

Original

Limitations of immunodeficient mice as models for osteoporosis studies

Álvaro del Real¹, Laura López-Delgado², Carolina Sañudo², Belén García-Montesinos³, Carlos Wert-Carvajal⁴, Esther Laguna⁵, Carmen García-Ibarbia², José A. Saiz-Aja⁶, Diego Ferreño⁶, José A. Casado⁶, Guillermo Menéndez⁵, M. Isabel Pérez-Núñez⁵, José A. Riancho^{1,2}

¹Department of Medicine and Psychiatry. Universidad de Cantabria. Santander, Spain. Departments of ²Internal Medicine, and ³Maxillofacial Surgery. Hospital Universitario Marqués de Valdecilla-IDIVAL. Universidad de Cantabria. Santander, Spain. 4Institute of Experimental Epileptology and Cognition Research. Medical Faculty. University of Bonn. Bonn, Germany. 5Department of Traumatology and Orthopedic Surgery. Hospital Universitario Marqués de Valdecilla. Universidad de Cantabria-IDIVAL. Santander, Spain. ⁶Department of Material Science. LADICIM. Universidad de Cantabria. Santander, Spain

Abstract

We aimed to establish a murine model to investigate the potential therapeutic role of human mesenchymal stem cells (MSCs) in skeletal disorders in vivo. Therefore, we specifically focused on 2 experimental models: the bisphosphonate-related osteonecrosis of the jaw (ONJ) and ovariectomy (OVX)-induced bone loss to simulate postmenopausal osteoporosis. Utilizing NOD.CB17-Prkdcscid/J (NOD-SCID) mice, known for their compromised immune systems, we examined the development of ONJ following varying dosages and routes of administration of zoledronic acid, with and without adjunctive dexamethasone treatment. Surprisingly, we found a very low incidence of ONJ compared to the results reported in immunocompetent mice, suggesting that factors intrinsic to the NOD-SCID mice, such as immune deficiency and possibly altered microbiota due to sterile housing conditions, may influence the development of this condition. On the other hand, these mice did not show the anticipated bone loss following bilateral OVX, challenging conventional wisdom and emphasizing the multifaceted nature of osteoporosis involving both the immune system and microbiota. This study reveals the limitations of immunodeficient mice as experimental models in bone research. On the other hand, it is consistent with experimental data suggesting a role of osteoimmunology and osteomicrobiology mechanisms in the pathogenesis of some skeletal disorders.

Keywords: Immunodeficient mice. Osteonecrosis. Ovariectomy. Bisphosphonates.

Received: 04/29/2024 • Accepted: 05/20/2024

Funding: supported by a grant from Instituto de Salud Carlos III (PI16/915), which can be cofunded by European Union FEDER funds. Alvaro del Real received support by the postdoctoral grant Margarita Salas in the University of Cantabria, Santander, Spain.

Conflicts of interest: the authors declare no conflict of interest.

Artificial intelligence: the authors declare that they did not used any artificial intelligence (AI) or Al-assisted technologies to write this the article.

Del Real A, López-Delgado L, Sañudo C, García-Montesinos B, Wert-Carvajal C, Laguna E, García-Ibarbia C, Saiz-Aja JA, Ferreño D, Casado JA, Menéndez G, Pérez-Núñez MI, Riancho JA. Limitations of immunodeficient mice as models for osteoporosis studies. Rev Osteoporos Metab Miner 2024;16(1):10-15

DOI: 10.20960/RevOsteoporosMetabMiner.00041

Correspondence:

José A. Riancho. Department of Internal Medicine. Hospital Universitario Marqués de Valdecilla-IDIVAL. Universidad de Cantabria. Av. de Valdecilla, s/n. 39008 Santander, Spain

e-mail: rianchoj@unican.es

INTRODUCTION

NOD.CB17-Prkdcscid/J mice, commonly known as NOD-SCID mice, are a genetically engineered strain of mice frequently used in biomedical research (1). These mice are characterized by a severely compromised immune system, due to a specific mutation in the Prkdc gene, which encodes a protein kinase vital for DNA repair during immune cell development, and by their background strain, the nonobese diabetic (NOD) mouse, which provides additional unique immune system characteristics. Due to the mutation in the Prkdc gene, these mice lack functional T cells and B cells, which makes them unable to mount effective adaptive immune responses (2). This deficiency prevents them from rejecting materials from foreign species, such as human cells. The NOD background contributes to defects in the natural killer (NK) cell function, which is typically less compromised in other SCID strains (2,3). This makes NOD-SCID mice particularly useful for xenograft experiments, in which human cells or tissues are grafted into mice. Therefore, they are extensively used in research involving the transplantation of human cells (1-4).

