



Trabajo Original

Obesidad y síndrome metabólico

Association of resistin (rs3138167) gene polymorphism with metabolic response after a hypocaloric Mediterranean diet

Asociación del polimorfismo del gen de la resistina (rs3138167) con la respuesta metabólica tras una dieta mediterránea hipocalórica

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Abstract

Background: the single nucleotide polymorphism (SNP) (rs3138167) is a polymorphism that has been associated with metabolic disorder in obese subjects and its effect on the metabolic response after a dietary intervention has not been evaluated.

Objective: our aim was to analyze the effects of the rs3138167 on metabolic changes secondary to weight loss with a hypocaloric diet with a Mediterranean pattern.

Method: one thousand and eight Caucasian obese patients were evaluated. Before and after 12 weeks on a hypocaloric diet with Mediterranean pattern, an anthropometric evaluation and a biochemical analysis were performed. The statistical analysis was performed as a dominant model (CC vs. CT + TT).

Results: the values of insulin, HOMA-IR and resistin were higher in T allele carriers than non-T allele carriers in pre- and post-intervention time. In non-T allele carriers, resistin, insulin, HOMA-IR, triglycerides and C-reactive protein levels decreased. The improvement was statistically superior in non-T allele carriers; resistin (-1.2 ± 0.2 ng/dl; $p = 0.02$), triglycerides (-18.3 ± 4.3 mg/dl; $p = 0.02$), C-reactive protein (-2.6 ± 0.3 mg/dl; $p = 0.02$), insulin (-4.4 ± 1.9 mU/l; $p = 0.02$) and HOMA-IR (-2.1 ± 0.7 ; $p = 0.03$).

Conclusion: we report an association of rs3138167 with a worse metabolic response (insulin, HOMA-IR, triglyceride and C-reactive protein) in T allele carriers after weight loss with a hypocaloric diet with Mediterranean pattern.

Keywords:

Mediterranean diet.
Obesity. Resistin.
rs3138167.

Resumen

Antecedentes: el polimorfismo de nucleótido único (SNP) (rs3138167) se ha asociado con trastorno metabólico en sujetos obesos y no se ha evaluado su efecto sobre la respuesta metabólica después de una intervención dietética.

Objetivo: nuestro objetivo fue analizar los efectos del polimorfismo rs3138167 sobre los cambios metabólicos secundarios a la pérdida de peso con una dieta hipocalórica de patrón mediterráneo.

Métodos: se evaluaron 1.008 pacientes caucásicos con obesidad. Antes y tras 12 semanas de dieta hipocalórica con patrón mediterráneo, se realizaron una evaluación antropométrica y un análisis bioquímico. El análisis estadístico se realizó como un modelo dominante (CC vs. CT + TT).

Resultados: los valores de insulina, HOMA-IR y resistina fueron más elevados en los portadores del alelo T, tanto antes como después de la intervención dietética. En los no portadores del alelo T, los niveles de resistina, insulina, HOMA-IR, triglicéridos y proteína C reactiva disminuyeron. Las mejoras fueron estadísticamente significativas, de manera superior en los no portadores del alelo T; resistina ($-1,2 \pm 0,2$ ng/dl; $p = 0,02$), triglicéridos ($-18,3 \pm 4,3$ mg/dl; $p = 0,02$), proteína C reactiva ($-2,6 \pm 0,3$ mg/dl; $p = 0,02$), insulina ($-4,4 \pm 1,9$ mU/l; $p = 0,02$) y HOMA-IR ($-2,1 \pm 0,7$; $p = 0,03$).

Conclusión: describimos una asociación del rs3138167 con una peor respuesta metabólica en los portadores del alelo T (insulina, HOMA-IR, triglicéridos y proteína C reactiva) tras la pérdida de peso con una dieta hipocalórica de patrón mediterráneo.

Palabras clave:

Dieta mediterránea.
Obesidad. Resistina.
rs3138167.

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INTRODUCTION

Resistin is a well-known adipokine that it is related initially to insulin resistance in animal models (1). It is secreted by adipocytes and macrophages in adipose tissue. Adipose tissue has a role of energy storage but it is also an endocrine organ when synthesizing many adipokines. These adipokines have important roles in metabolism, insulin sensitivity, inflammatory status, satiety and appetite (2), and one of them is the above-mentioned resistin. Circulating resistin levels have been associated with increased central obesity (3), metabolic syndrome (4) and type 2 diabetes mellitus (5). However, other studies have failed to demonstrate these associations (6).

Heritability of resistin levels has been described (7), and a high percentage of the variation in circulating resistin levels can be explained by genetic factors. The gene encoding resistin (RETN) is located in chromosome 19p13.2, and some genetic variants have been described in this locus (8,9). For example, single nucleotide polymorphisms (SNPs) of this gene and in decorin gene (DC) have been reported to increase diabetes mellitus type 2 susceptibility by increasing resistin levels (10,11). One of these SNPs is rs3138167 and it has been poorly evaluated. The SNP 5' UTR C/T intron variant (rs3138167) is a polymorphism associated with elevated resistin levels (12). Despite this previous data in cross-sectional studies, there are no investigations in the literature that evaluate the effect of rs3138167 on metabolic changes after weight loss with a diet. One of the dietary patterns with the greatest beneficial effect on biochemical parameters after weight loss is the Mediterranean diet pattern (13). The Mediterranean diet pattern has demonstrated a lot of cardiometabolic improvements such as improvement of insulin resistance or lipid profile (14). Some studies with other SNPs have shown different metabolic responses to dietary interventions (15), and even after bariatric surgery (16).

