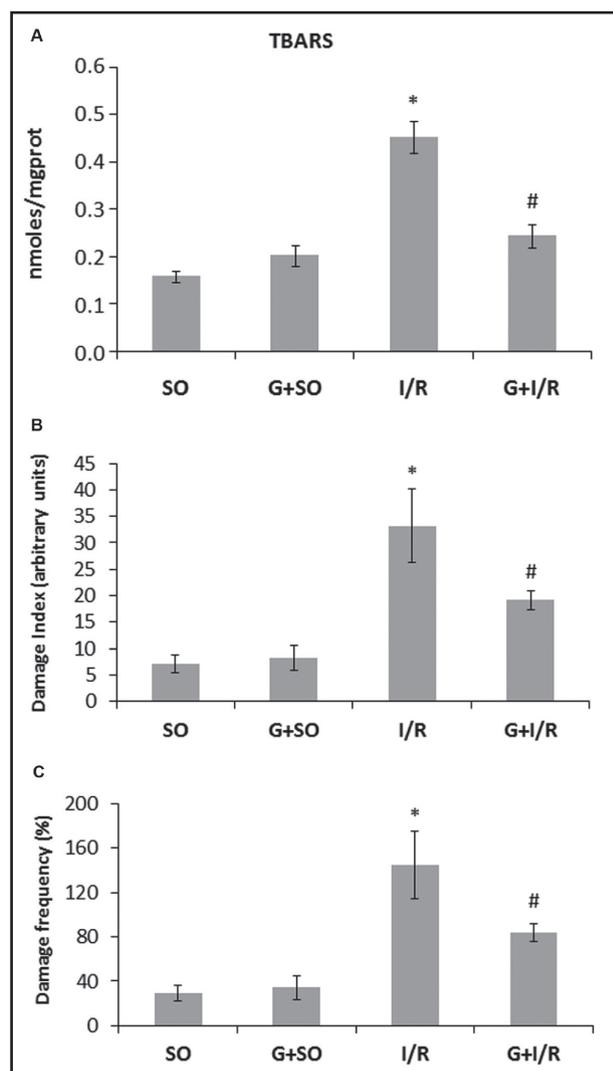


Figure 1.

Effect of glutamine administration on levels of lipid peroxidation in the liver tissue and DNA damage in the blood of animals subjected to the intestinal ischemia-reperfusion model. A. TBARS. B. Damage index. C. Damage frequency. Values are expressed as mean \pm SEM. A. *Significant difference between the I/R group and the SO and G+SO groups ($p < 0.001$). #Significant difference between the G+I/R group and the I/R group. ($p < 0.001$). B and C. *Significant difference between the I/R group and the SO and G+SO groups ($p < 0.01$). #Significant difference between the G+I/R group and the I/R group ($p < 0.05$). Sham operated (SO), glutamine + sham operated (G+SO), intestinal ischemia-reperfusion (I/R), glutamine + intestinal ischemia-reperfusion (G+I/R).

The SO and G+SO groups showed no positive staining. Pre-treatment with Gln reduced the staining for IL-6 and NF- κ B in the G+I/R group.

Similarly, the quantification of the expression of IL-6 (Fig. 4) and NF- κ B (Fig. 5) showed an increase in the expression of inflammatory mediators in the I/R group as compared with the SO and G+SO groups and a significant reduction in the G+I/R as compared with the I/R group (* $p < 0.001$, # $p < 0.05$). All images were magnified 400x.