In the early 2000s, physicians started observing osteonecrosis of the jaw (ONJ) in patients without a history of radiation exposure and realized that most of these patients had breast cancer metastatic to bone or myelomatous disease and were on bisphosphonate (BP) therapy. This led investigators to propose an association between BP use and ONJ (5). The estimated incidence is 1-90 cases per 100 000 patients on BPs per year (6). Most of these cases occur in patients on high doses of BPs (especially zoledronate and pamidronate) due to cancer. Treating ONJ may be rather difficult. In recent years, mesenchymal stem cells (MSCs) have been proposed as candidates for cellular therapies in different conditions including ONJ. MSCs are adult multipotent stromal cells that can differentiate into a variety of cell types, such as osteoblasts, chondrocytes and adipocytes (7,8). Given their ability to differentiate into osteoblasts, they are attractive candidates for bone regeneration therapies. Recent research underscores the multifaceted role of MSCs, mediated not only by differentiation into osteoblasts, but also by the modulation of the bone-healing environment through paracrine effects, involving the secretion of growth factors and cytokines (9-11). Therefore, some studies in animal models have obtained promising results with MSCs applied by either systemic or local routes (12-15). To explore the potentially beneficial effect of a systemic administration of human MSCs in vivo, we tried to develop a murine model of ONJ and ovariectomy (OVX)-induced osteoporosis. To prevent rejection, we used immunodeficient mice and protocols previously applied to induce ONJ and osteoporosis in immunocompetent mice.

MATERIAL AND METHODS

NOD.CB17-Prkdcscid/J called NOD-SCID mice were used. Founders were obtained from Jackson Laboratories (Bar, Harbor, Maine, United States) and the colony was housed at the animal facility of University of Cantabria, Santander, Spain under aseptic conditions and veterinary control.

For the ONJ models, 8 week-old mice were anesthetised and the maxillary right first molar was extracted, as published using immunocompetent mice (16). Mice were treated with several intraperitoneal (IP) or intravenous (IV) (retro-orbital plexus) doses of 540 μg/kg of zoledronic acid. The first dose was always given 1 week before the dental extraction, with subsequent doses at weekly intervals. Some mice additionally received subcutaneous dexamethasone (10 mg/kg, 3 times a week). Mice were euthanized 1 week after the last zoledronic acid dose. The maxillary bones were dissected, fixed in formaldehyde and preserved in ethanol prior to study by micro-CT (Bruker). In some experimental mice, as well as in control mice of the same age which did not undergo any experimental procedures, the femur and the tibia were dissected and the trabecular bone volume was determined as previously described (17). After micro-CT analysis, bones were decalcified, paraffin-embedded, and stained with hematoxylin-eosin. Regions of denuded mucosa and bone necrosis (defined as 5 adjacent empty osteocytic lacunae [18]) were evaluated by 2 independent observers.

For the osteoporosis model, female 8-week old NOD-SCID mice were used. Under general anesthesia, both ovaries were exposed and removed. Mice receive opioid analgesia for pain management right after the intervention. A sham-operated group was also included. Eight weeks later, the animals were euthanized, and both the femora and the tibiae were removed and analyzed by micro-CT. T-test was assessed as statistical analysis to compare the differences between mice groups (*p*-values indicated).

Protocol approval was obtained from University of Cantabria Research Ethics Committee and Consejeria de Sanidad de Cantabria, as established by current regulations (code 2016-27930).

RESULTS

OSTEONECROSIS OF THE JAW MODEL

In this study, 27 mice were used to establish an osteonecrosis of the jaw (ONJ) model, receiving varying doses of zoledronic acid administered via IV or IP acces. Table I illustrates the different treatment regimens, including the number of doses, administration route, duration 12 Á. DEL REAL ET AL.