Our aim was to analyze the effects of the rs3138167 polymorphism on metabolic changes secondary to weight loss after a hypocaloric diet with a Mediterranean pattern.

PATIENTS AND METHODS

SUBJECTS

A sample of 1,029 obese (body mass index [BMI] ≥ 30 kg/m²) non-diabetic Caucasian adult outpatients was recruited and a total of 1,011 subjects were analyzed (Fig. 1). These patients were enrolled in a prospective way with a consecutive method of sampling among patients from Primary Care physicians with obesity. This design was realized according to the Declaration of Helsinki and all procedures were passed by the Ethics Central Committee of Hospital Clínico Universitario de Valladolid (HCU-Va). All participants signed an informed consent to a protocol approved by the local ethical review boards.

For the inclusion of these patients, the following criteria were used: age over 18 years, BMI over 30 and absence of a diet during the three months previous to the study. The following criteria were used as exclusion criteria: cardiovascular events during the previous 12 months, history of hypertension or dyslipidemia or diabetes mellitus, as well as the use of metformin, sulphonylurea, dipeptidyl type IV inhibitor drugs, thiazolidinediones, insulin, glucocorticoids, angiotensin receptor blockers, angiotensin converting enzyme inhibitors, psychoactive medications, statins and other lipid drugs.

PROCEDURES

Fasting (12 hours) venous blood samples were obtained by venipuncture and collected in Vacutainer™ tubes. Basal fast-

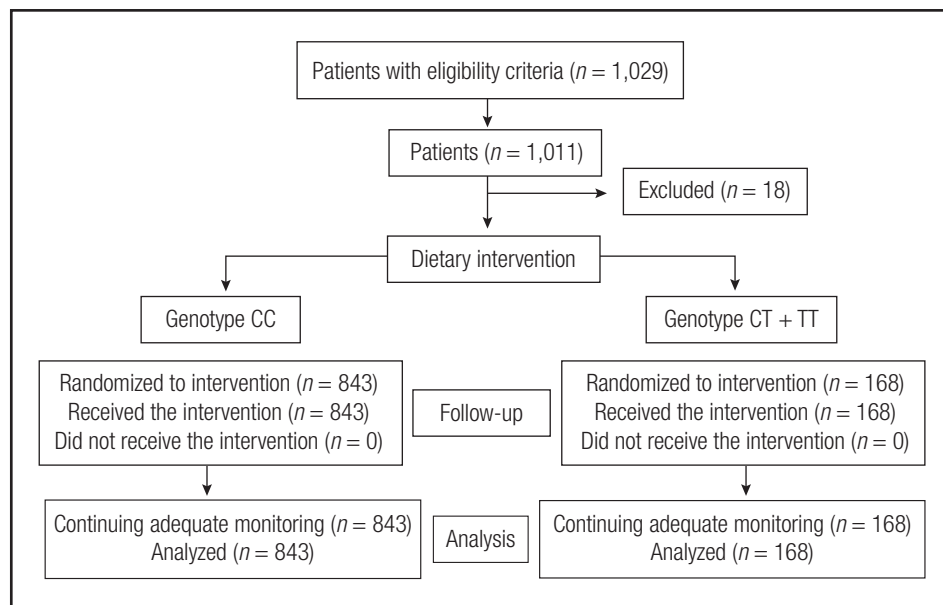


Figure 1.
Flow chart of patients.

ing glucose, C-reactive protein (CRP), insulin, insulin resistance (HOMA-IR), lipid profile (total cholesterol, LDL-cholesterol, HDL-cholesterol, plasma triglycerides concentration) and circulating adipokine levels (leptin, adiponectin and resistin) were determined within the basal time of the trial and repeated after 12 weeks of follow-up. In the two times of the study previously commented, the following parameters were determined: fat mass with impedance, weight and blood pressure. rs3138167 variant of RETN gene was evaluated at basal time.

GENOTYPING OF rs3138167 GENE POLYMORPHISM

Genomic DNA was obtained from peripheral blood leukocytes with the commercial Veneo Extract DNA kit (Biorad, Los Angeles, California, United States) according to the manufacturer's protocol. The real-time polymerase chain reaction (PCR) was carried out with 50 ng of this genomic DNA, 0.5 μ l of 100 μ M of each oligonucleotide primer (primer forward: 5'-ACGTTGGATGTTA-AGAAGAGTAGCACTGCC-3' and reverse 5'-ACGTTGGATGTCTCTGTGCTCACTGTATTG-3'). The DNA was denatured at 95 °C for three minutes; 45 denaturation cycles were performed at 95 °C for 15 seconds, and subsequently, at 59.3 °C for 45 seconds. PCR was performed in a final volume of 25 μ l containing 12.5 μ l of IQ™ Supermix (Bio-Rad®, Hercules, California, United States) with TAq DNA polymerase. If both strands grew in the sample, this sample was classified as heterozygous. If only one strand grew in a sample, this sample was classified as homozygous. The thermal cycler software classifies each subject as homozygous wild type (CC), heterozygous (CT), and homozygous mutant (TT). Moreover, a negative control and control samples representing all genotypes were included in each reaction. Hardy-Weinberg equilibrium was determined with a statistical test (Chi-squared). The variant of RETN gene was in Hardy-Weinberg equilibrium ($p = 0.43$).