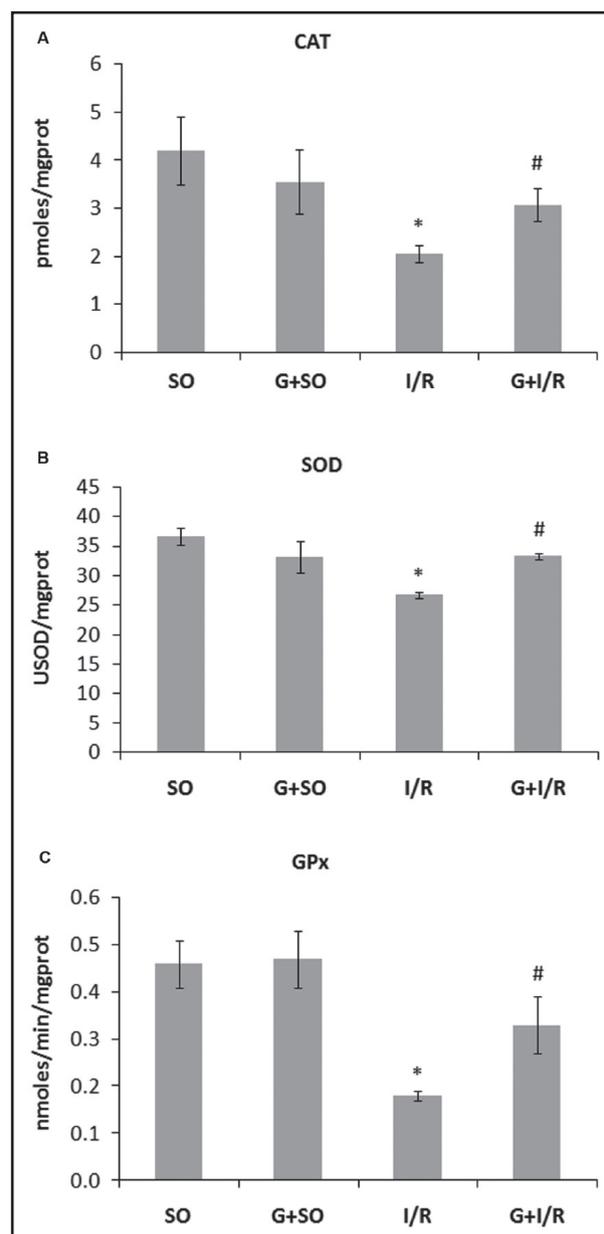


Figure 2.

Effect of glutamine administration on the CAT, SOD and GPx activity in the liver of animals subjected to the intestinal ischemia-reperfusion model. A. CAT. B. SOD. C. GPx. Values are expressed as mean \pm SEM. *Significant difference between the I/R group and the SO and G+SO groups ($p < 0.01$). #Significant difference between the G+I/R group and the I/R group ($p < 0.05$). Sham operated (SO), glutamine + sham operated (G+SO), intestinal ischemia-reperfusion (I/R), glutamine + intestinal ischemia-reperfusion (G+I/R).

WESTERN BLOT NF- κ B (p65)

The expression of nuclear factor NF- κ B (p65) (Fig. 6) showed an increase in the I/R group as compared with the SO and G+SO groups, and a significant decrease in the G+I/R as compared with the I/R group (* $p < 0.01$, # $p < 0.05$).

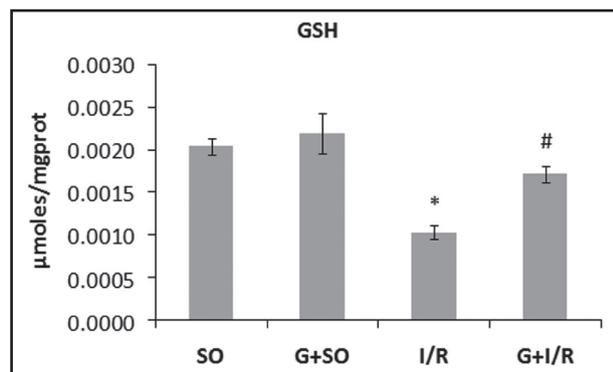


Figure 3.

Effect of glutamine administration on GSH levels in the liver of animals subjected to the intestinal ischemia-reperfusion model. Values are expressed as mean ± SEM. *Significant difference between the I/R group and the SO and G+SO groups ($p < 0.01$). #Significant difference between the G+I/R group and the I/R group ($p < 0.05$). Sham operated (SO), glutamine + sham operated (G+SO), intestinal ischemia-reperfusion (I/R), glutamine + intestinal ischemia-reperfusion (G+I/R).