Table I. Drug treatment and responses in the ONJ group					
Treatment	Doses of zoledronic acid (n)	Administration route	Time of study (weeks after molar removal)	Mice (n)	Osteonecrosis cases (n)
Zoledronic acid	5	IP	5	11	0
Zoledronic acid	3	IP	3	3	0
Zoledronic acid, dexamethasone	3	IP	3	2	0
Zoledronic acid	2	IV	2	6	0
Zoledronic acid, dexamethasone	2	IV	2	5	1
IP: intraperitoneal administration group; IV: intravenous administration group.					

of the study following molar extraction, and the incidence of osteonecrosis observed among the mice.

Micro-CT analysis confirmed the absence of the first molar, without abnormalities in the maxillary bone (Figs. 1 A and B). Only 1 out of the 27 mice studied showed some evidence of mild ONJ, with a small area of bone with several empty osteocytic lacunae. This mouse had been treated with 2 IV injections of zoledronic acid and dexamethasone (Figs. 1 C and D).

In some representative mice, we measured bone mass to confirm the bioactivity of zoledronic acid. As expected, trabecular bone volume was increased by zoledronic acid via IV or IP access (from a mean 30 ± 8 % to 48 ± 10 % at the femur, and 32 ± 6 % to 48 ± 12 % at the tibia).

OSTEOPOROSIS MODEL

In the evaluation of an osteoporosis model, 24 ovariectomized (OVX) mice and 11 controls (COVX) were successfully included for micro-CT analyses. Unexpectedly, bone mineral density (BMD), bone volume versus tissue volume (BV/TV) parameter, and cortical thickness were similar between the 2 groups (Fig. 2). Specifically, the mean BMD for the femur was 0.20 g/cm² in the OVX group, identical to the COVX group. The BV/TV mean values were $20.3 \pm 4.7 \%$ for OVX and 21.8 ± 3.2 for COVX. Hence, no significant differences were found in the bone measured parameters (BMD; p = 0.932; BV/TV p = 0.1507). Dissimilarities were not found either in the femur cortical thickness (p = 0.4242).

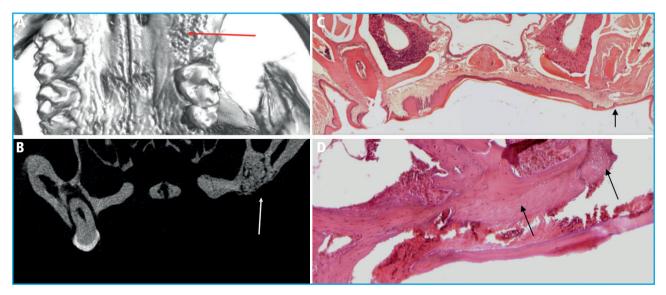


Figure 1. A. 3D reconstruction of the micro-CT analysis showing absence of first molar and cavity filled with new bone (red arrow). B. Coronal image, micro-CT analysis showing the absence of the first molar and the cavity filled with new bone (white arrow). C. Normal healing after tooth removal. HE staining showing complete regeneration of oral mucosa and filling of the cavity (black arrow). Image captured at 4× magnification. D. Mild bone necrosis. HE staining of the only mice showing an area of bone necrosis adjacent to the removed molar cavity (black arrows). Image captured at 10× magnification.

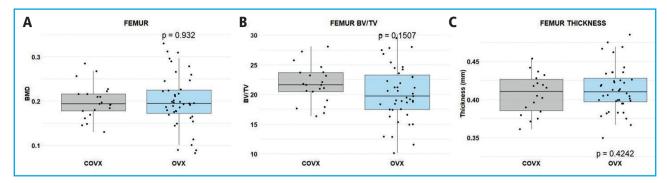


Figure 2. Comparative BMD and cortical thickness analyses with micro-CT. A. Femoral BMD of control (COVX, gray) and ovariectomized (OVX, blue) groups. B. Femoral BV/TV of COVX (gray) vs OVX (blue). C. Femoral cortical thickness of COVX (gray) and OVX (blue) groups. Statistical significance assessed via t-test (*p*-values indicated). Both lower limbs are represented for each mice as an individual point.

DISCUSSION

BPs are the most widely used antiresorptives in the treatment of OP and certain tumors. ONJ is a rare but serious side effect that can arise in patients on antiresorptives. As in other many complex disorders, ONJ seems to be the result of a combination of genetic and environmental factors. There are several well-established acquired risk factors associated with ONJ, such as previous dental extraction, periodontal disease, and the administration of BP (with differences depending on the type and administration route). Local infections and the immune response are also considered part of the ONJ pathogenesis. Different studies have proposed a potential role of $T\gamma\delta$ lymphocytes (18), neutrophils, and macrophages, as well as interleukins and proteins produced by cells involved in the immune system (19).