LABORATORY DETERMINATIONS

Serum biochemistry analysis for glucose, insulin, CRP, total cholesterol, HDL-cholesterol, and triglyceride levels was measured using the COBAS INTEGRA® 400 analyzer (Roche Diagnostic, Basel, Switzerland). LDL cholesterol was calculated using Friedewald formula (LDL cholesterol = total cholesterol - HDL cholesterol - triglycerides / 5) (17). Based on glucose and insulin levels, homeostasis model assessment for insulin resistance (HOMA-IR) was obtained using the next equation (glucose x insulin / 22.5) (18). Finally, all adipokine levels were determined by enzyme-linked immunosorbent assay (ELISA): resistin (Biovendor Laboratory, Inc., Brno, Czech Republic) with a sensitivity of 0.2 ng/ml with a normal range of 4-12 ng/ml (19), leptin (Diagnostic Systems Laboratories, Inc., Texas, United States) with a sensitivity of 0.05 ng/ml and a normal range of 10-100 ng/ml (20) and adiponectin (R&D systems, Inc., Minneapolis, United States) with a sensitivity of 0.246 ng/ml and a normal range of 8.65-21.43 ng/ml.

BLOOD PRESSURE AND ANTHROPOMETRIC PARAMETERS

Blood pressure was measured three times after a ten-minute rest time with a mercury sphygmomanometer (Omrom, Los Angeles, California, United States) and the results were averaged. Body weight was measured with a scale with a precision of 100 g (Omrom, Los Angeles, California, United States), and BMI was calculated as weight (kg) / height (m^2), classifying as obese patients with a BMI greater than 30 kg/ m^2 . The waist circumference (WC) was also measured with a measuring tape (Type SECA, SECA, Birmingham, United Kingdom) (narrowest diameter between the xiphoid process and the iliac crest). Total fat mass was obtained by impedance with an accuracy of 5 g (EFG BIA 101 Anniversary, Akern, Italy) (21). The following formula was used: $(0.756 \text{ height}^2 / \text{resistance}) + (0.110 \times \text{body mass}) + (0.107 \times \text{reactance}) - 5.463$.

DIETARY CHANGE

During this interventional study (12 weeks), subjects received individualized counseling on a hypocaloric diet with a Mediterranean profile. This caloric intake was calculated by subtracting 500 calories from the caloric intake obtained with the Harris-Benedict formula in our obese population ($2,039.4 \pm 93.7$ kcal per day). The ratio of nutrients in the diet was as follows: 25 % from lipids, 23 % from proteins and the main nutrient was 52 % from carbohydrates. The range of fats was: 50.7 % of monounsaturated dietary fats, 38.5 % of saturated dietary fats and 11.8 % of polyunsaturated dietary fats. The diet contained the following foods: extra-virgin olive oil (an amount of 30 ml/day) (OliDuero, Matarromera, S.L.), three portions of fish per each week, three servings of nuts per week, and 4-5 portions of vegetables and fresh fruits each day. The monitoring of the dietary intervention was performed each two weeks by a dietitian. All subjects received briefing to impress their intakes for three different days. Records were evaluated by a registered dietitian with a computer program (22). The recommended physical exercise program consisted of aerobic exercise at least three times per week (60 minutes each, reaching a total of 180 minutes per week) and the patient recorded it with a self-reported questionnaire.

STATISTICAL ANALYSIS

Sample size was realized to detect differences over 1 ng/ml in circulating resistin levels after diets with 90 % power and 5 % significance ($n = 1,000$). The Kolmogorov-Smirnov test was used to determine variable distribution. The data were reported as average \pm standard deviation. Numerical variables with normal distribution were analyzed with a two-tailed Student's t-test. Categorical variables were evaluated with the Chi-squared test, with Yates correction as necessary. Non-parametric variables were analyzed with the Mann-Whitney U test. The differences in anthropometric and

biochemical variables between the genotype groups were tested with analysis of the covariance (ANCOVA) adjusted age and sex. The statistical analysis was performed for the combined CT and TT genotypes as a group (risk genotype) and CC genotype as second group (wild genotype), in a dominant model. A *p*-value < 0.05 was considered as statistically significant. All analyses were carried out using SPSS version 23.0 (Illinois, United States).

RESULTS

One thousand and twenty-nine patients signed an informed consent and were enrolled in the dietary intervention trial. One thousand and eight patients received dietary intervention and completed the 12-week follow-up period (Fig. 1). The mean age was 47.3 ± 4.3 years and the mean BMI was 36.2 ± 2.3 , with 271 males (26.9 %) and 737 females (73.1 %). Eight hundred and forty-three patients (83.4 %) had the genotype CC (major allele group) and 168 (16.6 %) patients had the next genotypes groups; CT (165 patients, 16.3 %) or TT (three patients, 0.3 %) (minor allele group). Hardy-Weinberg equilibrium was assessed with the Chi-squared test to compare our expected

and observed counts. This genetic variant was in Hardy-Weinberg equilibrium (*p* = 0.43).