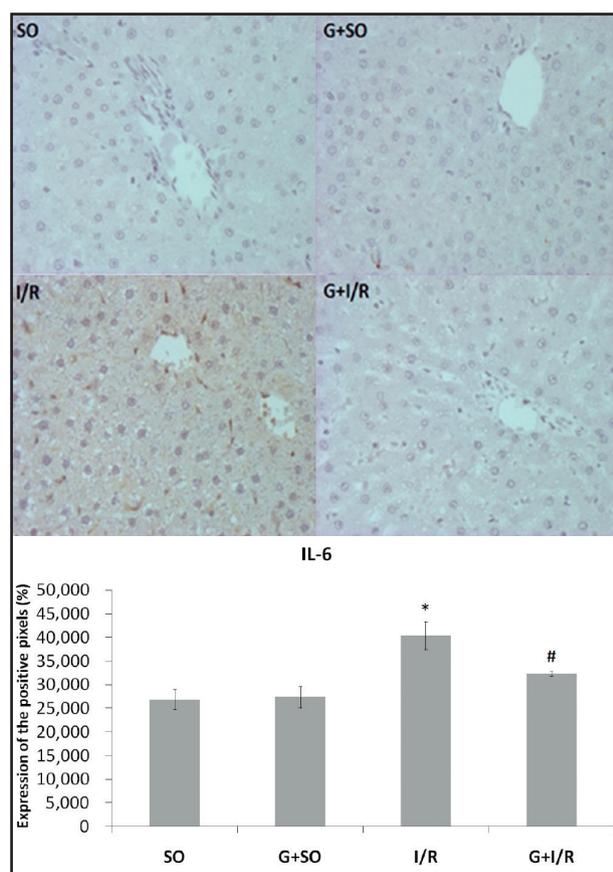


Figure 4.

Effect of glutamine administration on the immunohistochemistry and expression of IL-6 in the liver of animals subjected to the intestinal ischemia-reperfusion model. Values are expressed as mean ± SEM. *Significant difference between the I/R group and the SO and G+SO groups ($p < 0.001$). #Significant difference between the G+I/R group and the I/R group ($p < 0.001$). Sham operated (SO), glutamine + sham operated (G+SO), intestinal ischemia-reperfusion (I/R), glutamine + intestinal ischemia-reperfusion (G+I/R).

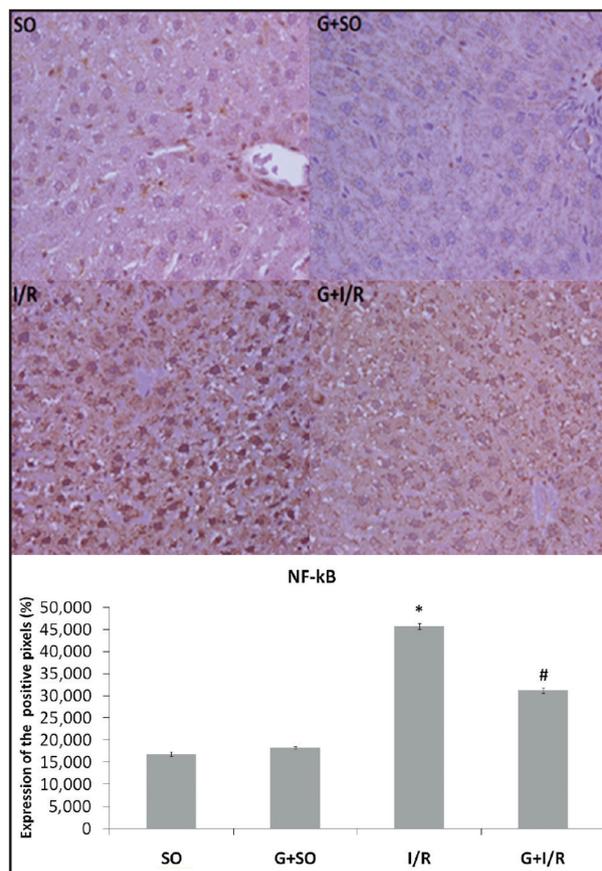


Figure 5.