Despite molar extraction and the injection of high doses of zoledronic acid, only 1 out of the 27 studied mice developed histological findings consistent with mild ONJ. These results widely differ from the 45%-50% of ONJ cases described in studies in immunocompetent mice using a similar methodology (20,21).

Unlike immunocompetent mice, it seems that immunosuppressed mice tend not to develop ONJ even after an insult to the oral mucosa and having received high doses of BP and steroids. Although the mechanisms involved await further studies, we can speculate that it could be due to the immune deficiency *per se* or the sterile conditions in which those mice were maintained, which likely modified oral microbiota. These failed experiments reinforce the concept that factors apart from medication, such as the immune system and the microbiota, play an important role in the occurrence of ONJ after BP therapy.

In line with the ONJ model, the absence of a functional immune system in NOD.CB17-Prkdcscid/J mice, and/ or the maintenance of the mice under sterile conditions seemed to prevent them from developing the expected bone loss after OVX. Studies using immu-

nocompetent mice typically report a significant decrease in BMD and changes to bone architecture after 3 weeks following ovariectomy. For example, Smith et al. (2020) observed a significant reduction in BMD and BV/TV and significant increases in trabecular separation in C57BL/6 mice 8 weeks after ovariectomy, highlighting the accelerated bone loss in this model due to normal immune function (22). These differences underscore the critical role of the immune system in post-ovariectomy bone remodeling and the potential for unique disease progression pathways in immunocompromised conditions.

The immune system plays a pivotal role in bone homeostasis, with T cells, B cells, and cytokines influencing both bone resorption and formation. In particular, T cells have been involved in osteoclastogenesis through the production of RANKL, a key osteoclast-activating factor (23). Additionally, classic studies from the Pacifici and Manolagas groups confirmed the role played by cytokines, such as IL-1, IL-6, and tumor necrosis factor, produced by immune and other cells, in OVX-induced osteoporosis (24-26). The lack of an adaptive immune response in these immunodeficient mice could therefore significantly ameliorate the osteoclast-mediated bone resorption expected to occur after ovariectomy.

Moreover, gut microbiota has been increasingly recognized for its influence on bone density and health (27). Microbiota dysbiosis has been associated with inflammatory responses capable of promoting osteoclast activity, eventually leading to bone loss (28-30). The sterile conditions under which NOD.CB17-Prkdcscid/J mice are kept could impact their microbiota, potentially affecting the incidence and progression of bone loss. This concept would be consistent with reports showing that estrogen deficiency causes a gut microbiome-dependent expansion of Th17 cells and TNF- α -producing T cells. This may be of pathophysiological importance, because the blockade of Th17 cell and TNF+ T cell egress from the gut prevented OVX-induced bone loss (31).

14 A. DEL REAL ET AL.

In conclusion, the blunted ability to induce BP-related ONJ and OVX-related decrease in bone mass is a limitation for using immunodeficient mice as experimental models in osteoporosis research. On the other hand, they highlight the role of osteoinmmunological mechanisms, and perhaps body microbiota, in bone pathophysiology. The correlation between these mechanisms and their collective impact on bone health requires a broader perspective when investigating osteoporosis and should be taken into account when conducting studies with animal models.