No differences were detected in average ages between genotype groups (major allele group: 47.8 ± 2.9 years vs minor allele group: 47.1 ± 4.3 years; ns). Sex distribution was similar in both genotype groups, males (27.6 % vs 23.4 %) and females (72.4 % vs 76.6 %), too. Patients followed the dietary recommendations and in both genotype groups patients reached the targets. The total caloric amount was similar in both genotype groups (CC vs CT + TT) ($1,543.1 \pm 192.2$ vs $1,496.1 \pm 210.1$ calories/day; ns). The percentage of macronutrient in both groups (CC vs CT + TT) was similar: carbohydrates = 35.8 ± 3.1 vs 36.2 ± 2.7 % (*p* = 0.21); fats = 41.0 ± 3.1 vs 41.1 ± 2.8 % (*p* = 0.22); and proteins = 23.2 ± 2.3 vs 22.7 ± 1.9 %; (*p* = 0.13), too. The distribution of dietary fats in both genotype groups (CC vs CT + TT) was similar: monounsaturated fats = 60.3 ± 4.0 vs 59.9 ± 4.4 % (*p* = 0.42); saturated fats = 24.7 ± 3.0 vs 25.1 ± 2.8 % (*p* = 0.30); and polyunsaturated fats = 16.0 ± 1.0 vs 15.0 ± 1.3 % (*p* = 0.13), too.

The changes in adiposity parameters and levels of blood pressure are reported in table I. After the hypocaloric diet with a Mediterranean pattern, the next parameters decreased: weight, BMI,

Table I. Basal and post-intervention anthropometric parameters of obesity and blood pressure measurement (mean ± SD)

Parameters	CC (n = 843)		CT + TT (n = 168)		p values - Time CC - Basal genotype - Time CT + TT - 12 weeks genotype
	Basal	12 weeks	Basal	12 weeks	
BMI	36.2 ± 2.0	$35.2 \pm 2.1^*$	36.1 ± 1.4	$35.3 \pm 1.2^*$	<i>p</i> = 0.01 <i>p</i> = 0.34 <i>p</i> = 0.01 <i>p</i> = 0.38
Weight (kg)	93.9 ± 3.6	$89.8 \pm 3.1^\dagger$	92.9 ± 2.2	$88.9 \pm 1.3^\dagger$	<i>p</i> = 0.02 <i>p</i> = 0.41 <i>p</i> = 0.01 <i>p</i> = 0.47
Fat mass (kg)	38.7 ± 2.2	$35.6 \pm 2.1^\dagger$	37.8 ± 1.2	$34.9 \pm 1.9^\dagger$	<i>p</i> = 0.03 <i>p</i> = 0.21 <i>p</i> = 0.02 <i>p</i> = 0.39
WC (cm)	111.4 ± 5.1	$102.8^\ddagger \pm 6.1\§$	109.9 ± 4.1	$102.3 \pm 2.1^\§$	<i>p</i> = 0.03 <i>p</i> = 0.47 <i>p</i> = 0.03 <i>p</i> = 0.51
SBP (mmHg)	128.1 ± 5.2	$123.7 \pm 4.1^\ddagger$	128.9 ± 4.1	$122.7 \pm 3.8^\ddagger$	<i>p</i> = 0.01 <i>p</i> = 0.39 <i>p</i> = 0.02 <i>p</i> = 0.53

(Continues on next page)

Table I (cont.). Basal and post-intervention anthropometric parameters of obesity and blood pressure measurement (mean \pm SD)

Parameters	CC (n = 843)		CT + TT (n = 168)		p values – Time CC – Basal genotype – Time CT + TT – 12 weeks genotype
	Basal	12 weeks	Basal	12 weeks	
DBP (mmHg)	83.1 \pm 4.1	82.0 \pm 3.1	83.9 \pm 3.1	81.4 \pm 4.2	p = 0.51 p = 0.69 p = 0.62 p = 0.53

BMI: body mass index; DBP: diastolic blood pressure; SBP: systolic blood pressure; WC: waist circumference. Statistical differences $p < 0.05$, in each genotype group (*BMI, †weight, ‡fat mass, §WC, ¶SBP). First p, significance of dietary intervention after 12 weeks in CC genotype; second p, significance between CC genotypes vs CT + TT baseline values; third p, significance of dietary intervention after 12 weeks in CT + TT genotype; and fourth p, significance between CC genotypes vs CT + TT post-treatment.

fat mass, systolic blood pressure and WC. These improvements were similar in both genotype groups, without statistical differences. In the CC group, the decrease in weight was -4.1 ± 1.1 kg (decrease in T allele carriers: -4.0 ± 1.2 kg; $p = 0.43$), BMI -1.0 ± 0.3 kg/m² (decrease in T allele carriers: -0.9 ± 0.3 kg/m²; $p = 0.41$), fat mass -3.1 ± 1.0 kg (decrease in T allele carriers: -3.0 ± 1.0 kg; $p = 0.34$) and WC -8.6 ± 2.1 cm (decrease in T allele carriers: -7.6 ± 2.1 cm; $p = 0.31$). In non-T allele carriers, the decrease in systolic blood pressure was -4.4 ± 1.9 mmHg (decrease in T allele carriers: -5.1 ± 2.1 mmHg; $p = 0.28$). Therefore, the improvement of all the previously mentioned parameters was similar in carriers and non-carriers of the T allele. No differences were detected in diastolic blood pressure after dietary intervention. Finally, no differences were detected among basal and post-treatment values of anthropometric parameters between both genotype groups CC vs CT/TT. An additional analysis was carried out in anthropometry, dichotomizing the variable WC (elevated: men > 102 cm and women > 88 cm), showing a similar percentage in elevated values before the dietary intervention (CC vs CT + TT) (47.4 % vs 46.4 %; $p = 0.43$) and after the dietary intervention (35.8 % vs 34.5 %; $p = 0.56$).