Effect of glutamine administration on the immunohistochemistry and expression of NF- κ B (p65) in the liver of animals subjected to the intestinal ischemia-reperfusion model. Values are expressed as mean ± SEM. *Significant difference between the I/R group and the SO and G+SO groups ($p < 0.001$). #Significant difference between the G+I/R group and the I/R group ($p < 0.001$). Sham operated (SO), glutamine + sham operated (G+SO), intestinal ischemia-reperfusion (I/R), glutamine + intestinal ischemia-reperfusion (G+I/R).

DISCUSSION

Intestinal I/R is often associated with small intestine transplant, strangulated hernia, aortic aneurysm surgery, and necrotizing enterocolitis, with high rates of mortality and morbidity in recent decades. Some studies suggest that this injury may trigger the involvement of other organs and it may be the main mechanism in the pathogenesis of MODS (6,13).

The small intestine is susceptible to ischemic injury, and the main factors involved are adhesion molecules, nitric oxide, pro-inflammatory cytokines, and reactive oxygen species. Some studies suggest that exacerbated generation of free radicals is one of the major causes of local and systemic injuries (8,9,12).

The gut-liver axis plays a key role in the deleterious effects induced by intestinal I/R because the intestinal bloodstream is connected to liver vascularization. Intestinal I/R leads to the production and subsequent release of deleterious substances, such as ROS and inflammatory mediators, which reach the liver through the portal vein causing severe liver injuries (31-33).

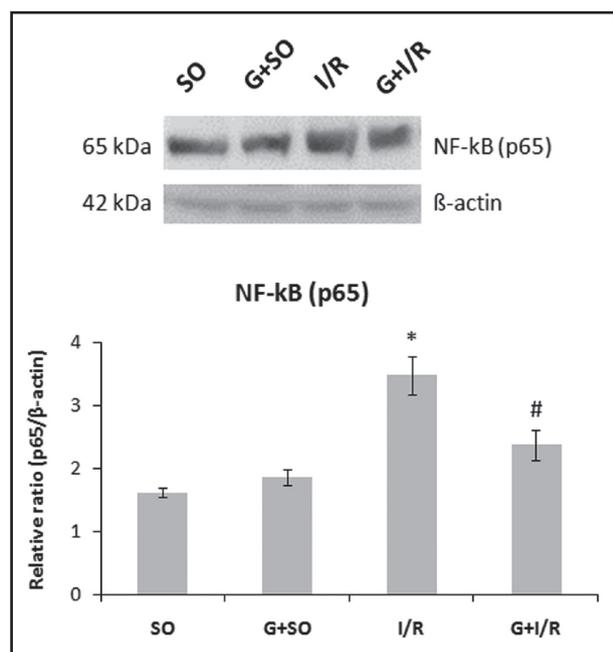


Figure 6.

Effect of glutamine administration on the expression of NF- κ B (p65) by western blot analysis in the liver of animals subjected to the intestinal ischemia-reperfusion model. Values are expressed as mean \pm SEM. *Significant difference between the I/R group and the SO and G+SO groups ($p < 0.01$). #Significant difference between the G+I/R group and the I/R group ($p < 0.05$). Sham operated (SO), glutamine + sham operated (G+SO), intestinal ischemia-reperfusion (I/R), glutamine + intestinal ischemia-reperfusion (G+I/R).

Gln has been studied as a possible treatment option for injuries associated with ROS and inflammatory substances because it plays an antioxidant and anti-inflammatory role, and may reduce these oxidative and inflammatory damages (7,18).

In our study, the liver damage was also evaluated based on the function of the hepatic enzymes AST, ALT, and ALP. Our results showed an increase in these enzymes in the I/R group, and pretreatment with Gln reduced these levels. Inan et al. (2012) found an increase in the activity of AST and ALT in animals subjected to an ischemia-reperfusion model. The administration of sildenafil reduced the levels of these enzymes (32).