REFERENCES

- Pearson T, Greiner DL, Shultz LD. Humanized SCID mouse models for biomedical research. Curr Top Microbiol Immunol 2008;324:25-51. DOI: 10.1007/978-3-540-75647-7_2
- 2. Shultz LD, Lyons BL, Burzenski LM, Gott B, Chen X, Chaleff S, et al. Human lymphoid and myeloid cell development in NOD/LtSz-scid IL2R gamma null mice engrafted with mobilized human hemopoietic stem cells. J Immunol 2005;174(10):6477-89. DOI: 10.4049/jimmunol.174.10.6477
- 3. Ito M, Hiramatsu H, Kobayashi K, Suzue K, Kawahata M, Hioki K, et al. NOD/SCID/gamma(c)(null) mouse: an excellent recipient mouse model for engraftment of human cells. Blood 2002;100(9):3175-82. DOI: 10.1182/blood-2001-12-0207
- Ito M, Kobayashi K, Nakahata T. NOD/Shi-scid IL2rgamma(null) (NOG) mice more appropriate for humanized mouse models. Curr Top Microbiol Immunol 2008;324:53-76. DOI: 10.1007/978-3-540-75647-7
- Ruggiero SL, Mehrotra B, Rosenberg TJ, Engroff SL. Osteonecrosis of the Jaws Associated with the Use of Bisphosphonates: A
 Review of 63 Cases. J Oral Maxillofac Surg 2004;62(5):527-34.
 DOI: 10.1016/j.joms.2004.02.004
- Khan A, Morrison A, Cheung A, Hashem W, Compston J. Osteonecrosis of the jaw (ONJ): diagnosis and management in 2015. Osteoporos Int 2016;27(3):853-9. DOI: 10.1007/s00198-015-3335-3
- 7. Friedenstein AJ, Chailakhjan RK, Lalykina KS. The development of fibroblast colonies in monolayer cultures of guinea-pig bone marrow and spleen cells. Cell Tissue Kinet 1970;3(4):393-403. DOI: 10.1111/j.1365-2184.1970.tb00347.x
- Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, et al. Multilineage Potential of Adult Human Mesenchymal Stem Cells. Science 1999;284(5411):143-7. DOI: 10.1126/science.284.5411.143
- Torrecillas-Baena B, Pulido-Escribano V, Dorado G, Gálvez-Moreno MÁ, Camacho-Cardenosa M, Casado-Díaz A. Clinical Potential of Mesenchymal Stem Cell-Derived Exosomes in Bone Regeneration. J Clin Med 2023;12(13):4385. DOI: 10.3390/ icm12134385
- Zou J, Yang W, Cui W, Li C, Ma C, Ji X, et al. Therapeutic potential and mechanisms of mesenchymal stem cell-derived exosomes as bioactive materials in tendon-bone healing. J Nanobiotechnology 2023;21(1):14. DOI: 10.1186/s12951-023-01778-6
- Fan S, Sun X, Su C, Xue Y, Song X, Deng R. Macrophages bone marrow mesenchymal stem cells crosstalk in bone hea-