Biochemical parameters are reported in table II. The decrease in biochemical variables was not significant in patients with the T allele. In non-T allele carriers, insulin, HOMA-IR, triglycerides and CRP levels decreased. The decrease of these parameters in non-T allele carriers was statistically significant; triglycerides (-18.3 ± 4.3 mg/dl; $p = 0.02$), CRP (-2.6 ± 0.3 mg/dl; $p = 0.02$), insulin (-4.4 ± 1.9 mU/l; $p = 0.02$) and HOMA-IR (-2.1 ± 0.7 ; $p = 0.03$). Therefore, the improvement of all the previously mentioned parameters was statistically superior in non-carriers of the T allele than in non-T allele carriers. Finally, statistical differences in insulin and HOMA-IR were detected among basal and post-treatment values of variables between major allele genotype CC and minor allele genotype (CT + TT). The values of these parameters were higher in T allele carriers than in non-T allele carriers.

Table III reports changes of circulating adipokines. Leptin levels decrease in both genotype groups after dietary intervention (-22.3 ± 4.5 ng/dl in non-T allele carriers vs -18.0 ± 4.2 ng/dl in T allele carriers; $p = 0.45$). Resistin levels decrease in non-T allele carriers after dietary intervention (-1.2 ± 0.2 ng/dl in non-T allele carriers vs -0.6 ± 0.1 ng/dl in T allele carriers; $p = 0.02$). Adiponectin levels remained unchanged in both groups. Differences in resistin levels were detected among basal and post-treatment between both genotype groups CC vs CT/TT. The values of resistin were higher in T allele carriers than in non-T allele carriers.

DISCUSSION

As far as we know, this is the first interventional study that analyzes the effects of a hypocaloric diet and variant rs3138167 on metabolic response after a significant weight loss. In our study analyzing this genetic variant, we have observed a significant association between T allele of this SNP and worse response of insulin resistance, triglycerides, CRP and circulating resistin than in non-T allele carriers after weight loss with a hypocaloric Mediterranean diet.

The role of resistin on metabolism is controversial. Some authors have observed that resistin levels were related to obesity and its comorbidities (23) and other studies did not report these associations (24,25). On the other hand, the effect on resistin levels of dietary or other interventions that produce weight loss are also scarce and contradictory. There are studies with hypocaloric diets (15) in which the levels of resistin have not changed. Other dietary interventions studies have demonstrated a decrease of resistin levels after weight loss (26) or even an increase of serum levels after weight loss (27). The biological response of resistin is complex since after adjustable gastric banding, a biphasic response with a decrease in resistin levels initially and an increase at 12 months of follow-up have been reported (28).

Table II. Basal and post-intervention levels biochemical parameters (mean ± SD)

Parameters	CC (n = 843)		CT + TT (n = 168)		p values – Time CC – Basal genotype – Time CT + TT – 12 weeks genotype
	Basal	12 weeks	Basal	12 weeks	
Glucose (mg/dl)	102.2 ± 4.1	99.1 ± 5.1	99.9 ± 4.2	96.9 ± 5.3	p = 0.17 p = 0.34 p = 0.18 p = 0.48
Total cholesterol (mg/dl)	206.9 ± 10.7	189.2 ± 13.2	203.9 ± 9.1	197.1 ± 8.2	p = 0.12 p = 0.51 p = 0.31 p = 0.38
LDL-cholesterol (mg/dl)	128.7 ± 9.3	120.1 ± 8.1	123.2 ± 7.1	119.1 ± 4.0	p = 0.13 p = 0.45 p = 0.12 p = 0.46
HDL-cholesterol (mg/dl)	53.5 ± 4.1	52.4 ± 4.2	55.3 ± 4.0	54.8 ± 3.2	p = 0.29 p = 0.41 p = 0.50 p = 0.49
Triglycerides (mg/dl)	135.6 ± 11.9	117.5 ± 9.4*	129.1 ± 13.1	118.6 ± 11.2	p = 0.02 p = 0.60 p = 0.13 p = 0.44
Insulin (mU/l)	14.6 ± 1.9	10.2 ± 2.1 [†]	17.4 ± 2.2 [¶]	13.7 ± 2.0 [¶]	p = 0.02 p = 0.01 p = 0.13 p = 0.04
HOMA-IR	3.6 ± 0.8	2.6 ± 0.9 [‡]	5.4 ± 1.0 [¶]	5.0 ± 0.9 [¶]	p = 0.03 p = 0.03 p = 0.12 p = 0.03
CRP	6.5 ± 2.0	3.9 ± 1.3 [§]	6.6 ± 1.1	5.8 ± 1.1	p = 0.02 p = 0.49 p = 0.23 p = 0.15

HOMA-IR: homeostasis model assessment; CRP: C-reactive protein. Statistical differences $p < 0.05$, in each genotype group (*triglycerides, [†]insulin, [‡]HOMA IR, [§]CRP). Statistical differences $p < 0.05$, in different genotype groups ([¶]insulin, [¶]HOMA-IR). First p, significance of dietary intervention after 12 weeks in CC genotype; second p, significance between CC genotypes vs CT + TT baseline values; third p, significance of dietary intervention after 12 weeks in CT + TT genotype; and fourth p, significance between CC genotypes vs CT + TT post-treatment values.