Some studies have linked the involvement of oxidative processes, such as LPO and DNA damage (34), oxidative damage, inflammatory process and DNA damage, to the progression of tissue and cell injuries caused by intestinal I/R in other organs (7,10,12). We measured the levels of LPO by TBARS and DNA damage by comet assay. We found an increase in LPO levels and DNA damage index and frequency in the I/R group; however, there was a significant decrease in terms of oxidative damage in the group pretreated with Gln, possibly due to its antioxidant activity.

Our results corroborate other studies showing similar results in the reduction of LPO levels when there is use of a substance with antioxidant effect. Shafik (2012) found an increase in the serum levels of malondialdehyde of animals with I/R. After the administration of febusostat, these levels were reduced, thus

demonstrating a possible antioxidant action of this substance (35). Cámara-Lemarroy et al. (2011) evaluated the possible antioxidant and anti-inflammatory effects of triflusal, S-adenosylmethionine and dextromethorphan, suggesting that the administration of these substances reduced the serum levels of malondialdehyde of animals with I/R (36). Oliveira et al. (2013) showed that glutamine reduced DNA damage in animals receiving cisplatin chemotherapy, a medication that affects the DNA synthesis, suggesting a protective effect of glutamine against damage of DNA bases (37,38).

The antioxidant system is a compensation mechanism for the oxidation process involving enzymatic substances, such as catalase, superoxide dismutase and glutathione peroxidase, and non-enzymatic substances, such as GSH. These substances play an important role against oxidative damage (10,11). In our study, we found a significant reduction in the activity of CAT, SOD and GPx, as well as in the levels of GSH in the I/R group, whereas there was a significant increase in the antioxidant parameters in the group pretreated with Gln.

Zhang et al. (2011) found that pretreatment with Gln was able to enhance the antioxidant capacity in an ischemia-reperfusion model. The animals treated with different doses of Gln showed an increase in SOD activity, suggesting a protective role of Gln against the oxidative damage caused by hepatic I/R-mediated liver injury (18).

Similar results were found in the study by Zhao et al. (2010), where a reduction in the activity of SOD and GPx and in the levels of glutathione in the liver of animals subjected to an intestinal I/R model was observed. The administration of sulforaphane compound (SFN), which is present in several plants, was able to increase the antioxidant capacity of the liver, suggesting a protective role against oxidative damage (39).

Intestinal I/R may lead to increased generation of free radicals and to the consequent production of pro-inflammatory cytokines (IL-1 β , IL-6, TNF- α) that may be mediated by NF- κ B (5,40). We measured the expression of IL-6 and NF- κ B by immunohistochemistry and western blot analysis in the liver. Increased expression of inflammatory markers (IL-6 and NF- κ B) was observed in the I/R group, whereas it was significantly reduced in the group pretreated with Gln, suggesting a possible anti-inflammatory effect of this amino acid.

Cho et al. (2013) evaluated the systemic inflammatory response based on the IL-6 plasma level of animals subjected to an intestinal I/R model. They found an increase in these levels in the intestinal I/R group; conversely, these levels were reduced with the administration of remifentanyl, suggesting an inhibitory action on NF- κ B activity thus reducing the production of IL-6 (3). Ma et al. (2014) found a positive expression of NF- κ B in the liver tissue of the intestinal I/R group, whereas, when protocatechuic acid was administered, a polyphenolic compound with antioxidant and anti-inflammatory effect, there was a reduction in the expression of this inflammatory mediator (11). Similar results were found by Yao et al. (2009), showing that the treatment with carnosol, a compound with antioxidant effect, inhibited the expression of NF- κ B, thereby reducing intestinal I/R-induced liver injuries (40).

In conclusion, our findings suggest that Gln inhibits LPO, restoring the activity of antioxidant enzymes, as well as reducing the

IL-6 and NF- κ B expression, possibly due to its antioxidant and anti-inflammatory effect. Therefore, these results support the use of Gln to treat liver injuries caused by intestinal ischemia-reperfusion. Nevertheless, further studies should be conducted to validate its use in clinical practice.

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