- ling. Front Cell Dev Biol 2023;11:1193765. DOI: 10.3389/fce-ll.2023.1193765
- Kikuiri T, Kim I, Yamaza T, Akiyama K, Zhang Q, Li Y, et al. Cell-based immunotherapy with mesenchymal stem cells cures bisphosphonate-related osteonecrosis of the jaw-like disease in mice. J Bone Miner Res 2010;25(7):1668-79. DOI: 10.1002/jbmr.37
- Rodríguez-Lozano FJ, Oñate-Sánchez R, Gonzálvez-García M, Vallés-Bergadá M, Martínez CM, Revilla-Nuin B, et al. Allogeneic Bone Marrow Mesenchymal Stem Cell Transplantation in Tooth Extractions Sites Ameliorates the Incidence of Osteonecrotic Jaw-Like Lesions in Zoledronic Acid-Treated Rats. J Clin Med 2020;9(6):1649. DOI: 10.3390/jcm9061649
- Ogata K, Katagiri W, Osugi M, Kawai T, Sugimura Y, Hibi H, et al. Evaluation of the therapeutic effects of conditioned media from mesenchymal stem cells in a rat bisphosphonate-related osteonecrosis of the jaw-like model. Bone 2015;74:95-105. DOI: 10.1016/j.bone.2015.01.011
- Barba-Recreo P, Del Castillo Pardo De Vera JL, Georgiev-Hristov T, Ruiz Bravo-Burguillos E, Abarrategi A, Burgueño M, et al. Adipose-derived stem cells and platelet-rich plasma for preventive treatment of bisphosphonate-related osteonecrosis of the jaw in a murine model. J Craniomaxillofac Surg 2015;43(7):1161-8. DOI: 10.1016/j.jcms.2015.04.026
- 16. Park S, Kanayama K, Kaur K, Tseng HCH, Banankhah S, Quje DT, et al. Osteonecrosis of the jaw developed in mice: Disease variants regulated by $\gamma\delta$ T cells in oral mucosal barrier immunity. J Biol Chem 2015;290(28):17349-66. DOI: 10.1074/jbc. M115.652305
- Bouxsein ML, Boyd SK, Christiansen BA, Guldberg RE, Jepsen KJ, Müller R. Guidelines for assessment of bone microstructure in rodents using micro-computed tomography. J Bone Miner Res 2010;25(7):1468-86. DOI: 10.1002/jbmr.141
- 18. Park S, Kanayama K, Kaur K, Tseng HCH, Banankhah S, Quje DT, et al. Osteonecrosis of the jaw developed in mice: Disease variants regulated by $\gamma\delta$ T cells in oral mucosal barrier immunity. J Biol Chem 2015;290(28):17349-66. doi: 10.1074/jbc. M115.652305
- Kim S, Williams DW, Lee C, Kim T, Arai A, Shi S, et al. IL-36 Induces Bisphosphonate-Related Osteonecrosis of the Jaw-Like Lesions in Mice by Inhibiting TGF-β-Mediated Collagen Expression.
 J Bone Miner Res 2017;32(2):309-18. DOI: 10.1002/jbmr.2985
- Kikuiri T, Kim I, Yamaza T, Akiyama K, Zhang Q, Li Y, et al. Cell-based immunotherapy with mesenchymal stem cells cures bisphosphonate-related osteonecrosis of the jaw-like disease in mice. J Bone Miner Res 2010;25(7):1668-79. DOI: 10.1002/jbmr.37
- Zhao Y, Wang L, Liu Y, Akiyama K, Chen C, Atsuta I, et al. Technetium-99 conjugated with methylene diphosphonate ameliorates ovariectomy-induced osteoporotic phenotype without causing osteonecrosis in the jaw. Calcif Tissue Int 2012;91(6):400-8. DOI: 10.1007/s00223-012-9649-7
- Cline-Smith A, Axelbaum A, Shashkova E, Chakraborty M, Sanford J, Panesar P, et al. Ovariectomy Activates Chronic Low-Grade Inflammation Mediated by Memory T Cells, Which Promotes Osteoporosis in Mice. J Bone Miner Res 2020;35(6):1174-87. DOI: 10.1002/jbmr.3966
- 23. Yu M, Pal S, Paterson CW, Li JY, Tyagi AM, Adams J, et al. Ovariectomy induces bone loss via microbial-dependent trafficking of intestinal TNF+ T cells and Th17 cells. J Clin Invest 2021;131(4):e143137. DOI: 10.1172/JCI143137

- 24. Jilka RL, Hangoc G, Girasole G, Passeri G, Williams DC, Abrams JS, et al. Increased osteoclast development after estrogen loss: mediation by interleukin-6. Science 1992;257(5066):88-91. DOI: 10.1126/science.1621100
- 25. Kimble RB, Bain S, Pacifici R. The functional block of TNF but not of IL-6 prevents bone loss in ovariectomized mice. J Bone Miner Res 1997;12(6):935-41. DOI: 10.1359/jbmr.1997.12.6.935
- 26. Manolagas SC. Role of cytokines in bone resorption. Bone 1995;17(2 Suppl):63S-67S. DOI: 10.1016/8756-3282(95)00180-l
- Pacifici R. Bone Remodeling and the Microbiome. Cold Spring Harb Perspect Med 2018;8(4):a031203. DOI: 10.1101/cshperspect.a031203
- 28. Hsu E, Pacifici R. From Osteoimmunology to Osteomicrobiology: How the Microbiota and the Immune System Regulate Bone.

- Calcif Tissue Int 2018;102(5):512-21. DOI: 10.1007/s00223-017-0321-0
- 29. Lyu Z, Hu Y, Guo Y, Liu D. Modulation of bone remodeling by the gut microbiota: a new therapy for osteoporosis. Bone Res 2023;11(1):31. DOI: 10.1038/s41413-023-00264-x
- Wang Y, Zhang X, Tang G, Deng P, Qin Y, Han J, et al. The causal relationship between gut microbiota and bone mineral density: a Mendelian randomization study. Front Microbiol 2023;14:1268935. DOI: 10.3389/fmicb.2023.1268935
- 31. Yu M, Pal S, Paterson CW, Li JY, Tyagi AM, Adams J, et al. Ovariectomy induces bone loss via microbial-dependent trafficking of intestinal TNF+ T cells and Th17 cells. J Clin Invest 2021;131(4):e143137. DOI: 10.1172/JCI143137