Different nutrients present in the diet and also genetic variants may be involved in these observed responses. For example, Cabrera et al. (29) showed that resistin level is positively associated with saturated fat intake and inversely associated with monounsaturated fat intake. In other interventional study (30), resistin levels decreased after a dietary intervention supplemented with wakame and carobs. Interestingly, on the basis of dif-

ferent components of Mediterranean diet (omega 3 fatty acids, monounsaturated fatty acids, resveratrol, quercetin, etc.), it has been suggested that bioactive molecules in the Mediterranean diet may improve different cardiovascular risk factors through modulation of gene expression (31).

A previous study (15) with a hypocaloric diet of 1,500 calories and a macronutrient distribution of 52 % of calories in the form

Table III. Basal and post-intervention levels of serum adipokines (mean \pm SD)

Parameters	CC (n = 843)		CT + TT (n = 168)		p values – Time CC – Basal genotype – Time CT + TT – 12 weeks genotype
	Basal	12 weeks	Basal	12 weeks	
Resistin (ng/dl)	3.9 \pm 0.5	2.7 \pm 0.4†	4.9 \pm 0.6‡	4.1 \pm 0.5‡	p = 0.01 p = 0.03 p = 0.13 p = 0.02
Adiponectin (ng/dl)	33.2 \pm 9.1	37.2 \pm 4.1	30.1 \pm 7.0	39.3 \pm 5.2	p = 0.23 p = 0.51 p = 0.29 p = 0.54
Leptin (ng/dl)	93.1 \pm 10.6	71.2 \pm 8.5*	93.2 \pm 10.2	65.8 \pm 8.1*	p = 0.02 p = 0.41 p = 0.02 p = 0.44

Statistical differences $p < 0.05$, in each genotype group (*leptin, †resistin). Statistical differences $p < 0.05$, in different genotype groups (‡resistin). First p, significance of dietary intervention after 12 weeks in CC genotype; second p, significance between CC genotypes vs CT + TT baseline values; third p, significance of dietary intervention after 12 weeks in CT + TT genotype; and fourth p, significance between CC genotypes vs CT + TT post-treatment values.

of carbohydrates, 25 % in the form of lipids (50 % monounsaturated fats) and 23 % in the form of proteins, reported a different metabolic response based on the genetic variant rs10401670 of the RETN gene. These results were similar with a diet that reached the same caloric restriction, around 1,500 calories but with a higher fat percentage of 41 % with 60 % monounsaturated fats (32). The composition of this last diet is very similar to that of our present study, and when the variant rs3138167 was analyzed, we also found differences in metabolic parameters and resistin levels.

An interesting result of our work is the increased levels of resistin in T allele carriers with a worse metabolic status (insulin and HOMA-IR). It has been showed that resistin induces the synthesis of inflammatory cytokines in macrophages in adipose tissue and that inflammatory status induces expression of the resistin gene (33), as a vicious circle. Inflammation environment is well-known to be involved in the pathogenesis of insulin resistance, and circulating resistin could link inflammation and insulin resistance. For example, resistin is shown to elevate tumor necrosis factor- α (TNF- α) levels (34) and resistin decreases the secretion of anti-inflammatory adipokines such as adiponectin (35). All these metabolic environments generate resistance to insulin. In our present intervention study, no T-allele carriers improved inflammatory parameters (CRP), insulin and HOMA-IR with the same weight loss as the T allele carriers. The observed improvement in triglyceride levels may also be related to the improvement in the inflammatory status of patients that is observed after weight loss in T allele carriers. The role of resistin as an inflammatory molecule has been shown in other pathologies not related to obesity such as rheumatoid arthritis (36) and prothrombotic status (37).

The increased levels of resistin finding in our study in T allele carriers can be explained by various theories. First, it can be associated to specific binding of transcription factors Sp1 and Sp3 to a promoter element leading to increase promoter activity as showed with other SNPs (38). Second, this genetic variant may be in linkage disequilibrium with another SNP that modulates expression of resistin (38). Finally, this genetic variant would be part of a sequence of regulatory element that mediates the binding of coregulatory protein involved in the regulation of RETN and DC genes expression (39).

There are some limitations of our study. First, only one genetic variant has been evaluated. Second, it lacks a control group without a dietary intervention in order to compare the effect of weight loss. Third, new studies are necessary to validate these findings in overweight patients, as well as in diabetic patients and other high risk populations. Fourth, further studies are necessary to validate these findings in overweight patients, as well as in diabetic patients. Finally, the short duration of the clinical trial does not allow us to observe what would happen to the resistance levels during a longer period.

We report an association of rs3138167 with a worse metabolic response (insulin, HOMA-IR, triglyceride and CRP) in T allele carriers after weight loss with a hypocaloric diet with Mediterranean pattern.

REFERENCES

1. Stepan CM, Bailey ST, Bhat S, Brown EJ, Banerjee RR, Wright CM, et al. The hormone resistin links obesity to diabetes. *Nature* 2001;409:307-12. DOI: 10.1038/35053000
2. Di Raimo T, Azzara G, Corsi M, Cipollone D, Vasco VR, Businaro R. Adipokines and their involvement as a target of new drugs. *Aust J Pharm* 2015;3:166-70.

3. Pagano C, Marin O, Calcagno A, Schiappelli P, Pilon C, Milan G, et al. Increased serum resistin in adults with Prader-Willi syndrome is related to obesity and not to insulin resistance. *J Clin Endocrinol Metab* 2005;90:4335-40. DOI: 10.1210/jc.2005-0293
4. Burnett MS, Devaney JM, Adenika RJ, Lindsay R, Howard BV. Cross-sectional associations of resistin, coronary heart disease and insulin resistance. *J Clin Endocr Metab* 2006;91:64-8. DOI: 10.1210/jc.2005-1653
5. Zou CC, Liang L, Hong F, Fu JF, Zhao ZY. Serum adiponectin, resistin levels and non-alcoholic fatty liver disease in obese children. *Endocrine J* 2005;52:519-24. DOI: 10.1507/endocrj.52.519
6. Steppan CM, Wang J, Whiteman EL, Birnbaum MJ, Lazar MA. Activation of SOCS-3 by resistin. *Mol Cell Biol* 2005;25:1569-75.
7. Menzaghi C, Coco A, Salvermini L, Thompson R, De Cosmo S, Doria A, et al. Heritability of serum resistin and its genetic correlation with insulin resistance-related features in nondiabetic Caucasians. *J Clin Endocrinol Metab* 2006;91:2792-5. DOI: 10.1210/jc.2005-2715
8. Hiwert M, Manning A, LcAteer J, Dupuis J, Fox C, Cuples LA. Association of variant in RETN with plasma resistin levels and diabetes-related traits in the Framingham offspring Study. *Diabetes* 2009;58:750-6. DOI: 10.2337/db08-1339
9. Ortega L, Navarro P, Riestra P, Gavela-Pérez T, Soriano-Guillén L, Garcés C. Association of resistin polymorphisms with resistin levels and lipid profile in children. *Mol Biol Rep* 2014;41:7659-64. DOI: 10.1007/s11033-014-3658-8
10. Engert JC, Vohl MC, Williams SM, Lepage P, Loredó-Osti JC, Faith J, et al. 5' flanking variants of resistin are associated with obesity. *Diabetes* 2002;51:1629-34. DOI: 10.2337/diabetes.51.5.1629
11. Osawa H, Yamada K, Onuma H, Murakami A, Ochi M, Kawata H, et al. The G/G genotype of a resistin single-nucleotide polymorphism at -420 increases type 2 diabetes mellitus susceptibility by inducing promoter activity through specific binding of Sp1/3. *Am J Hum Genet* 2004;75:678-86. DOI: 10.1086/424761
12. Onuma H, Tabara Y, Kawamura R, Ohashi J, Nishida W, Takata Y, et al. Plasma resistin is associated with single nucleotide polymorphisms of a possible resistin receptor, the decorin gene, in the general Japanese population. *Diabetes* 2013;62:649-52. DOI: 10.2337/db12-0058
13. Mancini JG, Filion KB, Atallah R, Eisenberg MJ, Mancini JG, Filion KB, et al. Systematic review of the Mediterranean diet for long-term weight loss. *Am J Med* 2016;129:407-15. DOI: 10.1016/j.amjmed.2015.11.028
14. Widmer RJ, Flammer AJ, Lerman LO, Lerman A. The Mediterranean diet, its components, and cardiovascular disease. *Am J Med* 2015;128:229-38. DOI: 10.1016/j.amjmed.2014.10.014
15. Antonio de Luis D, Aller R, Izaola O, Primo D, Bachiller R. The rs10401670 variant in resistin gene improved insulin resistance response and metabolic parameters secondaries to weight loss after a hypocaloric diet. *Clin Nutr ESPEN* 2016;14:14-8. DOI: 10.1016/j.clnesp.2016.04.028
16. Antonio de Luis D, Izaola O, Primo D, Aller R, Pacheco D. Effect of two polymorphisms of the resistin gene (rs10401670 and rs1862513) on resistin levels and biochemical parameters in morbidly obese patients 1 year after a biliopancreatic diversion surgery. *Clin Nutr* 2016;35:1517-21. DOI: 10.1016/j.clnu.2016.04.005
17. Friedewald WT, Levy RJ, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma without use of the preparative ultracentrifuge. *Clin Chem* 1972;18:499-502. DOI: 10.1093/clinchem/18.6.499
18. Mathews DR, Hosker JP, Rudenski AS. Homeostasis model assessment: insulin resistance and beta cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985;28:412-4. DOI: 10.1007/BF00280883
19. Pflutzner A, Langefeld M, Kunt T. Evaluation of human resistin assays with serum from patients with type 2 diabetes and different degrees of insulin resistance. *Clin Lab* 2003;49:571-6.
20. Meier U, Gressner M. Endocrine regulation of energy metabolism: review of pathobiochemical and clinical chemical aspects of leptin, ghrelin, adiponectin and resistin. *Clin Chem* 2004;50:1511-25. DOI: 10.1373/clinchem.2004.032482
21. Lukaski H, Johnson PE. Assessment of fat-free mass using bioelectrical impedance measurements of the human body. *Am J Clin Nutr* 1985;41:810-7. DOI: 10.3138/cmlr.41.5.810
22. Mataix J, Mañas M. Tablas de composición de alimentos españoles. Granada: Universidad de Granada; 2003.
23. Steppan CM, Lazar MA. The current biology of resistin. *J Intern Med* 2004;255:439e47. DOI: 10.1111/j.1365-2796.2004.01306.x
24. Iqbal N, Seshadri P, Stern L, Loh J, Kundu S, Jafar T, et al. Serum resistin is not associated with obesity or insulin resistance in humans. *Eur Rev Med Pharmacol Sci* 2005;9:161e5.
25. Vendrell J, Broch M, Vilarrasa N. Resistin, adiponectin, ghrelin, leptin and proinflammatory cytokines: relationships in obesity. *Obes Res* 2004;12:962. DOI: 10.1038/oby.2004.118
26. Varady KA, Tussing L, Bhutani S, Braunschweig CL. Degree of weight loss required to improve adipokine concentrations and decrease fat cell size in severely obese women. *Metabolism* 2009;58:1096e101. DOI: 10.1016/j.metabol.2009.04.010
27. Elloumi M, Ben Ounis O, Makni E, Van Praagh E, Tabka Z, Lac G. Effect of individualized weight-loss programmes on adiponectin, leptin and resistin levels in obese adolescent boys. *Acta Paediatr* 2009;98:1487-93. DOI: 10.1111/j.1651-2227.2009.01365.x
28. Moschen AR, Molnar C, Wolf A, Weiss H, Graziadei I, Kaser S, et al. Effects of weight loss induced by bariatric surgery on hepatic adipocytokine expression. *J Hepatol* 2009;51:765-77. DOI: 10.1016/j.jhep.2009.06.016
29. Cabrera de León A, Almeida González D, González Hernández A, Domínguez Coelho S, Marrugat J, Juan Alemán Sánchez J, et al. Relationships between serum resistin and fat intake, serum lipid concentrations and adiposity in the general population. *J Atheroscler Thromb* 2014;21:454-62. DOI: 10.5551/jat.22103
30. Izaola O, Primo D, Rico Bargaúes D, Martín-Diana AB, Martínez Villaluenga C, Miranda J, et al. Effects of a snack enriched with carob and *Undaria pinnatifida* (wakame) on metabolic parameters in a double blind, randomized clinical trial in obese patients. *Nutr Hosp* 2020;34:465-73.
31. Toledo E, Wang DD, Ruiz-Canela M, Clish CB, Razquin C, Zheng Y, et al. Plasma lipidomic profiles and cardiovascular events in a randomized intervention trial with the Mediterranean diet. *Am J Clin Nutr* 2017;106:973-83.
32. De Luis D, Aller R, Izaola O, Primo D. Role of the rs10401670 variant in the resistin gene on the metabolic response after weight loss secondary to a high-fat hypocaloric diet with a Mediterranean pattern. *J Hum Nutr Diet* 2022;35(4):722-30. DOI: 10.1111/jhn.12975
33. Lehrke M, Reilly MP, Millington SC, Iqbal N, Rader DJ, Lazar MA. An inflammatory cascade leading to hyperresistinemia in humans. *PLoS Med* 2004;1:e45. DOI: 10.1371/journal.pmed.0010045
34. Aquilante CL, Kosmiski LA, Knutsen SD, Zineh I. Relationship between plasma resistin concentrations, inflammatory chemokines, and components of the metabolic syndrome in adults. *Metabolism* 2008;57:494-501. DOI: 10.1016/j.metabol.2007.11.010
35. Kim KH, Zhao L, Moon Y, Kang C, Sul HS. Dominant inhibitory adipocyte-specific secretory factor (ADSF)/resistin enhances adipogenesis and improves insulin sensitivity. *Proc Natl Acad Sci U S A* 2004;101:6780-5.
36. Senolt L, Housa D, Vernerova Z, Jirasek T, Svobodova R, Veigl D. Resistin is abundantly present in rheumatoid arthritis synovial tissue, synovial fluid, and elevated serum resistin reflects disease activity. *Ann Rheum Dis* 2007;66:458-63. DOI: 10.1136/ard.2006.054734
37. Qi Q, Wang J, Li H, Yu Z, Ye X, Hu FB, et al. Associations of resistin with inflammatory and fibrinolytic markers, insulin resistance, and metabolic syndrome in middle-aged and older Chinese. *Eur J Endocrinol* 2008;159:585-93. DOI: 10.1530/EJE-08-0427
38. Osawa H, Yamada K, Onuma H, Murakami A, Ochi M, Kawata H, et al. The G/G genotype of a resistin single-nucleotide polymorphism at -420 increases type 2 diabetes mellitus susceptibility by inducing promoter activity through specific binding of Sp1/3. *Am J Hum Genet* 2004;75:678-86.
39. Osawa H, Yamada K, Onuma H, Murakami A, Ochi M, Kawata H, et al. The G/G genotype of a resistin single-nucleotide polymorphism at -420 increases type 2 diabetes mellitus susceptibility by inducing promoter activity through specific binding of Sp1/3. *Am J Hum Genet* 2004;75:678-86. DOI: 10.1086/424761